Environmental Sciences

Vol.60: e17160359, January-December 2017 35http://dx.doi.org/10.1590/1678-4324-2017160359 ISSN 1678-4324 Online Edition

BRAZILIAN ARCHIVES OF BIOLOGY AND TECHNOLOGY

AN INTERNATIONAL JOURNAL

Effects of *Yucca schidigera* on gas mitigation in livestock production: A review

Deng-Sheng Sun¹⁺, Xiao Jin¹⁺, Binlin Shi^{1*}, Yuanqing Xu¹, Sumei Yan¹.

¹ Inner Mongolia Agricultural University - College of Animal Science - Huhhot, Inner Mongolia, China.

ABSTRACT

Yucca schidigera extract (YSE) has received much interest in the application of manure deodorization and hazardous gas mitigation in livestock rearing conditions. The main objective of this review article was to summarize the current knowledge regarding YSE towards its gas mitigation from livestock excrement. Saponins have been considered to be vital components of YSE in odor control and gas reduction in intensive farming industry due to their potentials in lowering methane for ruminants and ammonia for monogastric animals. This review article mainly covered the studies in ruminants, especially focused on in vitro environment. It also summarized possible reasons of the conflicting results among studies from the perspective of experimental design such as incubation time (in vitro) or storage time of manure, and some other factors such as feed source and dietary composition. In addition to traditional dietary inclusion of YSE alone, recent studies prone to apply YSE in new ways such as combining it with other natural compounds or using it to treat manure directly. Until now, there are still controversies in terms of the effectiveness of YSE in intensive-farming environment among researches, therefore further deeper studies on the expression of YSE bioactivity are needed, especially on the molecular level.

Key words: Yucca schidigera, livestock, ammonia, methane, ruminal fermentation

_

Author for correspondence: shibinlin@yeah.net

⁺ These authors contributed to the work equally and should be regarded as co-first authors.

INTRODUCTION

Increasing attention has been placed on greenhouse gas (GHG) emissions in recent years with deteriorating scenario of global warming nowadays ^{1,2}. Appropriately 7-18% of the global GHG emissions are originated from livestock sector ³. Each year domestic livestock can generate 80-115 million tons of methane (CH₄) on a global basis, which is equal to 15-20% of total anthropogenic methane ⁴. As another gas generated from livestock, ammonia (NH₃) is harmful to environment as well ^{5,6}, although not labeled as GHG ⁷. A statistics showed that agriculture industry took dominant part in ammonia emissions ⁸, which the main results were coming from barns (34-43%) and livestock waste storage (22-26%) ⁹.

Several measures have been undertaken to tackle environmental challenges in animal husbandry operations ¹⁰, and it was found that the additives such as fenugreek and cinnamomum verum ¹¹ in diets is one of the effective ways to mitigate GHG emissions and ammonia concentrations in livestock rearing process. Compared with chemical feed additives such as antibiotics and ionophores which were considered to be not suitable in the application of feed additives ¹², plant-derived substances have more integrated benefits ¹³: they can be incorporated as growth promoters (probiotics and prebiotics) ¹⁴, or show anti-inflammatory, anti-fungal, anti-infectious or antitoxigenic potentials in livestock production ^{15,16}. Therefore, phytogenic feed additives are the renewed hotspots of researchers who were aiming at finding their potential values in improving rearing conditions of farming animals in addition to aforementioned benefits ¹⁷⁻¹⁹. Saponins contained in plants, when labeled as feed additives, are one group of phytochemicals that have been studied continuously. This article exclusively discussed more recent studies of a typical saponin-containing plant - *Yucca schidigera*, and its effects on environmental control in livestock, and attempt to summarize opinions from latest studies in its application.

Yucca schidigera

Yucca schidigera (YS), also named as yucca, is a member of Agavaceae family. The potential of YS has been valued since it was used to treat inflammatory illnesses effectively ²⁰. Being a tropical plant, YS originally grows in North America, especially in arid Mexican dessert ²¹. Beneficial effects of Yucca schidigera extracts (YSE) are covering many aspects such as producing desired nutritional attribute that improving feed conversion efficiency thus enhancing animal growth, contributing to environmental control in commercial rearing conditions, and participating in microbial activity modification (e.g., anti-protozoal activity) ^{21,22}.

As a rich source of phytochemicals with promising bioactive functions ²³, YS has several components such as steroidal saponins, polyphenolics (e.g., resveratrol and some other stilbenes including yuccaols A, B, C, D and E) ^{20,24,25}. With 10% of steroidal saponins in its stem dry matter ²⁶, YS has been perceived as one of the two major commercial saponin sources, the other one is *Quillaia saponaria* ²¹. Saponins have been considered to be vital components of YS in odor control in intensive farming industry ^{19,27}. The 3-dimensional spatial orientation, its lipophilic aglycon, and the sugar composition all together contribute to the biological properties of YS ²⁸. In addition, new steroidal saponins with different structures included in YS have been detected continuously ²⁹⁻³¹. Future analysis of the YS molecule structures, isolation of YS bioactive components, and ascertaining its purity will provide more evidence for YSE application in terms of ameliorating the environmental pollution from livestock industry, and increase the feed efficiency in diets at the same time.

EFFECTS OF Yucca schidigera ON GAS MITIGATION IN LIVESTOCK

In Ruminants

Studies of YSE application in gas mitigation area have mostly been focused on ruminants, especially in cattle and sheep (Table 1). Emitting gases were mainly measured in cattle-based experiments. Singer et al. 32 reported that with increased feeding of YSE to lactating dairy cows, 4 h and 24 h gas production generated through these collected rumen fluids were increased, exhibiting a strong linear effect (P<0.05). A similar result was observed in another in vitro experiment which involved different ruminal substrates including soluble potato starch, cornstarch, or hay plus concentrate (1.5:1) in the incubation process ³³. Total gas productions at 6 h and 24 h were increased as dietary sarsaponin increased from 1.2 to 3.2 g/L, and the methane reduction rate was statistically up to different substrates. Methane production was decreased (P<0.05) by YSE addition in both gas production rate (mL/min) and extent (L) in the study of Pen et al. ³⁴. In another research, methane production at 24 h was decreased (P<0.05) by 110 g/kg of YSE addition, although in vitro gas production was not affected ²⁷. Holtshausen et al. ³⁵ indicated that in order to avoid the potential side effects of YSE on ruminal fermentation and feed digestion, saponin levels were reduced (10 g/kg of DM) that resulted in a nonsignificant difference of methane production in vitro among different treatments. However, when sarsaponin concentration was 1% of DM (22.4 g), YSE addition in diets resulted gas reduction in steers effectively, in which methane was inhibited by approximately 12.7% (P<0.05) from day 6 to day 9 of the 10 days feeding period without impairing animal performance ³⁶.

There are also some studies showed inconsistent results. YSE supplementation of 3 g/kg of DM did not reduce methane production in lactating dairy cows, as suggested by Zijderveld et al. ³⁷. Similar results were also observed by Li and Powers ³⁸ who measured gaseous emissions in room exhaust air of steers. In their study even the 1.5% YSE inclusion groups failed to alter either methane, or ammonia, or nitrous oxide emissions on a daily basis (per unit DMI). Methanol extract of YSE was used in an experiment in vitro, and YSE decreased (P<0.05) methane production when calculated by per unit of dry matter, but not by per unit of true digested dry matter ³⁹. Most researches using sheep as experimental animals were conducted to measure ruminal fermentation parameters related to gas production such as ruminal ammonia concentration. The results of an experiment in vitro showed that 100 mg/kg dietary sarsaponin of DM (600 mg/kg CP) reduced the ruminal ammonia over 21% throughout the measurements from day 5 to day 10 40. In the subsequent study in vivo, only 2 and 30 mg/kg of DM YSE were added in the diets 41. The results showed that dietary YSE only had slight trends to reduced gas emission without statistical effect over a 15 days period 41. Feeding the diets with 120 ppm YSE in sheep resulted that YSE reduced N losses in urine and total N losses, leading to a 50% higher retained N, and ammonia N concentration was lowered by 11.9% although not significant ⁴². In the subsequent experiment ⁴³, the supplementation of YSE in the basal diet was 240 ppm DM per day and dietary YSE feeding lasted 14 days in which it comprised of 8 days of dietary adjustment. Compared to the control diet, YSE reduced rumen ammonia N concentrations (P < 0.05) in Cheviot wethers ⁴³. A decline of rumen ammonia N was explained that caused by dietary YSE 44.

At 4 h and 6 h after feeding YSE-containing diets (300 mg/kg) in sheep, propionate concentration was increased and acetic concentration was lowered, but neither of them changed significantly ⁴⁵. At 2 h after feeding YSE-containing diets (300 mg/kg), protozoan population was decreased (*P*<0.05). And all 100, 200, and 300

mg/kg YSE feeding resulted a increasing in ammonia concentration 45 . When supplementing 170 mg/d YSE with two other additives (flavomycin and ropadiar) in sheep diets, rumen liquor samples were taken on the day 9 and day 11, and gas production from the sheep was measured from day 12 to day 14. Results indicated an increase in VFA concentration and a decrease in acetate:propionate ratio (P<0.05), while ammonia N concentration (P<0.05) and average methane production (P<0.05) were reduced compared to the control 46 . A later experiment *in vivo* reported that ruminal ammonia concentration, ammonia N concentration and protozoa population in sheep were suppressed especially by the 200 and 300 mg/kg YSE treatment groups in the experimental conditions where dietary YSE levels were 100, 200, 300 mg/kg 47 .

For methane production, a recent study revealed that methane production was not affected (P>0.05) by YSE-contained diets, even at highest levels (6 g/d saponins) ⁴⁸. Nonetheless, in a latest research, YSE reduced methane significantly (P<0.05) in a dose dependent manner, in both substrates (dates byproducts and the vetch-oat) used in the trials ⁴⁹. When saponin levels were over 8 mg/mL, the decreasing percentages of methane can be as high as 60% ⁴⁹. Decreased methane production (11%) in wether sheep by YSE (14 mL) in two 23 h runs (day 16 to day 17 of the 18 days period) was noted as well ³⁴.

In Monogastric Animals

This section summarized the studies of two typical monogastric animals: swine and poultry. A number of studies have been carried out to determine the effects of YSE on reducing ammonia in poultry farms. Cabuk et al. ⁵⁰ reported that feeding of 120 mg/kg dietary YSE resulted in a decreased ammonia concentration of broiler houses at day 19 without impairing broiler performance. However, in another experiment, the supplementation of 100 ppm of YSE and Quillaja saponaria was added in a corn-soybean control diet, and ammonia emission of broiler chicken litters was not altered compared with control in the 42 days experimental period ⁵¹. When YSE was applied to laying-hens, 100 ppm inclusion in diets significantly reduced ammonia emission by 44% and 28% for the first two days of manure storage ⁵². However, an experiment showed that ammonia N concentrations and microorganism levels of litter materials (half was wood shavings, the other was rice hull) among examined groups did not show statistical difference when pulverized YSE was applied to different litter materials at the level of 0, 4% and $8\%^{53}$. It was hypothesized that the efficiency of YSE could be amplified if litter was used in farming houses under bad situations 53. As a study to evaluate the effects of YSE on poultry manure alone or together with microbial preparation, YSE showed highest potentials in reducing volatile odorous compounds concentrations after 96 h of the process ¹⁹. This study also confirmed the ability of YSE to decrease the concentrations of odorous compounds emitted from poultry manure such as ammonia, trimethylamine, dimethylamine, isobutyric acid and hydrogen sulfide. In addition, applying YSE separately with microbial preparation at 48 h interval obtained best results ¹⁹ Only few studies have been carried out using swine as experimental animals. Panetta et al. 54 observed no significant effect of dietary YSE (0, 62.5, 125 mg/kg) on ammonia emission during 72 h of consecutive measurement after 4 days dietary adjustment. A decreasing tendency (P>0.05) in ammonia gas production of fecal samples was shown during a 30 days experiment period ⁵⁵. However, Liang et al. ⁵⁶

indicated that YSE added in the feed (125 mg/kg) decreased the emission of

Table 1 – Studies oF the effects of YSE on ammonia and methane mitigation in ruminants

ammonia and hydrogen disulfide in the 35 days trials.

Reference	e Animal	s YSE Levels	Results
27	Dairy Cows vitro)	(in0 or 110 g/kg	CH_4 production at 24 h was reduced (P <0.05).
32	Dairy Cows vitro)	(in0, 5, 10 or 15 g cow/d (Sarsaponin)	Rumen NH ₃ -N levels tended (<i>P</i> =0.06) to decrease with increased YSE level.
33	Dairy Cows vitro)	(in0, 1.2, 1.8, 2.4, 3.2 g/L (Sarsaponin)	Fermentation of soluble potato starch $(P<0.05)$, cornstarch $(P<0.05)$, or hay plus concentrate $(P<0.05)$ decreased CH_4 production with the concentration of sarsaponin increased.
34	Dairy Cows vitro)	(in (80-100 g/kg saponin)	Rate and extent of CH_4 production were reduced (P <0.001) by YSE addition in a dose-dependent manner by up to 42% and up to 32%.
39	Cattle <i>vitro</i>)	(in650 μg/ml (100 μg/mL steroida sapogenin).	alYSE treatments decreased CH_4 production when measured as per unit of DM (P <0.05).
40	Cow vitro)	(in1, 20 and 100 mg/kg DN (sarsaponin)	Substantive effects of the saponin-rich products on ruminal nitrogen metabolism (CH ₄ , ruminal NH ₃ level) were observed only at doses exceeding those recommended by the manufacturers.
35	Dairy Cows vitro and vivo)	(inin vitro: 15, 30, 45 g/kg DM; in vivo d in10 g/kg of DM (6.0% saponin)	Methane production <i>in vitro</i> was lowered c:(<i>P</i> <0.05), yet methane <i>in vivo</i> was not affected either at low or high supplementations.
36	Steers	0, 11.2, 22.4 g (sarsaponin)	Approximately 12.7% of CH_4 was inhibited (P <0.05).
37	Dairy Cows	3 g/kg DM	CH ₄ production was not affected.
38	Steers	Experiment 1: 0.64% YSE (8.5% saponin); Experiment 3: 1.5% YSE of DM	CH ₄ , NH ₃ , N ₂ O monitored in room exhaust air were not affected.
49	Sheep vitro)	(in0, 2, 4, 6, 8 mg/mL (saponins: 44 g/kg DM)	CH ₄ production was reduced (<i>P</i> <0.05).
41	Lambs	2 and 30 mg/kg DM (sarsaponin)	CH ₄ production was not affected (<i>P</i> >0.05).
42	Lambs	120 ppm	NH_3 -N was not affected ($P>0.05$).
43	Lambs	240 ppm	Ruminal NH ₃ concentration was decreased $(P<0.05)$, urinary N was lowered $(P<0.01)$.
44	Lambs	14 mL (1.31-1.64 g of saponing/wether/d)	
45	Lambs	100, 200, 300 mg/kg	Ruminal NH ₃ concentration was decreased $(P<0.05)$.
46	Sheep	170 mg/d	NH_3 -N concentrations was lowered (P <0.05).
		<u> </u>	

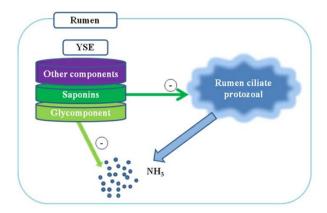
Sun, DS et al.

47	Sheep	0, 100, 200, 300 mg/kg	The 200 and 300 mg/kg YSE groups have a suppressing effect on ruminal NH ₃ concentration (<i>P</i> <0.05) than control.
48	Sheep	0, 1.5, 3.0, 4.5, 6.0 g/d of saponins	CH ₄ emissions were not affected (<i>P</i> >0.05).

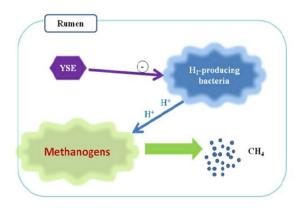
Gas Mitigation Mechanisms

Gas Mitigation Mechanisms of Ruminants

Based on the research of Headon et al. ²⁴, the two components of *Yucca schidigera*, the glycocomponent and the saponin fraction, act differently in binding ammonia in rumen (Fig. 1). The glycocomponent has an ability to bind ammonia directly, while saponin fraction may inhibit ammonia concentrations by membranolytic properties through altering rumen ciliate protozoa, as it (saponin fraction) can cause cell lysis by acting with cholesterol in membranes of protozoal cell ²¹. However, the indirect way to reduce ammonia concentration through saponin may contributes most to ammonia reduction since the suppressing potential of glycocomponent is limited ⁵⁷.



As it is shown by Figure. 2, the most convincing mechanism for methane suppressing effects of steroidal saponins containing plants, YSE specifically, is that methane is possibly reduced through an inhibition of the growth of H₂-producing bacteria ^{57,58}. It has been demonstrated that ciliate protozoa, which provides substrate (H₂) for methanogens ^{59,60}, is associated with 9-25% of ruminal methanogens ⁶¹. Reduced methane emission due to saponin addition is regarded as the result of its toxicities towards protozoa population ^{35,62}. The symbiotic relationship between methanogens and protozoa in the ruminal environment accounts at least partially for decrease in methane production due to YSE inclusion in diets ³⁷. When YSE is added in diets, the balance between methanogens and protozoa would be broken which will lead to methanogens reduction, and eventually influence the production of the emitting methane ³⁶.



Given that the equation (Methane = $(1.8 \times \text{acetate} - 1.1 \times \text{propionate} + 1.6 \times$ butyrate) / 4) presented by Moss et al. ⁶³, the improvement of propionate production can lead to the reduction of methane production in rumen ³⁴. This is also supported by Cieslak et al. 64 who reported that the propionate production contents with methane for available hydrogen. S. ruminantium, the most predominant bacterium in the process of succinate decarboxylation, accounts for the majority of propionate yield in the rumen ^{65,66}. Narvaez et al. ³⁹ further noticed that *S. ruminantium* were significantly increased with YSE supplementation, indicating a positive transaction of microbial population towards those propionate-producing bacteria. However, it is possibly that the interaction between protozoa and methanogen has been overestimated ²⁷, as the protozoa viability at specific time in the experiment had not been identified ³⁴. Goel et al. ⁶⁷ provided that there was no connection between methanogens, protozoal population and methane production when using different saponin-rich materials such as Carduus and Sesbania to conduct the study in vitro. According to Lila et al. 33, YSE addition can only decrease protozoal populations at 6 h of fermentation in in vitro batch cultures since samples collected at 24 h had no detectable protozoa. It seems that YSE has a short lived effect on protozoa in vitro, which gives us a partly explanation about reduction of methane with YSE addition. The pH value is another factor that has an impact on YSE function on the reduction of methane production in rumen. In general, reduction of methane production entails an alteration in total VFA concentration ⁶⁴ which relates strongly with the acetate/propionate ratio, and this ratio is dependent partly on pH ⁴⁹. Cardozo et al. ⁶⁸ observed that YSE increased the proportion of propionate at pH 5.5, but not at pH 7.0 in in vitro rumen environment. CO₂, an end product of lactate fermentation to propionate ⁶⁹, may contribute to the increasing of total gas production ³³. It is suggested that with the increasing of sarsaponin levels in rumen, CO2 would be generated through succinate:propionate pathway 33.

Lila et al. ³⁶ pointed out that the lowering of ruminal ammonia concentration in response to YSE to a less extent could be attributed to an inhibited deaminative activity. Gram-positive bacteria and protozoa may be inhibited due to the sarsaponin inclusion which resulted in the falling of ruminal ammonia concentration ³³. Although both experiments *in vitro* and *in vivo* have demonstrated the decreasing of ammonia ^{58,70,71}, the effects could only be observed when at higher application rates of YSE ⁷². When at low application rates, YSE processed little biological effects on ruminal ammonia utilization ⁷³. When high levels of YSE (i.e., > 5 mg/mL) ³² were added in diets, protease activity may be increased, while deaminase activity was unchanged to avoid dietary protein degrade to ammonia ⁵⁸. Rumen ammonia N levels tended to reduce with the increasing levels of YSE at high application rates ^{32,74}. However, it remains a question about the mechanisms of ammonia reduction in response to YSE when short incubation time was incorporated in the experiment ⁷³.

Sun. DS et al.

Gas Mitigation Mechanisms of Monogastric animals

According to Liang et al. ⁵⁶, urease activity might be inhibited efficiently with YSE inclusion, which would decrease the speed of ammonia N formation from urea, so the increasing trends of ammonia N concentrations would be reduced. The dynamic balance of N would be broken in this moderate manner, hence ammonia emission rate would be lowered down. As for hydrogen disulfide reduction, it is hypothesized that YSE may decompose the generation of dissolvable sulfide by inhibiting sulfate reducing bacteria or involves in the process where sulfate reductase participate. It is speculated that antimicrobial abilities of saponins may also accounted for the high efficiency of YSE in dealing with odor from poultry feces ¹⁹. The positive effect of YSE on ammonia reduction may also due to the readily volatilized ability of urinary ammonium which is part of ammonia emitted from manure ⁵⁴.

Uric acid also has a positive effect on ammonia volatilization ⁷⁵. Moisture concentration in manure, which can be changed by YSE ⁵², is linked to the transformation of decomposition of uric acid directly ⁷⁶. But these conversions (solid urea dissolution and urea hydrolysis) need to be finished prior to ammonia emission ⁷⁷. Higher pH levels (above 7.0) which can be observed with YSE inclusion ⁵², is favorable for ammonia release since ammonia is a major form of gas emitted under this condition ⁷⁸⁻⁸⁰. Factors such as different collection time correspond with varying degrees of manure moisture ⁵², which need to be noticed to minimize the inaccuracy of experiment as well. Onbasilar et al. ⁵³ attributed the lack of effect in the experiment to exactly relatively low moisture and pH levels.

Possible Reasons of Conflicting Results

Numerous researches have been carried out in an attempt to find the convincing benefits of YSE with regard to gas mitigation, but the results of different studies are contradictory which to some extent were attributed to experimental designs and some non-YSE-structure factors. This part summarized possible factors from the past studies that may have impacts on the accuracy of experimental results, providing basis for the designing of more comparable experiments in the subsequent studies. Feed Source and Dietary Composition

Feed source and dietary composition can not be ignored when the effects of YSE on gas mitigation are being investigated. An experiment in vitro showed that methane production from ruminal fluid was decreased differently with the increasing of sarsaponin concentration when using soluble potato starch, cornstarch, or hay plus concentrate (1.5:1) as substrates, either at 6 h and 24 h ³³. However, Hristov et al. ⁷³ did not observe significant effects of YSE on ruminal fermentation after 8 h incubations in vitro. But substituting 50% corn by barley grain in feed mixture (alfalfa hay, grain, soybean meal) had a positive effect on utilization of ammonia N in vitro 73, suggesting that changing of specific components in the diets might be beneficial to the digestion of animals, which may make it easier for YSE to show its potential. Singer et al. ³² indicated that 4 h gas production of YSE-modified rumen fluid was affected by different levels of starch in feedstuffs. It is hypothesized that fiber digesting bacteria was reduced with increasing levels of YSE, which would improve amylolytic bacterial population and/or activity, leading to higher yet different gas production among different feedstuffs, thus offered us the notion that the composition of the diet impacted the effects of YSE on gas mitigation.

Xu et al. ²⁷ observed no interaction between YSE dosage either with forage source or the ratio of forage:concentrate in diets. Nonetheless, decreased YSE dosages were accompanied with increasing dietary concentrate values ⁸¹⁻⁸³. So a relatively high concentrate-contained ration is recommended in order to obtain better effects of YSE on methane reduction. Propionate concentration, which is related to methane

production ⁶³, was increased by YSE addition in ruminal fluid *in vitro* ³⁵. This trend would be more obvious for a high-grain diet than for high-forage diets ^{74,84,85}. In addition, both methane production ^{86,87} and ammonia emission ⁸⁸ in rumen are vulnerable to dietary protein levels. Reduced nitrogen emission might be larger with more pronounced protein deficit (>10%) in diets than diets with adequate protein level ⁴¹. Therefore, maintaining the proper nutritional levels in diets is one of the prerequisites in expressing the potential benefits of YSE with regard to gas control. Incubation Time (*in vitro*) or Storage Time (Manure)

It is known that fermentation time is essential in fermentation process. Short time fermentation, 4 h in particular, was best chosen for better assessment of gas production, because bacterial population was on the peak value at this time and fermentation parameters would be more reliable ⁸⁹. Wang et al. ⁹⁰ reported that deglycosylation of saponins from YSE occurred at 4 h *in vitro*, which may result in microbial inactivation of the saponins by the formation of sapogenin. While 24 h is mainly for the investigation of feed Metabolic Energy ⁹¹. Singer et al. ³² confirmed the differences of YSE on gas production at different time by monitoring 4 h (P<0.01) and 24 h (P<0.05) gas production *in vitro*. Rumen microbial adaptation induced by high levels of YSE may be responsible for longer incubation such as 24 h ⁷⁰. Wang et al. ⁹² also confirmed the existence of microbial adaptation of YSE during ruminal fermentation. Therefore in order to make it more significant in terms of gas mitigation at longer incubation time, higher levels of YSE feeding may be more desirable.

When dealing with manure, determining gas emissions from feces at different storage time would lead to various results. In an experiment *in vivo* with 22 days storage time, ammonia emission from room exhaust air in steer house was not affected by YSE supplementation, accounting for only 12% of N losses because 33% of N was retained in manure in the form of ammonia N ³⁸. This was not corresponding with other studies that an average 44.3% of N losses was through volatilization, most likely as ammonia ⁹³. Furthermore, during the study period, if the transformation is consecutive, ammonia emissions may be also as a result of organic N to inorganic N ³⁸.

Chemical Composition of YSE Products

Saponins from different sources vary in their contents which would affect bioactivities in rumen fermentation ^{35,94}. In addition, with different extraction methods ⁹⁵, the active components of YSE measured using different methods could be inconsistent. For instance, whole-plant product containing polyphenolics may account for some of the bioactivities of YSE ⁹⁶, which made the effects of YSE on gas mitigation not exclusive to saponins. Furthermore, agronomic and environmental factors like vegetative stage of plant at harvest also influence plant content of YSE ^{23,28,97}. And some contents of YSE was suspected to show negative effects towards livestock even they possess a GRAS (generally regarded as safe) label ⁹⁸, and it has not been reported if this is related to extracting methods of YSE.

Different Expressions of YSE Addition Supplied

Although there are numerous reports about YSE application in gas mitigation process, it remains a challenge to make meaningful comparisons among these studies. Levels of YSE in different researches were expressed in various ways such as on the basis of substrate addition (i.e., g/kg of DM) or volume (i.e., g/L) ³⁵, or as actual saponin concentration, or saponin source concentration, but the purity of the saponins used can attribute to various effects ⁹⁹. These would make it even hard for effective comparisons.

Different Measurement Methods of YSE Concentration and Gas-Producing-Related Parameters

It is different among saponin determining methods (e.g., Wang et al. ⁵⁸: smilagenin equivalents; Holtshausen et al. ³⁵: butanol-extracted solubles). For methanogen determination, it would be not accurate if it is determined through culture-based techniques, because only part of microorganisms would be cultured due to its non-specificity ¹⁰⁰. But when incorporating a marker (purine) or ¹⁵N into the rumen, more integral results of YSE enhancement to microorganisms were obtained ¹⁰¹. In addition, the determination of ammonia concentration and its emissions was processed in different ways, which is probably one of the reasons for discrepancies in observations ¹⁰². The gap between two methane measurements, chamber measurements and SF6 technique, is over 4% since chamber measurements can also monitor methane emission of rectum besides respired and eructated emission ¹⁰³.

CONCLUSIONS

This review primarily covered studies of YSE with regard to its application in gas mitigation and summarized some characteristics related to gas mitigation of cattle and sheep in methane reduction, swine and poultry in ammonia reduction. In this article we also summarized possible factors that may affect the results of studies on YSE. Although the results of some studies are conflicting, many articles reported positive effects of YSE on methane/ammonia mitigation, and most of the studies were carried out *in vitro*. Further finely designed *in vivo* experiments of ruminants, especially in cattle, are recommended. Furthermore, the contradictory results of different studies to some extent are attributed to the experimental designs among various studies. So the unification of gas measurement methods and YSE supplementation would make it easier for the comparisons among different studies.

REFERENCES

- 1. O'Mara FP. The significance of livestock as a contributor to global greenhouse gas emissions today and in the near future. *Anim Feed Sci Tech*. 2011; 166-167: 7-15.
- Bennetzen EH, Smith P, Porter JR. Agricultural production and greenhouse gas emissions from world regions - The major trends over 40 years. *Global Environ Chang*. 2016; 37: 43-55.
- 3. Hristov AN, Oh, Lee J, Meinen C, MontesR, Ott F, et al. Mitigation of greenhouse gas emissions in livestock production A review of technical options for non-CO₂ emissions. In: Gerber JP, Henderson B, Makkar HPS, eds. Rome: Food and Agriculture Organization of the United Nations; 2013.
- 4. Houghton JT, Ding Y, Griggs DJ, Noguer M, Linder PJ. Climate Change 2001: The Scientific Basis: Contribution Of Working Group I To The Third Assessment Report Of The Intergovernmental Panel On Climate Change. Cambridge: Cambridge University Press; 2001.
- 5. Barrancos J, Briz S, Nolasco D, Melián G, Padilla G, Padrón E, et al. A new method for estimating greenhouse gases and ammonia emissions from livestock buildings. *Atmos Environ*. 2013; 74: 10-17.
- 6. Samer M. Emissions inventory of greenhouse gases and ammonia from livestock housing and manure management. *Agric Eng Int: CIGR Journal*. 2013; 15(3): 29-54.
- 7. Charlotte B. International commitment toward curbing global warming: The Kyoto protocol. *Envtl Law*. 1997; 4: 917-942.

- 8. Behera SN, Sharma M, Aneja VP, Balasubramanian R. Ammonia in the atmosphere: a review on emission sources, atmospheric chemistry and deposition on terrestrial bodies. *Environ Sci Pollut R*. 2013; 20(11): 8092-8131.
- 9. Skjøth CA, Geels C. The effect of climate and climate change on ammonia emissions in Europe. *Atmos Chem Phys Discuss*. 2012; 12(9): 23403-23431.
- 10. Hou Y, Velthof GL, Oenema O. Mitigation of ammonia, nitrous oxide and methane emissions from manure management chains: a meta-analysis and integrated assessment. *Global Change Biol*, 2015; 21(3): 1293-1312.
- 11. Lewis KA, Tzilivakis J, Green A, Warner DJ. Potential of feed additives to improve the environmental impact of European livestock farming: a multi-issue analysis. *Inter J Agr Sust.* 2015; 13(1): 55-68.
- 12. Russell JB, Rychlik JL. Factors that alter rumen microbial ecology. *Science*. 2001; 292(5519): 1119-1122.
- 13. Mueller K, Blum NM, Kluge H, Mueller AS. Influence of broccoli extract and various essential oils on performance and expression of xenobiotic- and antioxidant enzymes in broiler chickens. *Brit J Nutr.* 2012; 108(4): 588-602.
- 14. Alloui MN, Agabou A, Alloui N, Application of herbs and phytogenic feed additives in poultry production A Review. *Global J Anim Sci Res.* 2014; 2(3): 234-243.
- 15. Khan SH. The use of green tea (Camellia sinensis) as a phytogenic substance in poultry diets. *Onderstepoort J Vet Res.* 2014; 81(1): 1-8.
- 16. Steiner T, Syed B. Medicinal and aromatic plants of the world. In: Phytogenic Feed Additives in Animal Nutrition. Dordrecht: Springer Netherlands; 2015. p. 403-423.
- 17. Morsy AS, Soltan YA, Sallam SMA, Kreuzer M, Alencar SM, Abdalla AL. Comparison of the *in vitro* efficiency of supplementary bee propolis extracts of different origin in enhancing the ruminal degradability of organic matter and mitigating the formation of methane. *Anim Feed Sci Tech.* 2015; 199: 51-60.
- 18. Prayuwidayati M, Sunarti TC, Sumardi, Subeki, Wiryawan KG. Bioactive compounds isolated from lignin of empty bunch palm fiber and their effects on in vitro rumen fermentation. *Media Peternakan*. 2015; 38(3): 183-191.
- 19. Matusiak K, Oleksy M, Borowski S, Nowak A, Korczynski M, Dobrzanski Z, et al. The use of *Yucca schidigera* and microbial preparation for poultry manure deodorization and hygienization. *J Environ Manage*. 2016; 170: 50-59.
- 20. Cheeke, PR. Actual and potential applications of *Yucca schidigera* and *Quillaja saponaria* saponins in human and animal nutrition. *J Anim Sci.* 2000; 77(E-Suppl): 1-10.
- 21. Cheeke PR, Piacente S, Oleszek W. Anti-inflammatory and anti-arthritic effects of *Yucca schidigera*: A review. J Inflamm. 2006; 3: 6.
- 22. Piacente S, Pizza1 C, Oleszek W. Saponins and phenolics of *Yucca schidigera* Roezl: Chemistry and bioactivity. *Phytochem Rev.* 2005; 4(2-3): 177-190.
- 23. Francis G, Kerem Z, Makkar HP, Becker K. The biological action of saponins in animal systems: a review. *Brit J Nutr.* 2002; 88(6): 587-605.
- 24. Headon DR, Buggle KA, Nelson AB, Killeen GF. Glycofractions of the Yucca plant and their role in ammonia control. In: Lyons TP, editor. Biotechnology in the Feed Industry. Nicholasville, KY: Alltech Technical Publications; 1991. p. 95-108.
- Patel S. Yucca: A medicinally significant genus with manifold therapeutic attributes. Nat Prod Bioprospect. 2012; 2(6): 231-234.
- 26. Kaneda N, Nakanishi H, Staba EJ. Steroidal constituents of *Yucca schidigera* plants and tissue cultures. *Phytochemistry*.1987; 26(5): 1425-1429.
- 27. Xu M, Rinker M, McLeod KR, Harmon DL. *Yucca schidigera* extract decreases *in vitro* methane production in a variety of forages and diets. *Anim Feed Sci Tech.* 2010; 159(1-2): 18-26.
- 28. Sen S, Makkar HPS, Becker K. Alfalfa saponins and their implication in animal nutrition. *J Agric Food Chem.* 1998; 46(1): 131-140.
- 29. Miyakoshi M, Tamura Y, Masuda H, Mizutani K, Tanaka O, Ikeda T, et al. Antiyeast steroidal saponins from *Yucca schidigera* (Mohave Yucca), a new anti-food-deteriorating agent. *J Nat Prod.* 2000; 63(3): 332-338.
- 30. Oleszek W, Sitek M, Stochmal A, Piacente S, Pizza C, Cheeke P. Steroidal saponins of *Yucca schidigera* Roezl. *J Agric Food Chem.* 2001; 49: 4392-4396.

- 31. Kowalczyk M, Pecio Ł, Stochmal A, Oleszek W. Qualitative and quantitative analysis of steroidal saponins in crude extract and bark powder of *Yucca schidigera* Roezl. *J Agric Food Chem.* 2011; 59(15): 8058-8064.
- 32. Singer MD, Robinson PH, Salem AZM, DePeters EJ. Impacts of rumen fluid modified by feeding *Yucca schidigera* to lactating dairy cows on *in vitro* gas production of 11 common dairy feedstuffs, as well as animal performance. *Anim Feed Sci Tech.* 2008; 146(3-4): 242-258.
- 33. Lila ZA, Mohammed N, Kanda S, Kamada T, Itabashi H. Effect of sarsaponin on ruminal fermentation with particular reference to methane production *in vitro*. *J Dairy Sci*. 2003; 86(10): 3330-3336.
- 34. Pen B, Sar C, Mwenya B, Kuwaki K, Morikawa R, Takahashi J. Effects of *Yucca schidigera* and *Quillaja saponaria* extracts on *in vitro* ruminal fermentation and methane emission. *Anim Feed Sci Tech.* 2006; 129(3-4): 175-186.
- 35. Holtshausen L, Chaves AV, Beauchemin KA, McGinn SM, McAllister TA, Odongo NE, et al. Feeding saponin-containing *Yucca schidigera* and *Quillaja saponaria* to decrease enteric methane production in dairy cows¹. *J Dairy Sci*. 2009; 92(6): 2809-2821.
- 36. Lila ZA, Mohammed N, Kanda S, Kurihara M, Itabashi H. Sarsaponin effects on ruminal fermentation and microbes, methane production, digestibility and blood metabolites in steers. *Asian Austral J Anim.* 2005; 18(12): 1746-1751.
- 37. Zijderveld SMV, Dijkstra J, Perdok HB, Newbold JR, Gerrits WJJ. Dietary inclusion of diallyl disulfide, yucca powder, calcium fumarate, an extruded linseed product, or medium-chain fatty acids does not affect methane production in lactating dairy cows. *J Dairy Sci.* 2011; 94(6): 3094-3104.
- 38. Li W, Powers W. Effects of saponin extracts on air emissions from steers. *J Anim Sci*. 2012; 90(11): 4001-4013.
- 39. Narvaez N, Wang YX, McAllister T. Effects of extracts of Humulus lupulus (hops) and *Yucca schidigera* applied alone or in combination with monensin on rumen fermentation and microbial populations *in vitro*. *J Sci Food Agric*. 2013; 93(10): 2517-2522.
- 40. Sliwinski BJ, Soliva CR, Machmuller A, Kreuzer M. Efficacy of plant extracts rich in secondary constituents to modify rumen fermentation. *Anim Feed Sci Tech.* 2002a; 101(1): 101-114.
- 41. Sliwinski BJ, Kreuzer M, Wettstein HR, Machmuller A. Rumen fermentation and nitrogen balance of lambs fed diets containing plant extracts rich in tannins and saponins, and associated emissions of nitrogen and methane. *Arch Anim Nutr.* 2002b; 56(6): 397-392.
- 42. Santoso B, Mwenya B, Sar C, Gamo Y, Kobayashi T, Morikawa R, et al. Effects of supplementing galacto-oligosaccharides, *Yucca schidigera* or nisin on rumen methanogenesis, nitrogen and energy metabolism in sheep. *Livest Prod Sci.* 2004; 91(3): 209-217.
- 43. Santoso B, Mwenya B, Sar C, Takahashi J. Ruminal fermentation and nitrogen metabolism in sheep fed a silage-based diet supplemented with *Yucca schidigera* or Y. schidigera and nisin. *Anim Feed Sci Tech*. 2006; 129(3-4): 187-195.
- 44. Pen B, Takaura K, Yamaguchi S, Asa R, Takahashi J. Effects of *Yucca schidigera* and *Quillaja saponaria* with or without 1-4 galacto-oligosaccharides on ruminal fermentation, methane production and nitrogen utilization in sheep. *Anim Feed Sci Tech.* 2007; 138(1): 75-88
- 45. Liu CL, Li ZQ, Du J, Shan AS. The effect of *Yucca schidigera* extract on ruminal fermentation and parameters traits in sheep. *Agr Sci China*. 2007; 6(1): 121-128.
- 46. Wang CJ, Wang SP, Zhou H. Influences of flavomycin, ropadiar, and saponin on nutrient digestibility, rumen fermentation, and methane emission from sheep. *Anim Feed Sci Tech.* 2009; 148(2-4): 157-166.
- 47. Liu CL, Li ZQ. Effect of levels of *Yucca schidigera* extract on ruminal fermentation parameters, digestibility of nutrients and growth performance in sheep. *Adv Mater*. 2011; 343-344: 655-660.
- 48. Canul-Solis JR, Piñeiro-Vázquez AT, Briceño-Poot EG, Chay-Canul AJ, Alayón-Gamboa JA, Ayala-Burgos AJ, et al. Effect of supplementation with saponins from *Yucca schidigera* on ruminal methane production by Pelibuey sheep fed Pennisetum purpureum grass. *Anim Prod Sci.* 2014; 54(10): 1834-1837.

- 49. Rira M, Chentli A, Boufenera S, Bousseboua H. Effects of plants containing secondary metabolites on ruminal methanogenesis of sheep *in vitro*. *Energy Procedia*. 2015; 74: 15-24.
- 50. Cabuk M, Alcicek A, Bozkurt M, Akkan S. Effect of *Yucca schidigera* and natural zeolite on broiler performance. *Int J Poultry Sci.* 2004; 3(10): 651-654.
- 51. Corzo A, Kidd MT, Miles DM, Dozier WA, Cheeke PR. *Yucca schidigera* and *Quillaja saponaria* supplementation in broiler diets abstract. International Poultry Scientific Forum; 2007. 64 p.
- 52. Chepete HJ, Xin H, Mendes LB, Li H, Bailey TB. Ammonia emission and performance of laying hens as affected by different dosages of *Yucca schidigera* in the diet. *J Appl Poultry Res.* 2012; 21(3): 522-530.
- 53. Onbasilar EE, Erdem E, Ünal N, Kocakaya A, Torlak E. Effect of *Yucca schidigera* spraying in diffrent litter materials on some litter traits and breast burn of broilers at the fifth week of production. *Kafkas Univ Vet Fak Derg*. 2013; 19(5): 749-753.
- 54. Panetta DM, Powers WJ, Xin H, Kerr BJ, Stalder KJ. Nitrogen excretion and ammonia emissions from pigs fed modified diets. *J Environ Qual*. 2006; 35(4): 1297-1308.
- 55. Hong JW, Kim IH, Moon TH, Kwon OS, Lee SH, Kim YG. Effects of yucca extract and (or) far infrared emitted materials supplementation on the growth performance, serum characteristics and ammonia production of growing and finishing pigs. *Asian Austral J Anim.* 2001; 14(9): 1299-1303.
- 56. Liang GQ, Wang XP, Wang XM, Li CP, Chen AG. Effects of camphor familiar plant extract and Yucca extracts on emission of NH₃ and H₂S in slurry of weaned pigs. *Chinese J Anim Sci.* 2009; 45(13): 22-26 (in Chinese).
- 57. Wallace RJ, Arthaud L, Newbold CJ. Influence of *Yucca shidigera* extract on ruminal ammonia concentrations and ruminal microorganisms. *Appl Environ Microbiol*. 1994; 60(6): 1762-1767.
- 58. Wang Y, McAllister TA, Newbold CJ, Rode LM, Cheeke PR, Cheng KJ. Effect of *Yucca schidigera* extract on fermentation and degradation of steroidal saponins in rumen simulation technique (RUSITEC). *Anim Feed Sci Tech.* 1998; 74(2): 143-153.
- 59. Stumm CK, Zwart KB. Symbiosis of protozoa with hydrogen utilizing methanogens. *Microbiol Sci.* 1986; 3(4): 100-105.
- 60. Ushida K, Tokura M, Takenaka A, Itabashi H. Ciliate protozoa and ruminal methanogenesis. In: Onodera R, Itabashi H, Ushida K, Yano H, Sasaki Y, eds. Rumen Microbes and Digestive Physiology in Ruminants. Tokyo: Japan Scientific Societies Press & S. Karger AG Basel; 1997. p. 209-220.
- 61. Newbold CJ, Lassalas B, Jouany JP. The importance of methanogens associated with ciliate protozoa in ruminal methane production *in vitro*. *Lett Appl Microbiol*. 1995; 21(4): 230-234.
- 62. Guo YQ, Liu JX, Lu Y, Zhu WY, Denman SE, McSweeney CS. Effect of tea saponin on methanogenesis, microbial community structure and expression of mcrA gene, in cultures of rumen microorganisms. *Lett Appl Microbiol*. 2008; 47(5): 421-426.
- 63. Moss AR, Jouany JP, Newbold J. Methane production by ruminants: its contribution to global warming. *Ann Zootech*. 2000; 49(3): 231-253.
- 64. Cieslak A, Zmora P, Pers-Kamczyc E, Szumacher-Strabel M. Effects of tannins source (*Vaccinium vitis idaea* L.) on rumen microbial fermentation *in vivo*. *Anim Feed Sci Tech*. 2012; 176(1-4): 102-106.
- 65. Wolin MJ, Miller TL. Microbe-microbe interactions. In: The Rumen Microbial Ecosystem. London: Elsevier Applied Science; 1988. p. 343-359.
- 66. Strobel HJ, Russell JB. Non-proton-motive-force-dependent sodium efflux from the ruminal bacterium *Streptococcus bovis*: bound versus free pools. *Appl Environ Microbiol*. 1989; 55(10): 2664-2668.
- 67. Goel G, Makkar HPS. Methane mitigation from ruminants using tannins and saponins. *Trop Anim Health Pro*. 2012; 44(4): 729-739.
- 68. Cardozo PW, Calsamiglia S, Ferret A, Kamel C. Screening for the effects of natural plant extracts at two pH levels on *in vitro* rumen microbial fermentation of a high-concentrate diet for beef cattle. *J Anim Sci.* 2005; 83(11): 2572-2579.

- 69. Mackie RI, Gilcrist FMC, Heath S. An *in vitro* study of ruminal microorganisms influencing lactate turnover and its contribution to volatile fatty acid production. *J Agric Sci.* 1984; 103(1): 37-51.
- 70. Newbold CJ, EL Hassan SM, Wang J, Ortega ME, Wallace RJ. Influence of foliage from African multipurpose trees on activity of rumen protozoa and bacteria. *Brit J Nutr.* 1997; 78(2): 237-249.
- 71. Teferedegne B. New perspectives on the use of tropical plants to improve ruminant nutrition. *P Nutr Soc.* 2000; 59(2): 209-214.
- 72. Wallace RJ, McEwan NR, McIntosh FM, Teferedegne B, Newbold CJ. Natural products as manipulators of rumen fermentation. *Asian Austral J Anim.* 2002; 15(10): 1458-1468.
- 73. Hristov AN, Grandeen KL, Ropp JK, Greer D. Effect of *Yucca schidigera*-based surfactant on ammonia utilization *in vitro*, and in situ degradability of corn grain. *Anim Feed Sci Tech.* 2004; 115(3-4): 341-355.
- 74. Hristov AN, McAllister TA, Herk FHV, Cheng KJ, Newbold CJ, Cheeke PR. Effect of *Yucca schidigera* on ruminal fermentation and nutrient digestion in heifers. *J Anim Sci*. 1999; 77(9): 2554-2563.
- 75. Pratt EV, Rose SP, Keeling AA. Effect of ambient temperature on losses of volatile nitrogen compounds from stored laying hen manure. *Bioresource Technol*. 2002; 84(2): 203-205.
- 76. Lundeen T. Yucca extract improves feed efficiency, decreases abdominal fat in broilers. *Feedstuffs*. 2000; 72(33): 9.
- 77. Nahm KH. Evaluation of the nitrogen content in poultry manure. World Poultry Sci J. 2003; 59(1): 77–88.
- 78. Reece FN, Lott BD, Deaton JW. Ammonia in the atmosphere during brooding affects performance of broiler chickens. *Poultry Sci.* 1980; 59(3): 486-488.
- 79. Gay SW, Wheeler EF, Zajaczkowski JL, Topper PA. Ammonia emissions from U.S. tom turkey growout and brooder houses under cold weather minimum ventilation. *Appl Eng Agric*. 2006; 22(1): 127-134.
- 80. Choi IH, Moore PA. Effect of various litter amendments on ammonia volatilization and nitrogen content of poultry litter. *J Appl Poultry Res.* 2008; 17(4): 454-462.
- 81. Blummel M, Becker K. The degradability characteristics of fifty-four roughages and roughage neutral-detergent fibers as described by *in vitro* gas production and their relationship to voluntary feed intake. *Brit J Nutr.* 1997; 77(5): 757-768.
- 82. Blummel M, Steingaβ H, Becker K. The relationship between *in vitro* gas production, *in vitro* microbial biomass yield and 15N incorporation and its implications for the prediction of voluntary feed intake of roughages. *Brit J Nutr.* 1997; 77(6): 911-921.
- 83. Blummel M, Karsli A, Russell JR. Influence of diet on growth yields of rumen microorganisms *in vitro* and *in vivo*: influence on growth yield of variable carbon fluxes to fermentation products. *Brit J Nutr.* 2003; 90(3): 625-634.
- 84. Hess HD, Kreuzer M, Diaz TE, Lascano CE, Carulla JE, Soliva CR, et al. Saponin rich tropical fruits affect fermentation and methanogenesis in faunated and defaunated rumen fluid. *Anim Feed Sci Tech.* 2003a; 109(1): 79-94.
- 85. Hess HD, Monsalve LM, Lascano CE, Carulla JE, Diaz TE, Kreuzer M. Supplementation of a tropical grass diet with forage legumes and Sapindus saponaria fruits: Effects on *in vitro* ruminal nitrogen turnover and methanogenesis. *Aust J Agric Res.* 2003b; 54(7): 703-713.
- 86. Huque KS, Chowdhury SA. Study on supplementing effects or feeding systems of molasses and urea on methane and microbial nitrogen production in the rumen and growth performances of bulls fed a straw diet. *Asian Austral J Anim.* 1997; 10(1): 35-46.
- 87. Mehra UR, Khan MY, Lal M, Hasan QZ, Das A, Har R, et al. Effect of source of supplementary protein on intake, digestion and efficiency of energy utilization in buffaloes fed wheat straw based diets. *Asian Austral J Anim.* 2006; 19(5): 638-644.
- 88. Todd RW, Cole NA, Clark RN. Reducing crude protein in beef cattle diet reduces ammonia emission from artificial feedyard surfaces. *J Environ Qual.* 2006; 35(2): 404-411.
- 89. Hungate RE. The rumen and its microbes. New York: Academic Press; 1966.
- 90. Wang Y, Greer D, McAllister TA. Effect of a saponin-based surfactant on water absorption, processing characteristics and *in vitro* ruminal fermentation of barley grain. *Anim Feed Sci Tech.* 2005; 118(3-4): 255-266.

- 91. Menke, K.H., Steingass, H., Estimation of the energetic feed value from chemical analysis and *in vitro* gas production using rumen fluid. *Anim Res Dev.* 1988; 28: 7-55.
- 92. Wang Y, McAllister TA, Cheeke PR, Cheng KJ. Assessment of inhibitory effects of ruminal fluid on biological activity of saponins using hemolytic assay. *Can J Anim Sci*. 1999; 79(4): 561-564.
- 93. Cole NA, Todd RW. Nitrogen and phosphorus balance of beef cattle feedyards. Proceedings of the Texas Animal Manure Management Issues Conference; September 29-30, 2009; Round Rock, TX.
- 94. Patra AK, Stiverson J, Yu Z. Effects of quillaja and yucca saponins on communities and select populations of rumen bacteria and archaea, and fermentation *in vitro*. *J Appl Microbiol*. 2012; 113(6): 1329-1340.
- 95. Kowalczyk M, Pecio Ł, Stochmal A, Oleszek W. Qualitative and quantitative analysis of steroidal saponins in crude extract and bark powder of *Yucca schidigera* Roezl. *J Agric Food Chem.* 2011; 59(15): 8058-8064.
- 96. Duffy CF, Killeen GF, Connolly CA, Power RF. Effects of dietary supplementation with *Yucca schidigera* Roezl ex Ortgies and its saponin and non-saponin fractions on rat metabolism. *J Agric Food Chem.* 2001; 49(7): 3408-3413.
- 97. Mader TL, Brumm MC. Effect of feeding sarsaponin in cattle and swine diets. *J Anim Sci*, 1987; 65(1): 9-15.
- 98. Wisløff, H, Uhlig S, Scheie E, Loader J, Wilkins A, Flaøyen A. Toxicity testing of saponin-containing *Yucca schidigera* Roetzl. juice in relation to hepato- and nephrotoxicity of *Narthecium ossifragum* (L.) Huds. *Toxicon*. 2008; 51(1): 140-150.
- 99. Goel G, Makkar HPS, Becker K. Changes in microbial community structure, methanogenesis and rumen fermentation in response to saponin-rich fractions from different plant materials. *J Appl Microbiol*. 2008; 105(3): 770-777.
- 100. Makkar HPS, McSweeney CS. Methods in gut microbial ecology for ruminants. Dordrecht: Springer; 2005.
- 101. Makkar HPS. *In vitro* gas method for evaluation of feeds containing phytochemicals. *Anim Feed Sci Tech*. 2005; 123-124: 291-302.
- 102. Panetta DM, Powers WJ, Xin H, Kerr BJ, Stalder KJ. Nitrogen excretion and ammonia emissions from pigs fed modified diets. *J Environ Qual*. 2006; 35(4): 1297-1308.
- 103. McGinn SM, Beauchemin KA, Iwaasa AD, McAllister TA. Assessment of the sulfur hexafluoride (SF6) tracer technique for measuring enteric methane emission from cattle. *J Environ Qual*. 2006; 35(5): 1686-1691.

Received: February 03, 2016; Accepted: July 14, 2016