CHEMICAL COMPOSITION AND ANTIMICROBIAL ACTIVITY OF THE ESSENTIAL OIL FROM Aeolanthus suaveolens MART. ex Spreng

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The essential oils from leaves (sample A) and flowers (sample B) of *Aeolanthus suaveolens* Mart. ex Spreng were obtained by hydrodistillation and analyzed by GC, GC-MS, and chiral phase gas chromatography (CPGC). Six compounds have been identified from the essential oils, representing *ca* 94.3 and 93% of the oils corresponding to samples A and B, respectively. The major constituents of samples A and B essential oils were respectively, linalool (34.2%/34.9%), (-)-massoialactone (25.9%/17.0%) and (E)- β -farnesene (25.4%/29.1%). The enantiomeric distribution of the monoterpene linalool was established by analysis on heptakis-(6-O-methyl-2,3-di-O-pentyl)- β -cyclodextrin capillary column. The antimicrobial activity of the essential oil from leaves and isolated compounds was also evaluated.

Keywords: Aeolanthus suaveolens; essential oils; antimicrobial activity.

INTRODUCTION

The Lamiaceae is one of the most diverse and widespread plant families in terms of ethnomedicine. The medicinal value of Lamiaceae species is largely based on their volatile oils¹. Regarding its effects on the central nervous system (SNC), studies proved anticonvulsant activities in at least three genera, among which *Aeolanthus*², *Leonurus*³ and *Salvia*⁴ are represented in the state of Rio Grande do Sul (RS), Brazil.

Among many essential oils, those from plants within the Lamiaceae family have received considerable attention in the search for biologically active natural products against agricultural as well as stored products pests. In a recent study, was investigated the antifeedant properties of several Lamiaceae essential oils and some of their volatile constituents against onion thrips *Thrips tabaci*. This raised questions about possible other biological effects of these phytochemicals in the chemical ecology of the onion thrips⁵⁻⁷.

Aeolanthus suaveolens Mart. ex Spreng known as "Catinga de Mulata", is a widespread plant in the Northeastern region of Brazil (Brazilian Amazon), and belongs to the Lamiaceae family. It is commonly used in folk medicine because of its sedative and anticonvulsant effects. Earlier reports have demonstrated that this specie presents the monoterpene, linalool, which is the major component of its essential oil and possibly responsible for the plant anticonvulsant properties^{8,9}.

In the present work, the chemical composition of the essential oils from leaves and flowers and antimicrobial activity of the essential oil from leaves were further investigated together with some of isolated compounds from essential oil of *Aeolanthus suaveolens* collected in Rio Grande do Sul. In addition, we report on the enantiomeric distribution of the chiral monoterpene constituent linalool in essential oils of *A. suaveolens*.

EXPERIMENTAL

Plant material

Aeolanthus suaveolens was collected at the town of São Vicente do Sul, Rio Grande do Sul state, Brazil. Leaves and flowers were collected between November 2003 and September 2004. Voucher specimens (FOET-0504) have been deposited at Herbarium of Federal University of Santa Maria. The leaves of *Ocimum basilicum* were collected at the town of Santa Maria, RS.

General

¹H⁻ and ¹³C -NMR spectra were recorded on a Bruker DPX 400 (400.1/100.6 MHz) NMR spectrometer, in CDCl₃ with TMS as internal standard. TLC were performed on precoated silica gel 60 F254 plates (Merk) and detection was achieved by UV light (254 nm) and by spraying with solutions of 10% H_2SO_4 and 1% vanillin in ethanol/ H_2SO_4 , followed by heating.

Chemical analysis

Fresh leaves and flowers (sample A and B) of *Aeolanthus suaveolens* and leaves of *Ocimum basilicum*, were subjected to hydrodistillation for 4 h using a modified Clevenger-type apparatus, and followed by exhaustive extraction of the distillate with diethyl ether.

The oils were analyzed by GC and GC/MS. GC analyses were performed using a Varian CP-3800 gas chromatograph with a FID detector and SE-54 fused silica column (25 m x 0.25 mm, film thickness 0.2 μ m). Operation conditions were as follows: injector and detector temperatures, 220 and 280 °C, respectively; carrier gas, H₂; oven temperature program from 50 - 250 °C at 4 °C/min. GC-MS analyses were performed using a Varian model 3800 Saturn system operating in the EI mode at 70 eV equipped with a CP-SIL cross-linked capillary column (30 m x 0.25 mm, film thickness 0.2 μ m). The identity of the oil components was based on comparison

of the retention times and Kovat's retention indexes and mass spectra with those of NBS/NIST Library and those described by Adams¹⁰, and by co-injection with standards available in our laboratories.

The enantiomeric distribution of chiral monoterpene linalool in essential oils of *A. suaveolens* and *O. basilicum* was determined by peak enrichment by enantioselective capillary GC with fused capillary column, 25 m x 0.25 mm, coated with heptakis-(6-O-methyl-2,3-di-O-pentyl)- β -cyclodextrin diluted with polysiloxane OV-1701. A Varian-3800 apparatus equipped with a flame ionization detector (FID) was used with hydrogen as the carrier gas. All runs were performed with the temperature program from 40 to 180 °C at 3 °C/min.

Isolation procedure

Part of the resulting oils of leaves from *Aeolanthus suaveolens* (100 mg) was further subjected to preparative TLC (SiO₂; hexane-EtOAc, 90:15) and afforded massoialactone (16 mg), β -farnesene (18 mg), and linalool (15 mg), respectively (Figure 1). The authenticity of linalool was determined by direct comparison with authentic samples in GC. Massoialactone and β -farnesene were characterized by GC/MS and NMR data. The absolute configuration of massoialactone [(*R*)-isomer¹¹] was determined by optical rotation.

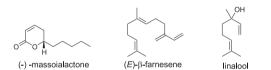


Figure 1. Structures of the compounds isolated from essential oil of Aeolanthus suaveolens

(-)-massoialactone: $[\alpha]_{\rm D}$: -91 (in CDCl₃, c = 0.006); ¹H NMR (400 MHz, CDCl₃): δ 0.82 (brs, 3H), 1.24-1.93 (m, 8H), 2.23-2.29 (m, 2H), 4.32-4.40 (m, 1H), 5.93-5.97 (m, 1H), 6.78-6.83 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.9, 20.7, 24.4, 29.4, 31.5, 34.8, 78.0, 121.4, 145.0, 164.5.

(*E*)-β-farnesene: ¹H NMR (400 MHz, CDCl₃): δ 1.58 (s, 6H), 1.66 (s, 3H), 1.96-2.23 (m, 8H), 4.97-5.24 (m, 6H), 6.36 (dd, 1H, J_1 10 Hz, J_2 17 Hz). ¹³C NMR (100 MHz, CDCl₃) δ 16.0, 17.6, 25.7, 26.6, 26.7, 31.4, 39.7, 113.0, 115.7, 124.0, 124.3, 131.3, 135.4, 139.0, 146.1.

Antimicrobial bioassay

The antimicrobial activity of sample A was assayed using the Minimal Inhibitory Concentration (MIC). A collection of eleven microorganisms was used, including seven bacteria: Staphylococcus aureus (ATCC 6538p), Staphylococcus epidermidis (ATCC 12228), Bacillus subitilis (ATCC 6633), Klebsiella pneumoniae (ATCC 10031), Eschericchia coli (ATCC 11103), Pseudomonas aeruginosa (ATCC 27873), Salmonella setubal (ATCC 19196); and four yeasts: Candida albicans (ATCC 10231), Saccharomyces cerevisae (ATCC 2601), Cryptococcus neoformans (ATCC 28952) and Candida dublinensis (SM26) (Candida dublinensis was isolated from patients of the University Hospital of Santa Maria). Standard microorganism strains were obtained from American Type Culture Collection (ATCC), and standard antibiotics chloramphenicol and nistatine were used in order to control microbial test sensitivity¹². The Minimal Inhibitory Concentration (MIC) was determined on 96well culture plates by a micro dilution method using a microorganism suspension with a density of 105 CFU/mL in Casein Soy Broth (CSB) incubated for 24 h at 37 °C for bacteria, and Sabouraud Broth (SB) incubated for 72 h at 25 °C for yeasts. The

cultures that did not present growth were used to inoculate solid medium plates (Muller Hinton Agar and Sabouraud Agar) in order to determine the Minimal Lethal Concentration (MLC). Proper blanks were assayed simultaneously and samples were tested in triplicate. Technical data was described previously¹³⁻¹⁵.

Bioautographic bioassay

The antimicrobial activity of isolated compounds (linalool, β -fanesene and (-)-massoialactone) and crude essential oil (sample A) were assayed using the bioautography technique¹⁶⁻¹⁸ with a collection of six microorganisms including Staphylococcus aureus, Staphylococcus epidermidis, Bacillus subitilis, Eschericchia coli, Pseudomonas aeruginosa and Salmonella setubal. Compound concentrations from 1.56 to 100 µg were applied to precoated TLC plates. Muller Hinton Agar and Sabouraud Agar media inoculated with microorganisms suspended in saline solution (10⁵ CFU/mL) were distributed over TLC plates. Bacterium plates were incubated for 24 h at 37 °C. Chloramphenicol was used in order to control microbial test sensitivity¹⁴. The results were stained with an aqueous solution of 2,3,5-triphenyl-tetrazolium chloride (TTC, 1 mg/mL). The appearance of inhibition zones was used to determine the lowest sample amount capable of inhibiting microbial growth. Samples were tested in triplicate.

RESULTS AND DISCUSSION

The content of essential oils from *Aeolanthus suaveolens* expressed in percentage was as follows: leaves 0.8% and flowers 0.5%.

The essential oils of *Aeolanthus suaveolens* obtained from leaves and flowers were analyzed by GC and GC-MS to determine their main constituents. Six compounds were identified, making up 94.3% of the oil from leaves (Sample A) and 92.6% of the oil from flowers (Sample B). Table 1 lists the chemical components of the essential oils and differences between sample A and the sample B. The main constituents of the essential oils were linalool (34.2%/34.9%), (-)-massoialactone (25.9%/17.0%) and β -farnesene (25.4%/29.1%) in samples A and B respectively. Others compounds found in the oils were linalool acetate, α -santalene and α -bergamotene.

 Table 1. Percentage composition of the essential oils of Aeolanthus suaveolens Mart. ex Spreng

Compounds ^a	Leaves Sample A	Flowers Sample B	RI ^b	Identification ^c
(-)-linalool	22.5	21.6	1098	GC-MS, co
(+)-linalool	12	14.3	1098	GC-MS, co
linalool acetate	5.7	6.8	1255	GS-MS
α -santalene	1.9	2.8	1418	GC-MS
α-bergamotene	0.9	1.4	1414	GC-MS
(E) - β -farnesene	25.4	29.1	1458	NMR, GC-MS
(-)-massoialactone	25.9	17.0	1476	NMR, GC-MS
TOTAL	94.3	93.0		

^aCompounds listed in order of elution from a SE-54 column; ^bRetention indices determined on apolar SE-54 column (50-250 °C; 4 °C min⁻¹); ^eIdentification: RI, GC-MS, gas chromatography-mass spectroscopy; NMR, nuclear magnetic ressonance.

The enantiomeric composition of the chiral monoterpene linalool was investigated, resulting in the detection of an enantiomeric distribution of the (-)-linalool (22.5%)/(+)-linalool (12%) in oil from leaves and (-)-linalool (21.6%)/(+)-linalool (14.3%) in oil from flowers. In accordance with reports in the

literature^{19,20}, the monoterpene linalool can to bear a racemisation process during hydrodistillation. For that reason, was investigated the enantiomeric composition of the linalool in essential oil obtained from *Ocimum basilicum* (manjericão) (a specie well known by to contain high content of linalool). Through this analysis was observed the enantiomerically pure presence of the isomer (-)-linalool in the essential oil of *O. basilicum*. So that, was concluded that the presence of the enantiomeric mixture of the linalool in the essential of *A. suaveolens* is natural and not due to an artifact formation.

The antimicrobial activity of the oil from leaves of A. suaveolens was evaluated by determining of the minimal inhibitory concentration (MIC). The results are given in Table 2. The oil was active against all microorganisms tested; being that the best MIC value observed was 1.25 mg/mL for Escherichia coli and Cryptococcus neoformans. The antimicrobial activity of the essential oil from leaves and isolated compounds (linalool, β-farnesene and massoialactone) was also tested by bioautography techique. According to results showed in Table 3 was observed that the (-)-massoialactone is the more active component and appear to be the principal responsible for biological properties of the oil. This compound exhibited a more potent antibacterial activity, when compared with others isolated compounds (linalool and β-farnesene) and crude essential oil. (-)-Massoialactone exhibited antibacterial activity against S. setubal (3.12 µg), B. subtilis (3.12 µg), S. aureus (6.25 µg), S. epidermidis (6.25 µg), E. coli (6.25 µg) and P. *aeruginosa* (6.25 μ g). The sesquiterpene β -farmesene showed activity against four microorganisms and the monoterpene linalool was active only against S. aureus (100 µg). In spite with (-)-massoialactone, this compound has been previously isolated from the bark oil of Critocarya massoia11 and jasmine flowers21, is also found in the defense secretion of two species of formicin ants of the genus Camponotus²².

In conclusion, the present study indicated that the leaf/flower essential oils of *Aeolanthus suaveolens* possessed a high content of the monoterpenes (66.1%/59.7%) and sesquiterpene (28.2%/33.3%), respectively. Linalool, β -farnesene and (-)-massoialactone were the isolated substances that are presented in the essential oils. The antimicrobial activity of these compounds was evaluated by bioautography technique, which showed the remarkable antimicrobial

Table 2. Antimicrobial activity of the essential oil from leaves of the *Aeolanthus suaveolens* (minimal inhibitory concentration (MIC) and the minimal letal concentration (MLC) in mg mL⁻¹)

Microorganisms	Leaves Sample A		Standard ^{a,b}	
	CIM ^a	CLM ^a	CIM	
Staphylococcus aureus	10	>20	6.25x 10 ⁻³	
Staphylococcus epidermidis	10	>20	3.12x10 ⁻³	
Escherichia coli	1.25	5	1.67x10 ⁻³	
Pseudomonas aeruginosa	5	>20	3.12x10 ⁻³	
Klebsiella pneumoniae	10	>20	6.25x 10 ⁻³	
Bacillus subtilis	10	>20	3.12x10 ⁻³	
Salmonella setubal	10	>20	1.67x10 ⁻³	
Cryptococcus neoformans	1.25	2.5	4.06x10 ⁻³	
Saccharomyces cereviseae	2.5	5	8.12x10 ⁻³	
Candida albicans	5	10	8.12x10 ⁻³	
Candida dublinensis	2.5	10	8.12x10 ⁻³	

^a Mean of 3 replicates in mg mL⁻¹. ^bStandard antimicrobial agents: chloramphenicol against bacteria and nistatine against yeasts.

potential of the (-)-massoialactone. The crude essential oil from *A. suaveolens* also exhibited a good antimicrobial activity.

According the found results, it can be concluded that the essential oil of *Aeolanthus suaveolens*, from leaves and flowers, are similar in terms of the chemical composition. The presence of an enantiomerically mixture of the linalool in essential oil of *A. suaveolens* is a natural occurrence, and is not due to an artifact formation during hydrodistillation process. The component (-)-massoialactone appear to be the main responsible for the biological properties of the oil.

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Table 3. Antimicrobial activity: minimum amount required, for essential oil and isolated compounds, for inhibition of bacterial growth on TLC plates (μg)

Microorganisms	Essential oil (leaves)	(E) - β -farnesene	linalool	massoialactone	Standard ^a
S. aureus	50	50	100	6.25	0.7
S. epidermidis	-	-	-	6.25	0.7
E.coli	50	-	-	6.25	0.4
S. setubal	50	50	-	3.125	0.6
P. aeruginosa	50	100	-	6.25	0.6
B. subtilis	25	12.5	-	3.125	0.6

^aStandard antibiotic chloramphenicol for bacteria.

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