Cysticercosis and the immunossupression: what are the mechanisms involved?

Cisticercose e a imunossupressão: quais os mecanismos envolvidos?

Ednéia Casagrande Bueno

PhD; Professor of Immunology and Clinical Immunology, Pharmacy Graduation Program and Biomedical Graduation Program, Universidade do Vale do Itajaí (UNIVALI), Itajaí SC, Brazil.

Correspondence:

Ednéia Casagrande Bueno Rua Uruguai 458 / bloco 17 / sala 309 88302-202 Itajaí SC - Brasil E-mail: ecbueno@univalli.br

Conflict of interest:

There is no conflict of interest to declare.

Received 09 January 2012 Accepted 16 January 2012 he human neurocysticercosis (NC) is caused by the presence of the larval form of *Taenia solium* in the central nervous system, after the consumption of water or food contaminated with parasite eggs. The prevalence of NC is related to socioeconomic and cultural factors, representing an important public health problem in countries with deficient sanitary conditions, and in industrialized countries receiving immigrants from epidemic areas. The disease is one of the most severe parasite infections affecting the central nervous system, with complex biological parasite-host interactions due to the occurrence of different parasite antigens in different stages of evolution, and individual genetic variations interfering with the host response, impairing the understanding of the dynamics of parasite survival and host defense mechanisms¹.

The diagnosis of NC is based on clinical, epidemiological, and laboratorial criteria using both imaging and laboratory analyses of cerebrospinal fluid (CSF) samples, including cytomorphological, biochemical, and immunological examinations². The detection of antibodies in serum is another important marker for the NC diagnosis³.

There are only few studies available regarding aspects of the cellular immune response, most of them showing immunosuppression in humans⁴, pigs⁵, and in experimental mouse models⁶⁻⁹. A study from our group using a lymphoproliferation assay with *T. solium* and *T. crassiceps* antigens as stimuli showed an antigen-specific suppression in NC-patients, suggesting that the antigenic components play a suppressor role in the host immune response¹⁰.

Correa et al.⁵ observed a decrease in the CD4/CD8 ratio in these patients, and they have suggested that CD8 cells are involved in immunosuppression. We also identified these cells as predominant in peripheral blood of patients with NC presenting degenerating cysts¹¹, with higher expression of activation cell marker (CD69) during this inflammatory form¹². The CSF samples from inflammatory NC-patients also showed a predominance of CD8 cells and a higher expression of HCAM and ICAM adhesion molecules. CD19 and CD56 cells, as well anticysticercus antibodies, were observed in the CSF samples from both inflammatory and non-inflammatory NC-patients, with high CD69 expression¹².

The immunosuppression observed in experimental cysticercosis seems to be related to the cytokine profile $^{8,13-15}$. Our contribution in this field showed that by using T. solium antigen as stimuli, there is a predominance of Th1 response in inflammatory NC-patients and a mixed Th1/Th2 pattern in noninflammatory NC-patients 12 .

Some experimental cysticercosis studies have attempted to determine the causes of immunosuppression: release of antigenic products¹⁶ that form circulating immunocomplexes¹⁷, suppressing lymphocyte cytokine production or inducing chromosome instability¹⁸; increase and/or decrease in CD8 cells depending on the evolution phase of the disease¹⁹; genetic instability and chromosome alterations in circulating lymphocytes induced by infection^{20,21}; macrophages with activity involving the programmed death ligand 1 pathway⁷; and Th1 and/or Th2 cytokines^{6,8,9,13-15,22,23}. Recently, it was reported the Toll-like receptor 2-dependent pathways as involved in the recognition of *T. crassiceps*, with subsequent activation of the innate response and production of the inflammatory cytokines²⁴. However, these studies on the cellular immunological aspects of cysticercosis do not permit a precise conclusion about the alterations in the host immune response, caused by the parasite or its products.

In this Issue of Arquivos de Neuropsiquiatria, Camargo and Bertolucci²⁵, from Federal University of São Paulo, report a study that investigated the correlation between neuronal death and NC, quantifying the soluble FAS apoptotic factor in CSF from 36 NC-patients by the immunoenzymatic assay. The authors showed higher CSF of soluble FAS protein at NC-patients with active and calcified form. The result represents a contribution to the field here pointed, since the presence of this pro-apoptotic protein could also suggest the involvement of apoptosis in the suppression observed in the NC, indicating that more studies in this area will be helpful to elucidate such field.

Also in this Issue, Matos-Silva et al.²⁶, from Federal University of Goiás, report the development of an

experimental model to NC studies involving two mice lineages by using *T. crassiceps* cysticerci. The authors observed in this original study that C57BL/6 mice showed greater capability on provoking early necrosis in the cysticerci with a chronic inflammation pattern, while BALB/c mice showed necrosis on late stage parasites with an acute inflammation pattern. This experimental model may become a reproducible method for human NC studies, since as pointed by the authors, the *T. crassiceps* cysticerci caused a dynamic inflammatory response from the host and present other advantages such as its rapid development cycle, facilities in maintenance and antigenic similarities to *T. solium* cysticerci.

References

- Schantz PM, Moore AC, Munoz JL, et al. Neurocysticercosis in an Orthodox Jewish community in New York City. N Engl J Med 1992;327:692-695.
- Colli BO, Martelli N, Assirati Junior JA, et al. Cysticercosis of the central nervous system. I. Surgical treatment of cerebral cysticercosis: a 23 years experience in the Hospital das Clinicas of Ribeirao Preto Medical School. Arq Neuropsiquiatr 1994;52:166-186.
- Bragazza LM, Vaz AJ, Passos AD, et al. Frequency of serum anticysticercus antibodies in the population of a rural Brazilian community (Cassia dos Coqueiros, SP) determined by ELISA and immunoblotting using *Taenia crassiceps* antigens. Rev Inst Med Trop Sao Paulo 2002;44:7-12.
- Molinari JL, Tato P, Reynoso OA, et al. Depressive effect of a Taenia solium cysticercus factor on cultured human lymphocytes stimulated with phytohaemagglutinin. Ann Trop Med Parasitol 1990;84:205-208.
- Correa D, Tovar A, Espinoza B, et al. Cisticercosis humana: relación inmunologica hosped-parasito. In: Flisser A, Malagón F (Eds). Cisticercosis humna y porcina: su conocimiento e investigación en México. México: Editorial Limusa; 1989.
- Terrazas LI, Bojalil R, Govezensky T, et al. Shift from an early protective Th1-type immune response to a late permissive Th2-type response in murine cysticercosis (*Taenia crassiceps*). J Parasitol 1998;84:74-81.
- Terrazas LI, Montero D, Terrazas CA, et al. Role of the programmed Death-1 pathway in the suppressive activity of alternatively activated macrophages in experimental cysticercosis. Int J Parasitol 2005;35:1349-1358.
- Toenjes SA, Spolski RJ, Mooney KA, et al. The systemic immune response of BALB/c mice infected with larval *Taenia crassiceps* is a mixed Th1/Th2-type response. Parasitol 1999;118:623-633.
- Villa OF, Kuhn RE. Mice infected with the larvae of *Taenia crassiceps*exhibit a Th2-like immune response with concomitant anergy
 and downregulation of Th1-associated phenomena. Parasitol
 1996;112:561-570.
- Bueno EC, Vaz AJ, Machado LR et al. Antigen-specific suppression of cultured lymphocytes from patients with neurocysticercosis. Clin Exp Immunol 2001;126:304-310.
- Bueno EC, Vaz AJ, Oliveira CA, et al. Analysis of cells in cerebrospinal fluid from patients with neurocysticercosis by means of flow cytometry. Cytometry 1999;38:106-110.
- Bueno EC, Machado LR, Livramento JA, et al. Cellular immune response of patients with neurocysticercosis (inflammatory and noninflammatory phases). Acta Trop 2004;91:205-213.

- Robinson P, Atmar RL, Lewis DE, et al. Granuloma cytokines in murine cysticercosis. Infect Immun 1997;65:2925-2931.
- Terrazas LI, Cruz M, Rodríguez-Sosa M, et al. Th1-type cytokines improve resistance to murine cysticercosis caused by *Taenia* crassiceps. Parasitol Res 1999;85:135-141.
- Toenjes SA, Kuhn RE. The initial immune response during experimental cysticercosis is of the mixed Th1/Th2 type. Parasitol Res 2003;89: 407-413
- Tato P, Castro AM, Rodríguez D, et al. Supression of murine lymphocyte proliferation induced by small RNA purified from the *Taenia solium* metacestode. Parasitol Res 1995;81:181-187.
- Lentoja T, Hammerberg C, Schurig G. Evaluation of spleen lymphocyte responsiveness to a T-cell mitogen during early infection with larval Taenia taeniaeformis. Parasitol Res 1987;73:265-270.
- Arechavaleta F, Molinari JL, Tato P. A Taenia solium metacestode factor nonspecifically inhibits cytokine production. Parasitol Res 1998:84:117-122.
- Meeusen E, Gorrell MD, Rickard MD, et al. Lymphocyte subpopulations of sheep in protective immunity to *Taenia hydatigena*. Parasite Immunol 1989;11:169-181.
- 20. Flisser A, Gonzalez D, Plancarte A, et al. Praziquantel treatment of brain and muscle porcine *Taenia solium* cysticercosis. 2. Immunological and cytogenetic studies. Parasitol Res 1990;76:640-642.
- Herrera LA, Santiago P, Rojas G, et al. Immune response impairment, genotoxicity and morphological transformation induced by *Taenia* solium metacestode. Mutation Res 1994;305:223-228.
- 22. Sciutto E, Fragoso G, Baca M, et al. Depressed T-cell proliferation associated with susceptibility to experimental *Taenia crassiceps* infection. Infect Immun 1995;63:2277-2281.
- Patil S, Robinson P, Actor JK, et al. Proinflammatory cytokines in granulomas associated with murine cysticercosis are not the cause of seizures. J Parasitol 2006;92:738-741.
- Reyes JL, González MI, Ledesma-Soto Y, et al. TLR2 mediates immunity to experimental cysticercosis. Int J Biol Sci 2011;7:1323-1333.
- Camargo JA, Bertolucci PHF. Quantification of Fas protein in CSF of patients with neurocysticercosis. Arq Neuropsiquiatr 2012;70: 264-268.
- Matos-Silva H, Reciputti BP, De Paula EC, et al. Experimental encephalitis caused by *Taenia crassiceps* cysticerci in mice. Arq Neuro-Psiquiatr 2012;70:302-308.