

Synthesis, Characterization and *in vitro* Antioxidant Activity of New Chiral *N*-boc Organotellurium Compounds, (CH₃)₃OC(O)NHCH(R)C(O)NHCH₂-CH₂Te-C₆H₄-4-OCH₃, Containing Carbamate and Peptide Groups

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Synthesis, characterization and antioxidant activity of a new series of chiral *N*-boc oraganotellurium compounds, $(CH_3)_3OC(O)NHCH(R)C(O)NHCH_2CH_2Te-C_6H_4-4-OCH_3$, containing carbamate and peptide groups have been reported in this paper. These chiral peptides were synthesized in good to excellent yields, via acid-amine coupling reaction of *N*-boc *L*-amino acids with 2-(4-methoxyphenyltelluro) ethylamine in presence of dicyclohexyl carbodimide (DCC) at room temperature. The elemental analyses, Fourier transform infrared (FTIR), ¹H and ¹³C {¹H} nuclear magnetic resonance (NMR) spectra and mass spectra were characteristic. Specific optical rotation (SOR) was also determined. *In vitro* antioxidant activity of these multi-functional compounds in methanol has been evaluated against 2,2-diphenyl-1-picrylhydrazyl (DPPH) radicals with 2,6-di-*tert*-butyl-4-methylphenol (BHT) as a standard reference compound. The IC₅₀ (inhibitory concentration 50) values of these chiral peptides revealed significant inhibition against DPPH radicals and found to be effective antioxidants.

Keywords: organochalcogen, aminoacid derivative, antioxidant

Introduction

An antioxidant is "any substance that delays or prevents oxidation of an oxidisable species".^{1,2} Oxidative stress, induced by reactive oxygen species (ROS) produced in the body, is one of the main factors of current slow killer diseases, that population suffering from, like cancer, diabetes, cardiovascular, neurological (Alzheimer's and Parkinson's), inflammatory viral diseases and digestive disorders.³⁻⁶

Since the discovery of "Ebselen", a natural antioxidant, a number of organochalcogen (S, Se, Te) compounds have been designed, synthesized and investigated as free radical scavengers or antioxidants to mimic the natural ones.⁷⁻³⁰ The divalent organochalcogen compounds have the potential to act as catalytic antioxidants because they readily react with many types of oxidants such as peroxides, peroxynitrites, singlet oxygen and ozone resulting tetravalent organochalcogens (that is chalcogenides to chalcogenoxides and chalcogenols to dichalcogenides) which in turn reduced by many mild reducing agents. The antioxidant activity of organochalcogen compounds is also related to their redox potentials and rates of oxidation in solutions.³¹⁻³⁵ Even though a large number of antioxidants are available, they have many disadvantages such as they are costly and many of them cause various side effects due to the toxic byproducts.^{36-40,22}

Organotellurium compounds have shown many advantages as free radical scavengers or antioxidants. They act as both peroxide decomposers and chain-breaking, donating antioxidants even under biologically relevant conditions. They have also demonstrated higher radical scavenging efficiency than the corresponding S and Se compounds.^{41,42}

Organotellurium compounds have been much less explored in biology unlike the compounds of its lower congeners (S and Se), even though tellurium exhibit various similar properties of sulfur and selenium⁴³⁻⁴⁶ which may be due to the fact that tellurium was considered non-essential biological trace element, commercial non-availability of a wide variety of organotellurium compounds as starting materials and some misconception that they are air sensitive, foul smelling and even toxic. A few reasons among enhanced interest currently in the biological chemistry of organotellurium compounds are the availability of modern analytical techniques (¹²⁵Te Fourier transform nuclear magnetic resonance, FT-NMR; atomic absorption spectroscopy, AAS; and inductively coupled plasma-atomic

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emission spectrometry, ICP-AES) to study and understand the solution behavior of Te containing systems, such an option does not exist for sulfur compounds, the secondary interactions (Te^{...}X where X = O, Cl, Br, I or M) arises from the hypervalent nature of Te, thus increases the stability of organotellurium compounds and higher redox activity of the tellurium.

Numerous organochalcogen compounds containing amine, peptide/amide and carbamate groups enhances the rates of their redox reactions^{16,22,23,47} and the effect of these groups on the activity of organotellurium compounds was expected to be more pronounced than that of corresponding organoselenium compounds.^{47,23} The biological activity of chiral organochalcogen compounds may be specific. Recently reported that the organoselenium and tellurium compounds which contain the secondary interactions between chalcogen atom and other heteroatoms [TeX, X = N, O, S, Cl, Br, I] through internal chelation play an important role in the catalytic antioxidant activity.⁴⁸⁻⁵⁰

There are numerous synthetic and natural chiral organoselenium compounds^{20-26,43-46} but there has not been studied antioxidant properties of chiral organotellurium compounds containing peptide and carbamate functionalities.

In view of the above advantages and to expand the applications of chiral organotellurium compounds in biological activities⁵¹ particularly as antioxidants, we have reported in this paper the synthesis, spectral and analytical characterization data of chiral *N*-boc organotellurium compounds containing amide and carbamate groups. These chiral multi-functionalized compounds (**4a-4g**) synthesized from *N*-boc *L*-amino acids and 2-(4-methoxyphenyltelluro) ethylamine via condensation in presence of dicyclohexyl carbodimide (DCC). The *in vitro* antioxidant activity of these compounds against 2,2-diphenyl-1-picrylhydrazyl (DPPH) radicals has been studied. All the compounds showed significant antioxidant activity when compared to 2,6-di-*tert*-butyl-4-methylphenol (BHT).

Results and Discussion

Synthesis and characterization of chiral *N*-boc organotellurated peptides (**4a-4g**)

A new series of chiral *N*-boc organotellurated compounds (**4a-4g**, Figure 1) were synthesized by condensation of *N*-boc *L*-amino acids and 2-(4-methoxyphenyltelluro) ethylamine in presence of DCC as a dehydrating agent according to the reactions given in the Scheme 1. The reactions were carried out in dry dichloromethane. The products were purified by flash column chromatography. The compounds, **4a-4g**, were obtained with good to

excellent yields of about 80-95%.

FTIR spectra of chiral *N*-boc organotellurated peptides (4a-4g)

Fourier transform infrared (FTIR) spectra were recorded in the range of 400-4000 cm⁻¹. In the IR spectra of compounds, **4a-4g**, characteristic bands were obtained for NH, C–H, C=C, C=O, C–O, C–S and C–Te functional groups. The –NH– stretching bands were observed at around at 3300-3450 cm⁻¹ and the C=O stretching vibrational bands were obtained at 1630-1660 cm⁻¹. The C–S stretching band in **4d** and **4e** were appeared at 733 cm⁻¹. The C–Te aryl stretching vibrational band was appeared in the region of 510 ± 5 cm⁻¹.⁵²⁻⁵⁵

¹H NMR spectra of chiral *N*-boc organotellurated peptides (4a-4g)

The ¹H NMR spectra of the compounds **4a-4g** were found characteristic. The CH₂Te and CH₂N signals were observed at δ , 2.734-2.814 and 3.290-3.375 ppm and appeared as triplets and multiplets respectively. These signals were found deshielded about 0.4 to 0.5 ppm when compared to 2-(4-methoxyphenyltelluro) ethylamine.55 There are two types of NH groups such as the carbamate (Boc-NH-) and amide (>CO-NH-). The carbamate NH proton gave a doublet in the range of δ , 6.5 to 6.9 ppm while the amide NH gave a triplet between 7.95 to 8.25 ppm due to coupling with CH and CH2 protons respectively. In all the compounds a singlet was observed at around 1.35 ppm for all the 9 protons of t-Bu group. In 4a the methyl group showed a doublet at 1.118-1.136 ppm. In 4b the benzylic CH2 protons were split into two doublets of a doublet at 2.672-2.705 and 2.891-2.924 ppm due to their diastereotopic nature. In 4c the CH, CH2 and CH3 protons all gave multiplets between 0.77 to 1.25 ppm. The protons of -CH₂-CH₂-S-CH₃ group in 4d were appeared as a multiplet, triplet and a singlet at 1.698-1.771, 2.384-2.423 and 2.013 ppm respectively. In 4e, the -CH₂SCH₂Ph, protons were observed as two multiplets. In 4f the phenolic OH was obtained as a singlet at 9.12 ppm. The aromatic protons present at meta and ortho to Te were observed at 6.804-6.9 and 7.606-7.663 ppm respectively in all the compounds which were in agreement with that reported for 2-(4-methoxyphenyltelluro) ethylamine.⁵⁵ The peaks for other aromatic protons were characteristic.

¹³C NMR spectra of chiral *N*-boc organotellurated peptides (4a-4g)

In the ¹³C nuclear magnetic resonance (NMR) spectra of



Figure 1. Structure of chiral N-boc organotellurated peptides (4a-4g).



Scheme 1. Synthesis of chiral *N*-boc organotellurated peptides (**4a-4g**): (a) EtOH, NaBH₄/aq. NaOH, CH₂Cl₂; (b) **3a-3f**, DCC, THF; (c) **3g**, DCC, THF; (d) antioxidant activity against DPPH radicals.

4a-4g, the CH₂Te and CH₂N carbon signals were appeared between δ , 22-25 ppm and 40-41 ppm, respectively.⁵⁶ The methyl carbons of 'Bu group in 4d and 4f are slightly deshielded and appeared at 28.63 and 29.87 ppm, respectively, whereas in others these carbon were appeared at 28.07-28.16 ppm. The quaternary carbon peaks of 'Bu group were produced at δ ca. 78 ppm. The carbons atoms present at position meta and ortho to Te were appeared at around 115.5 and 140.0 ppm, respectively. The carbamate $[(CH_3)_3CO(CO)NH_-]$ and amide $[-NH(CO)CH_]$ carbons are highly deshielded and appeared in the range of 155.0-155.5 and 170-171 ppm, respectively, in all 4a-4g. The methyl carbon was observed at 18.29 ppm in 4a. In 4b and 4f the benzylic methylene carbon signal was appeared at ca. 37 ppm. In 4d the carbons of CH₃SCH₂CH₂- chain were obtained at 15.09, 32.21 and 30.24 ppm, respectively. The signals for CH₂SCH₂- carbons were observed at 35.10 and 47.44 ppm, respectively, in 4e. The carbon atoms meta and ortho to Te were observed between 115.2-115.7 and 139.0-140.5 ppm, respectively in 4a-4g.⁵⁶ The other aromatic signals are characteristic for their respective carbons.

Mass spectra of 4a, 4b and 4g

Mass spectra of compounds **4a**, **4b** and **4g** were recorded using atmospheric-pressure chemical ionization (APCI) in the positive mode. The [M + 1] ion peak were obtained at m/z 450.9, 527 and 476.9 with 20, 30 and 10% abundance. In all, the peaks were also obtained with [M + 1 + H₂O] at 467.9, 545 and 494.9 with 100% abundance.

Radical scavenging activity of chiral *N*-boc organotellurated peptides (**4a-4g**)

In order to understand the influence of tellurium atom along with carbamate and amide groups by varying the substituent at chiral carbon in the biological activity, such as the antioxidant activity of compounds **4a-4g** in comparison with BHT, a common antioxidant that finds application in the food industry,^{57,58} was determined by DPPH assay *in vitro* as given in Table 1.

DPPH is characterized as a stable free radical by virtue of the delocalization of the single electron over the molecule as a whole, so that the molecules do not dimerise, as would be the case with most other free radicals. The delocalization also gives rise to the deep violet color, characterized by an absorption band in ethanol solution centered at about 520 nm.

When a solution of DPPH is mixed with that of a substance that can donate an electron or hydrogen atom, then this gives rise to the reduced form with the loss of

Table 1. Antioxidant capacities of chiral N-boc organotellurated peptides (4a-4g)

Concentration / µM	Inhibition / %						
	4a	4b	4c	4d	4e	4f	BHT
0	0	0	0	0	0	0	0
4	20	30	33	33	30	31	38
8	42	48	53	51	51	49	60
12	59	57	66	58	62	63	67
16	64	61	68	64	64	65	70

this violet color.⁵²⁻⁵⁵ This latter will then undergo further reactions which control the overall stoichiometry, that is, the number of molecules of DPPH reduced (decolorized) by one molecule of the reductant. The reaction is therefore intended to provide the link with the reactions taking place in an oxidizing system, such as the autoxidation of a lipid or other unsaturated substance; the DPPH molecule is thus intended to represent the free radicals formed in the system whose activity to be suppressed by the substance.

Antioxidant potential of chiral *N*-boc organotellurated amides were assessed through DPPH radicals with varying concentration as given in the Table 1 and also shown in Figure 2 which demonstrated that these multi-functionalized compounds (**4a-4g**) are effective antioxidants than BHT towards DPPH radicals due to the presence of Te and which is also influenced by carbamate and amide groups.



Figure 2. Graphical representation of antioxidant activity of 4a-4g against DPPH radicals.

The IC₅₀ (inhibitory concentration 50) values (sample concentration required to inhibit 50% of the radicals) of **4a-4g** are in the range of 6.00 to 10.00 μ M mL⁻¹ as given in the Table 2 and also represented graphically in Figure 3. These values suggested that the chiral *N*-boc organotellurated compounds containing carbamate and amide functionalities showed good antioxidant activities.



Figure 3. IC₅₀ values of chiral *N*-boc organotellurated peptides (4a-4g).

Table 2. IC₅₀ values of 4a-4g

Compound	IC ₅₀ / µM			
4a	9.9			
4b	8.8			
4c	7.4			
4d	7.7			
4e	7.8			
4f	8.2			
4g	6.2			

IC₅₀: inhibitory concentration 50.

Conclusions

In summary, we presented the synthesis of new chiral N-boc oraganotellurium compounds (4a-4g) containing carbamate and peptide groups from N-boc L-amino acids and 2-(4-methoxyphenyltelluro) ethylamine in presence of DCC. All these compounds were characterized by elemental analysis, FTIR, ¹H, and ¹³C {¹H} NMR spectroscopy and mass spectrometry. Specific optical rotations were determined for 4a-4g. In vitro antioxidant activity of 4a-4g in methanol has been investigated against DPPH radicals with BHT as a standard reference compound. The IC₅₀ values of the chiral N-boc organotellurrated peptides (4a-4g) were found in the range of 6 to 10 µM concentration. These results revealed that the new chiral N-boc organotellurated peptides (4a-4g) proved to possess significantly high antioxidant activity against DPPH radicals and among which compound 4g, L-proline based peptide showed highest antioxidant activity.

Experimental

Analytical techniques

 1H and ^{13}C { 1H } NMR spectra were recorded on a Bruker DPX-400 Hz or AVANCE II Bruker 400 Hz spectrometer at

399.65 and 100.64 Hz, respectively. Spectra were recorded in dimethyl sulfoxide, DMSO- d_6 , solutions. Chemical shifts were reported in parts *per* million (ppm) with tetramethylsilane (TMS) as a reference standard. Specific optical rotations (SOR) were measured using Rudolph Autopol-I automatic Polarimeter in a cell of 100 mm path length. Melting points were recorded in open capillary tubes and are reported uncorrected. FTIR spectra were recorded on a JASCO FTIR-4100 instrument in the range of 400 to 7000 cm⁻¹. Column chromatography was performed using Merck silica gel (230-400 mesh). Thin layer chromatography (TLC) was performed using Merck 60 F254 silica gel on pre-coated aluminum sheets. Compounds were visualized on TLC with UV-light, potassium permanganate stain, iodine vapor or with ninhydrin.

Materials

Solvents such as tetrahydrofuran (THF) and benzene were dried using sodium metal and benzophenone. *N*-boc *L*-aminoacids, sodium borohydride, elemental tellurium, anisole, 2-chloroethylamine hydrochloride, DCC, DPPH and BHT were purchased from Sigma Aldrich Ind. Ltd., and they were used as received. The following precursor compounds were prepared according to literature procedures: 1,2-bis(4-methoxyphenyl) ditellane,⁵⁹ 2-(4-methoxyphenyltelluro) ethylamine.⁵²⁻⁵⁵

General procedure for the synthesis of chiral *N*-boc *L*-organotellurated amides (**4a-4f**)

The stirred solution of *N*-boc *L*-amino acids, (10 mmol) in dry CH₂Cl₂ (50 mL) was cooled to 0 °C and then was added DCC (2.063 g, 10 mmol). The reaction mixture was slowly brought to room temperature and continued the stirring for about 2-3 h. A solution of 2-((4-methoxyphenyl) tellanyl) ethylamine dissolved in 15-20 mL of CH₂Cl₂ was added dropwise. The whole reaction mass was further stirred at room temperature. The progress of the reaction was checked by TLC. After completion of the reaction, the resulting mixture was diluted with CH₂Cl₂ and the organic layer was washed thrice with brine (3 × 50 mL). The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered. The solvent was removed under reduced pressure. The crude product obtained above was subjected to column chromatography hence gave compounds **4a-4f**.

(S)-tert-Butyl(1-((2-((4-methoxyphenyl)tellanyl)ethyl) amino)-1-oxopropan-2-yl) carbamate (4a)

Light brown viscous liquid; yield 85%; $[\alpha]_{D}^{28.5}$ –20.0 (*c* 1.0, CHCl₃); IR (KBr) v / cm⁻¹ 3338, 3306, 2976, 2926,

2849, 1686, 1644, 1524, 1488, 1449, 1365, 1320, 1280, 1240, 1162, 1065, 1024, 810, 638, 585, 511, 463, 446, 419; ¹H NMR (399.65 Hz, DMSO- d_6) δ 1.118-1.136 (d, 3H, *J* 7.2 Hz, H12), 1.357 (s, 9H, 'Bu), 2.779-2.814 (t, 2H, *J* 7.59 Hz, H5), 3.290-3.375 (m, 2H, H4), 3.73 (s, 3H, OCH₃), 3.843-3.879 (m, 1H, H2), 6.804-6.833 (m, 3H, H8, H10, NH), 7.612-7.663 (d, 2H, *J* 8.4 Hz, H7, H11), 7.964-7.993 (t, H, *J* 5.99 Hz, NH); ¹³C NMR (100.64 Hz, DMSO- d_6) δ 18.29 (CH₃), 24.43 (C5), 28.17 ('Bu), 40.72 (C2), 40.72 (C4), 55.23 (OCH₃), 77.93 ('BuCO), 100.18 (C6), 115.57 (C8, C10), 139.90 (C7, C11), 154.94 (C1), 159.27 (C9), 172.34 (C3); MS (+ pESI) *m/z*, calcd. for C₁₇H₂₆N₂O₄Te [M⁺]: 450; found (% abundance): 150 (90), 288 (30), 330 (70), 450.9 (20), 467.9 (100).

(S)-tert-Butyl(1-((2-((4-methoxyphenyl)tellanyl)ethyl) amino)-1-oxo-3-phenylpropan-2-yl) carbamate (**4b**)

Colourless solid; yield 90%; $[\alpha]_{D}^{29.0}$ –9.5 (*c* 1.0, CHCl₃); mp 108-110 °C; elemental anal. calcd. for $C_{23}H_{30}N_2O_4Te$ (found): C 52.51 (52.98), H 5.75 (5.75), N 5.32 (5.41); IR (KBr) v / cm⁻¹ 3392, 3061, 2968, 2935, 1688, 1656, 1585, 1525, 1487, 1455, 1389, 1365, 1302, 1285, 1243, 1175, 1159, 1066, 1029, 1005, 948, 846, 817, 788, 754, 701, 659, 635, 589, 511; ¹H NMR (399.65 Hz, DMSO-*d*₆) δ 1.279 (s, 9H, ^tBu), 2.672-2.705 (dd, 1H, J13.18 Hz, H12), 2.739-2.777 (t, 2H, J7.59 Hz, H5), 2.891-2.924 (dd, 1H, J13.18 Hz, H12), 3.288-3.396 (m, 2H, H4), 3.726 (s, 3H, OCH₃), 4.058-4.068 (m, 1H, H2), 6.809-6.831 (d, 2H, J 8.4 Hz, H8, H10), 6.849-6.870 (d, 1H, J 8.39 Hz, NH), 7.162-7.240 (m, 5H, H14 to H18), 7.614-7.636 (d, 2H, J 8.7 Hz, H7, H11), 8.117 (bt, H, NH); ¹³C NMR (100 Hz, CDCl₃) δ 25.05 (C5), 28.07 (^tBu), 37.61 (C12), 40.79 (C4), 55.69 (C2), 54.95 (OCH₃), 77.87 ('BuCO), 100.21 (C6), 115.29 (C8, C10), 126.06 (C16), 127.91 (C14, C18), 129.10 (C15,17), 138.08 (C13), 139.80 (C7, C11), 155.07 (C1), 159.23 (C9), 171.17 (C3); MS (+ pESI) m/z, calcd. for C₂₃H₃₀N₂O₄Te [M⁺]: 526.1; found (% abundance): 365 (40), 404 (70), 527 (30), 545 (100).

(S)-tert-Butyl((2S,3S)-1-((2-((4-methoxyphenyl)tellanyl) ethyl)amino)-3-methyl-1-oxopentan-2-yl) carbamate (**4c**)

Colourless solid; yield 95%; $[\alpha]_D^{27.5} - 13.5$ (*c* 1.0, CHCl₃); mp. 61-62 °C; elemental anal. for C₂₀H₃₂N₂O₄Te: calcd. (found): C 48.82 (48.90), H 6.55 (6.55), N 5.69 (5.75); IR (KBr) v / cm⁻¹ 3314, 3061, 3085, 2962, 2930, 2876, 2853, 1679, 1644, 1518, 1486, 1281, 1243, 1164, 1019, 640, 586, 512, 447, 420; ¹H NMR (400.23 Hz, DMSO-*d*₆) δ 0.769-0.807 (m, 6H, H14, H15), 1.011-1.065 (m, 2H, H13), 1.197-1.247 (m, 1H, H12), 1.357 (s, 9H, 'Bu), 2.779-2.817 (t, 2H, *J* 7.99 Hz, H5), 3.328-3.413 (m, 2H, H4), 3.692-3.794 (m, 1H, H2), 3.731 (s, 3H, OCH₃), 6.591-6.613 (d, 1H, *J* 8.79 Hz, NH), 6.799-6.835 (d, 2H, *J* 8.4 Hz,

H8, H10), 7.606-7.642 (d, 2H, *J* 8.7 Hz, H7, H11), 8.068 (bt, 1H, NH); ¹³C NMR (100.64 Hz, DMSO- d_6) δ 10.97 (C10), 15.47 (C18), 24.38 (C13), 25.30 (C5), 28.16 ('Bu), 40.15 (C2), 40.77 (C4), 54.99 (OCH₃), 77.90 ('BuCO), 100.23 (C6), 115.33 (C8, C10), 139.91 (C7, C11), 155.26 (C1), 159.28 (C9), 171.09 (C3).

(S)-tert-Butyl(1-((2-((4-methoxyphenyl)tellanyl)ethyl) amino)-4-(methylthio)-1 oxobutan-2-yl)carbamate (4d)

Light brown viscous liquid; yield 80%; $[\alpha]_D^{28.0} - 2.0$ (*c* 1.0, CHCl₃); IR (KBr) v / cm⁻¹ 3320, 3061, 2974, 2929, 1696, 1586, 1520, 1489, 1439, 1391, 1366, 1283, 1245, 1174, 1046, 1025, 963, 861, 822, 788, 758, 695, 635, 587, 515, 485; ¹H NMR (399.65 Hz, DMSO-*d*₆) δ 1.361 (s, 9H, 'Bu), 1.698-1.771 (m, 2H, H12), 2.013 (s, 3H, H14), 2.384-2.423 (t, 2H, *J* 7.19 Hz, H13), 2.791-2.825 (t, 2H, *J* 6.39 Hz, H5), 3.295-3.409 (m, 2H, H4), 3.730 (s, 3H, OCH₃), 3.887-3.939 (m, 1H, H2), 6.896-6.916 (d, 1H, *J* 7.99 Hz, NH), 6.804-6.826 (d, 2H, *J* 8.4 Hz, H8, H10), 7.611-7.633 (d, 2H, *J* 8.7 Hz, H7, H11), 8.022-8.049 (t, H, *J* 5.59 Hz, NH); ¹³C NMR (100.64 Hz, DMSO-*d*₆) δ 15.09 (C12), 28.63 ('Bu), 30.24 (C14), 32.21 (C13), 33.79 (C5), 41.17 (C4), 47.96 (C2), 55.46 (OCH₃), 78.50 ('BuCO), 100.66 (C6), 115.79 (C8, C10), 140.36 (C7, C11), 155.76 (C1), 159.74 (C9), 171.81 (C3).

(*S*)-*tert*-Butyl(3-(benzylthio)-1-((2-((4-methoxyphenyl) tellanyl)ethyl)amino)-1-oxopropan-2-yl) carbamate (**4e**)

Bright yellow solid; yield 95%; $[\alpha]_{D}^{27.0}$ -6.5 (c 1.0, CHCl₃); mp 89-90 °C; elemental anal. for C₂₄H₃₂N₂O₄STe calcd. (found): C 50.38 (50.38), H 5.64 (5.65), N 4.90 (4.95); IR (KBr) v / cm⁻¹ 3357, 3323, 3058, 3023, 2925, 2850, 1685, 1654, 1514, 1487, 1456, 1365, 1282, 1244, 1227, 1156, 1018, 697, 608, 563, 507, 464, 436; ¹H NMR (399.65 Hz, DMSO-d₆) δ 1.394 (s, 9H, 'Bu), 2.672-2.720 (dd, 1H, J 5.59 Hz, C12), 2.803-2.839 (t, 2H, J 6.79 Hz, H5), 3.354-3.442 (m, 1H, C13), 3.701-3.735 (m, 2H, H4), 3.740 (s, 3H, OCH₃), 4.075-4.131 (m, 1H, H2), 6.815-6.837 (d, J 8.4 Hz, 2H, H8, H10), 6.899-6.920 (d, 1H, J 8.39 Hz, NH), 7.221-7.308 (m, 5H, H15 to H19), 7.624-7.646 (d, J 8.7 Hz, 2H, H7, H11), 8.210-8.237 (t, H, J 5.19 Hz, NH); ¹³C NMR (100.64 Hz, DMSO-*d*₆) δ 24.39 (C5), 28.12 ('Bu), 35.10 (C12), 40.86 (C4), 47.44 (C13), 53.70 (C2), 54.95 (OCH₃), 78.16 (BuCO), 100.20 (C6), 115.28 (C8, C10), 126.73 (C17), 128.25 (C15, C19), 128.83 (C16, C18), 138.32 (C14), 139.86 (C7, C11), 155.14 (C1), 159.24 (C9), 170.14 (C3).

(*S*)-*tert*-Butyl(3-(4-hydroxyphenyl)-1-((2-((4-methoxyphenyl) tellanyl)ethyl)amino)-1-oxopropan-2-yl) carbamate (**4f**)

Light brown solid; yield 80%; $[\alpha]_D^{26.0}$ – 5.7 (*c* 1.0, CHCl₃); mp 96-97 °C; elemental anal. for C₂₃H₃₀N₂O₅Te calcd. (found): C 50.96 (50.75), H 5.58 (5.60), N 5.17 (5.25); IR (KBr) v / cm⁻¹ 3319, 2927, 2850, 1646, 1513, 1487, 1438, 1364, 1242, 1158, 1023, 820, 641, 587, 515, 463, 413; ¹H NMR (400.33 Hz, DMSO- d_6) δ 1.30 (s, 9H, 'Bu), 2.570-2.627 (m, 1H, H12), 2.764-2.807 (t, 2H, *J* 7.60 Hz, H5), 2.844-2.924 (dd, 1H, *J* 14.80 Hz, H12), 3.334-3.405 (m, 1H, H4), 3.486 (bs, 1H, H4), 3.731 (s, 3H, OCH₃), 3.913-3.973 (m, 1H, H2), 6.809-6.829 (d, *J* 8.4 Hz, 2H, H7, H10), 6.696-6.717 (d, 1H, *J* 8.40 Hz, NH), 6.614-6.633 (d, 2H, *J* 8.7 Hz, H14, H18), 6.976-6.995 (d, 2H, *J* 8.4 Hz, H15, H17), 7.612-7.633 (d, 2H, *J* 8.7 Hz, H7, H11), 8.034 (bt, H, NH), 9.123 (bs, 1H, OH); ¹³C NMR (100.64 Hz, DMSO- d_6) δ 25.88 (C5), 29.87 ('Bu), 37.40 (C12), 41.28 (C4), 47.96 (C2), 55.46 (OCH₃), 78.35 ('BuCO), 100.72 (C6), 115.24 (C7, C10), 115.80 (C15, C17), 128.54 (C13), 130.49 (C14, C18), 140.30 (C7, C11), 155.56 (C16), 156.53 (C1), 159.73 (C9), 171.83 (C3).

(*S*)-*tert*-Butyl2-((2-((4-methoxyphenyl)tellanyl)ethyl) carbamoyl)pyrolidine-1-carboxylate (**4g**)

Colourless solid; yield 90%; $[\alpha]_{D}^{28.5}$ -36.7 (*c* 1.0, CHCl₃); mp 105-106 °C; elemental anal. for C₁₉H₂₈N₂O₄Te, calcd. (found): C 47.94 (47.54), H 5.93 (5.65), N 5.88 (6.01); IR (KBr) v / cm⁻¹ 3077, 3305, 2998, 2974, 2934, 2870, 2835, 1677, 1658, 1584, 1544, 1486, 1455, 1415, 1368, 1343, 1280, 1244, 1160, 1122, 1060, 1027, 918, 890, 832, 812, 777, 753, $673, 586, 539, 513; {}^{1}\text{H} \text{NMR} (400.23 \text{ Hz}, \text{DMSO-}d_6) \delta 1.306$ (s, 9H, 'Bu), 1.609-1.652 (dd, 1H, J 4.00 Hz, H13), 1.730-1.748 (m, 4H, H12-H14), 2.048-2.069 (dd, 1H, J 5.79 Hz, H2), 2.790-2.830 (t, 2H, H5), 3.215-3.275 (m, 2H, H4), 3.729 (s, 3H, OCH₃), 3.907-3.949 (dd, 1H, J 6.79 Hz, H2), 6.809-6.829 (d, 2H, J 8.4 Hz, H8, H10), 7.609-7.631 (d, 2H, J 8.7 Hz, H7, H11), 8.047-8.061 (t, H, J 5.59 Hz, NH); 13 C NMR (100.64 Hz, DMSO- d_6) δ 23.13 (C13), 25.30 (C5), 28.13 (Bu), 30.97 (C12), 40.82 (C2), 46.62 (C4), 47.48 (C14), 55.25 (OCH₃), 78.37 (BuCO), 100. 27 (C6), 115.32 (C8, C10), 139.81 (C7, C11), 153.61 (C1), 159.28 (C9), 172.18 (C3); MS (+ pESI) m/z, calcd. for C₁₉H₂₈N₂O₄Te [M⁺]: 476.04, found (% abundance): 315 (50), 354 (70), 377 (25), 476.9 (15), 494.9 (100).

Antioxidant activity DPPH radical scavenging evaluation of compounds (4a-4g)

Antioxidant activity was evaluated for the series of newly synthesized chiral *N*-boc organotellurated peptides, **4a-4g**, *in vitro* using DPPH assay following the modified method of Brand-Williams.⁶⁰ DPPH is a stable free radical with purple color which is generally used for determination of electron donating capacity of an antioxidant compound *in vitro*.⁶¹ In the presence of an antioxidant, the DPPH radical decays and the change in absorbance at 520 nm was measured by spectrophotometry. Hence the scavenging

activity of compounds was determined. The solutions of concentrations of 4, 8, 16 and 20 μ M of compounds, **4a-4g**, in 50% methanol were prepared. To this 140 μ L of 1 mM DPPH were added and then incubated at 37 °C for 30 min. The absorbance's for these solutions were measured at 520 nm against 50% methanol as blank by UV-Vis, spectrophotometer. A control reaction was also carried out without addition of the test sample. The actual absorbance was taken as the difference in the absorbance of the control and the test samples and hence IC₅₀ (inhibitory concentration 50) values were determined.

Supplementary Information

Supplementary data (NMR, MS and FTIR spectra) are available free of charge at http://jbcs.sbq.org.br as PDF file.

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References

- Engman, L.; Persson, J.; Vessman, K.; Ekstrom, M.; Berglund, M.; Anderson, C. M.; *Free Radical Biol. Med.* **1995**, *19*, 441.
- Halliwell, B.; Gutteridge, J. M. C.; *Free Radicals in Biology* and Medicine, 2nd ed.; Clarendon Press: Oxford, UK, 1989.
- 3. Andersen, J. K.; Nat. Rev. Neurosci. 2004, 5, S18.
- Seet, R. C. S.; Lee, C.-Y. J.; Lima, E. C. H.; Tan, J. J. H.; Quek, A. M. L.; Chong, W. L.; Looi, W. F.; Huang, S. H.; Wang, H.; Chand, Y. H.; Halliwell, B.; *Free Radical. Biol. Med.* **2010**, *48*, 560.
- Shohat, M.; Mimouni, D.; Ben-Amitai, D.; Sredni, B.; Sredni, D.; Shohat, B.; *Clin. Exp. Dermatol.* 2005, *30*, 432.
- Halliwell, B.; Free Radicals and Other Reactive Species in Disease; John Wiley & Sons: Chichester, 2015.
- Spallholz, J. E.; Boylan, L. M. In *Peroxidases in Chemistry and Biology*, Vol. I; Everse, J.; Everse, K. E.; Grisham, M. B., eds.; CRC Press: Boca Raton, 1991.
- Halliwel, B.; Gutteridge, J. M. C. In *Free Radicals in Biology* and *Medicine*, 4th ed.; Oxford University Press: Oxford, 2007.
- Gerzson, M. F. B.; Victoria, F. N.; Radatz, C. S.; Gomes, M. G.; Boeira, S. P.; Jacob, R. G.; Alves, D.; Jesse, C. R.; Savegnago, L.; *Pharmacol., Biochem. Behav.* **2012**, *102*, 21.
- Savegnago, L.; Vieira, A. I.; Seus, N.; Goldani, B. S.; Castro, M. R.; Lenardão, E. J.; Alves, D.; *Tetrahedron Lett.* **2013**, *54*, 40.
- Engman, L.; Laws, M. J.; Malmstrom, J.; Schiesser, C. H.; Zugaro, L. M.; *J. Org. Chem.* **1999**, *64*, 6764.
- 12. Nogueira, C. W.; Rocha, J. B. T. In Patai's Chemistry of

Functional Groups; Rappoport, Z., ed.; Wiley: Chichester, 2011.

- Manjare, S. T.; Kim, Y.; Churchill, D. G.; Acc. Chem. Res. 2014, 47, 2985.
- Nobre, P. C.; Borges, E. L.; Silva, C. M.; Casaril, A. M.; Martinez, D. M.; Lenardão, E. J.; Alves, D.; Savegnago, L.; Perin, G.; *Bioorg. Med. Chem.* **2014**, *22*, 6242.
- 15. Sies, H.; Arteel, G. E.; Free Radical. Biol. Med. 2000, 28, 1451.
- Nogueira, C. W.; Rocha, J. B. T.; J. Braz. Chem. Soc. 2010, 21, 2055.
- Engman, L.; Stern, D.; Frisell, H.; Vessman, K.; Berglund, M.; Ek, B.; Andersson, C. M.; *Bioorg. Med. Chem.* **1995**, *3*, 1255.
- Vessman, K.; Ekstram, M.; Berglund, M.; Andersson, C. M.; Engman, L.; *J. Org. Chem.* **1995**, *60*, 4461.
- Mugesh, G.; Panda, A.; Kumar, S.; Apte, S. D.; Singh, H. B.; Butcher, R. J.; *Organometallics* 2002, *21*, 884.
- Mugesh, G.; Mont, W.-W.; Sies, H.; Chem. Rev. 2001, 101, 2125.
- Zimmerman, M. T.; Bayse, C. A.; Ramoutar, R. R.; Brumaghim, J. L.; *J. Inorg. Biochem.* 2015, 145, 30.
- Nogueira, C. W.; Zeni, G.; Rocha, J. B. T.; *Chem. Rev.* 2004, 104, 6255.
- Alberto, E. E.; Nascimento, V.; Braga, A. L.; J. Braz. Chem. Soc. 2010, 21, 2032.
- Rocha, J. B. T.; Saraiva, R. A.; Garcia, S. C.; Gravina, F. S.; Nogueira, C. W.; *Toxicol. Res.* 2012, *1*, 85.
- 25. Jacobs, J. L.; The Year in Immunology; 1989, 4, 276.
- Victoria, F. N.; Radatz, C. S.; Sachini, M.; Jacob, R. G.; Alves, D.; Savegnago, L.; Perin, G.; Motta, A. S.; Silva, W. P.; Lenardão, E.; *Food Control* 2012, 23, 95.
- Kanski, J.; Drake, J.; Aksenova, M.; Engman, L.; Butterfield, D. A.; *Brain Res.* 2001, *911*, 12.
- Engman, L.; Stem, D.; Cotgreave, I. A.; Andersson, C. M.; J. Am. Chem. Soc. 1992, 114, 9737.
- Engman, L.; Stem, D.; Pelcman, M.; Andersson, C. M.; J. Org. Chem. 1994, 59, 1973.
- Andersson, C. M.; Brattsand, R.; Hallberg, A.; Engman, L.; Persson, J.; Moldeus, I. P.; Cotgreave, A.; *Free Radical Res.* 1994, 20, 401.
- Morya, V. K.; Shin, J. D.; Eun-ki, K.; *Appl. Biochem. Biotechnol.* 2014, 172, 3390.
- Ba, L. A.; Döring, M.; Jamier, V.; Jacob, C.; Org. Biomol. Chem. 2010, 8, 4203.
- Giles, G. I.; Tasker, K. M.; Johnson, R. J. K.; Jacob, C.; Peers, C.; Green, K. N.; *Chem. Commun.* 2001, 2490.
- Młochowski, J.; Lisiak, R.; Halina, W. M. In: PATAI'S Chemistry of Functional Groups, The Chemistry of Organic Selenium and Tellurium Compounds; Rappoport, Z., ed.; John Wiley & Sons: Chichester, 2012.
- Cotgreave, L. A.; Moldus, P.; Engman, L.; Hallberg, A.; Biochem. Pharmacol. 1991, 42, 1481.
- 36. Sies, H.; Exp. Physiol. 1997, 82, 291.

- Sies, H.; Jones, D. In *Encyclopedia of Stress*, Vol. 3, 2nd ed.; Fink, G., ed.; Elsevier: Amsterdam, 2007, pp 45-48.
- 38. Halliwel, B.; Plant Physiol. 2006, 141, 312.
- Poorna, C. A.; Resmi, M. S.; Soniya, E. V.; *Int. J. Agr. Chem.* 2012, 1, 1.
- Donato, F.; Gomes, M. G.; Goes, A. T. R.; Seus, N.; Alves, D.; Jesse, C. R.; Savegnago, L.; *Life Sci.* 2013, *93*, 393.
- Rooseboom, M.; Vermeulen, N. P. E.; Durgut, F.; Commandeur, J. N. M.; *Chem. Res. Toxicol.* **2002**, *15*, 1610.
- Andersson, C. M.; Hallberg, A.; Brattsand, R.; Cotgreave, I. A.; Engman, L.; Persson, J.; *Bioorg. Med. Chem. Lett.* **1993**, *3*, 2553.
- Romano, B.; Plano, D.; Encío, I.; Palop, J. A.; Sanmartín, C.; Bioorg. Med. Chem. 2015, 23, 1716.
- 44. Nogueira, C. W.; Rocha, J. B. T.; Arch. Toxicol. 2011, 85, 1313.
- Romano, B.; Font, M.; Encío, I.; Palop, J. A.; Sanmartín, C.; *Eur. J. Med. Chem.* 2014, 83, 674.
- Sanmartína, C.; Garmendia, I.; Romano, B.; Díaz, M.; Palop, J. A.; Goicoecheac, N.; *Sci. Hortic.* **2014**, *180*, 40.
- 47. Bhabak, K. P.; Mugesh, G.; Chem. Eur. J. 2008, 14, 8640.
- Sudha, N.; Singh, H. B.; Coord. Chem. Rev. 1994, 135-136, 469.
- 49. Mugesh, G.; Singh, H. B.; Acc. Chem. Res. 2002, 35, 226.
- Mukherjee, A. J.; Zade, S. S.; Singh, H. B.; Sunoj, R. B.; *Chem. Rev.* 2010, *110*, 4357.
- Asakura, T.; Shibutani, Y.; Reilly, M. P.; de Meio, R. H.; *Blood* 1984, 64, 305.
- Nakamoto, K.; Infrared and Raman Spectra of Inorganic and Coordination Compound, 3rd ed.; John Wiley & Sons: Hoboken, New Jersey, 1978.
- Kumar, P. R.; Upreti, S.; Singh, A. K.; *Inorg. Chim. Acta* 2008, 361, 1426.
- Vargas, F.; Sehnem, J. A.; Galetto, F. Z.; Braga, A. L.; *Tetrahedron* 2008, 64, 392.
- Kumar, P. R.; Singh, A. K.; Butcher, R. J.; Sharma, P.; Toscano, R. A.; *Eur. J. Inorg. Chem.* **2004**, 1107.
- Singh, A. K.; Srivastava, V.; Phosphorus, Sulfur Silicon Relat. Elem. 1990, 47, 471.
- Suresh, D.; Udayabhanu, Nethravathi, P. C.; Lingaraju, K.; Rajanaika, H.; Sharma, S. C.; Nagabhushana, H.; *Spectrochim. Acta, Part A* 2015, *136*, 1467.
- 58. Karamac, M.; Amarowicz, R.; Grasas Aceites 1997, 48, 83.
- Irgolic, K. J.; Zingaro, R. A.; *Reactions of Organotellurium Compounds in Organometallic Synthesis*, Vol. 2; Becker, E.; Tsutsui, M., eds.; John Wiley & Sons: New York, NY, USA, 1971.
- 60. Brand-Williams, W.; Food Sci. Technol. Int. 1995, 28, 25.
- You, Y.; Ahsan, K.; Detty, M. R.; J. Am. Chem. Soc. 2003, 125, 4918.