

## BROMOPYRROLE ALKALOIDS FROM THE CARIBBEAN SPONGE *Agelas cerebrum*

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Bioguided fractionation of *Agelas cerebrum* crude extract resulted in isolation of four bromopyrrole and four bromopyrrole aminoimidazole alkaloids, identified as 5-bromopyrrole-2-carboxylic acid (**1**), 4-bromopyrrole-2-carboxylic acid (**2**), 3,4-bromopyrrole-2-carboxylic acid (**3**), 4,5-bromopyrrole-2-carboxylic acid (**4**), oroidin (**5**), bromoageliferin (**6**), dibromoageliferin (**7**) and dibromosceptrin (**8**) on the basis of spectroscopic data analyses (UV, IR, HRMS, 1D and 2D NMR) and comparison with literature data. This is the first report of compounds **2** and **3** in a marine sponge belonging to the *Agelas* genus and the first evidence of the presence of **1** from a natural source.

Keywords: *Agelas cerebrum*; bromopyrrole alkaloids; antitumoral and antiprotozoal activity.

## INTRODUCTION

Interest in the biology and chemistry of sponges that belong to the genus *Agelas* continues despite many years of study devoted to these species. *Agelas* sponges are commonly found on the Caribbean and Indo-Pacific coral reefs, and new species are reported every year. For example, in 1992 it was noted that 12 species of *Agelas* were documented,<sup>1</sup> while today the taxonomic record shows that there are 35 such species,<sup>2</sup> from which more than 200 novel molecules have been isolated,<sup>3</sup> some of them with high therapeutic potential.<sup>4</sup>

Characteristic metabolites from this genus of sponges are bromopyrrole alkaloids which present a broad range of biological activities,<sup>5</sup> including anticancer<sup>6</sup> and antimalarial activity.<sup>7</sup> Structurally, most of them are built up by a 4-bromo- or 4,5-dibromopyrrole-2-carboxylic acid moiety, which is often connected with a 2-aminoimidazole group through an aliphatic segment. Monomers of this chemotype can dimerize or trimerize to give a large variety of pyrrole 2-aminoimidazole alkaloids.

As part of our ongoing work on secondary metabolites produced by Caribbean marine sponges, we undertook the chemical study of the marine sponge *Agelas cerebrum* Assmann, van Soest & Köck, 2001 (phylum Porifera, class Demospongiae, order Agelasida, family Agelasidae). This is the first report of a chemical study on the isolation and identification of metabolites from the marine sponge *A. cerebrum*.

## EXPERIMENTAL

### General procedures

UV measurements were performed on a Varian Cary 300 Scan

UV-visible spectrophotometer. IR spectra were obtained with a PerkinElmer Paragon 1000 FT-IR spectrophotometer. NMR experiments were performed on a Bruker Avance 500 MHz spectrometer. Low resolution electrospray ionization (ESI) mass spectra were obtained with a Bruker Esquire 3000 Plus spectrometer in the positive or negative mode. High resolution mass spectra (HRESIMS) were conducted on a LTQ Orbitrap mass spectrometer (Thermo Finnigan). HPLC purification was carried out on a Waters 600 system equipped with a Waters 717 plus autosampler, a Waters 996 photodiode array detector, and a Sedex 55 evaporative light-scattering detector (Sedere, France).

### Biological material

A specimen of the marine sponge *A. cerebrum* was collected at a depth of about 20 m from "Boca de Calderas", Havana, Cuba (23° 05' 55" N 82° 28' 30" W) in March 2008 and identified by Dr. P. M. Alcolado (Institute of Oceanology, Havana, Cuba). A voucher sample (ANC.02.010) has been deposited in the sponge collection of the Cuban National Aquarium. The sponge was kept frozen from collection until the extraction process.

### Extraction and isolation

A portion of *A. cerebrum* (250 g) was freeze-dried and ground to obtain a dry powder (15 g), which was exhaustively extracted with a mixture of MeOH/CH<sub>2</sub>Cl<sub>2</sub> (1:1) to give 2.3 g of a crude extract after concentration under reduced pressure. The crude extract was fractionated by RP-C<sub>18</sub> flash chromatography (elution with a decreasing polarity gradient of H<sub>2</sub>O/MeOH from 1:0 to 0:1, then MeOH/CH<sub>2</sub>Cl<sub>2</sub> from 1:0 to 0:1). Fractions obtained were submitted to antitumoral assays by a colorimetric high-throughput screen.

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The bioactive H<sub>2</sub>O/MeOH (1:3) fraction (180 mg) was chosen for a chemical investigation and was further fractionated by RP-C<sub>18</sub> semi-preparative HPLC (Phenomenex, Luna C<sub>18</sub>, 250 × 10 mm, 5 μm) with a gradient of H<sub>2</sub>O/MeOH/Formic acid from 60:40:0.1 to 20:80:0.1 in 35 min (flow 3.0 mL/min) and the subsequent mixtures were finally purified by RP-C<sub>18</sub> analytical HPLC (Phenomenex Luna C<sub>18</sub>, 150 × 4.6 mm, 5 μm, flow 1.0 mL/min) to afford a new compound (**1**, 1.2 mg), together with seven known metabolites: **2** (5.2 mg), **3** (1.0 mg), **4** (2.5 mg), **5** (3.6 mg), **6** (2.7 mg), **7** (1.8 mg) and **8** (1.4 mg). Due to higher yield requirements for antimalarial assays, a preparative fraction was further obtained by RP-C<sub>18</sub> chromatography after stepwise elution with H<sub>2</sub>O (which was discarded) and MeOH/CH<sub>2</sub>Cl<sub>2</sub> (1:1).

**Compound 1**: Amorphous white solid; UV (MeOH) λ<sub>max</sub> (log ε) 234 (3.80), 273 (4.10) nm; IR (thin film) ν<sub>max</sub> 3356, 3122, 1649 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD + CDCl<sub>3</sub>) δ 6.26 (1H, d, *J* = 3.5 Hz, H-4), 6.59 (1H, d, *J* = 3.5 Hz, H-3); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD+CDCl<sub>3</sub>) δ 129.3 (C-2), 111.7 (C-3), 112.3 (C-4), 104.3 (C-5), 164.1 (C-6); HRESI-MS (-): *m/z* 190, 188 [M-H]<sup>-</sup> (calcd for C<sub>5</sub>H<sub>3</sub><sup>79</sup>BrNO<sub>2</sub>, 187.9396, Δ 0.3 ppm).

#### Antitumoral assay

A colorimetric assay using sulforhodamine B was adapted for a quantitative measurement of cell growth and viability following a technique described in the literature.<sup>8</sup> *In Vitro* cytotoxicity was evaluated against three tumor cell lines: lung carcinoma A549, colon carcinoma HT29, and breast MDA-MB-231 and samples were tested at concentrations of 100, 10 and 1 μg/mL.

#### Antimalarial assay

*In vitro* drug susceptibility was determined in standard short-term cultures of *Plasmodium berghei* ANKA blood stages, as described before.<sup>9</sup> Briefly, erythrocytes infected with parasites of *P. berghei* ring forms/young trophozoites were incubated at 2% parasitemia at a final cell concentration of 1% in complete culture medium (RPMI 1640; 20% Fetal Calf Serum, Sigma) containing serial dilutions of samples from *A. cerebrum*, each in duplicate wells of 96-well culture plates. These plates were incubated for a period of 24 h at 37 °C under standardized *in vitro* culture conditions. The antimalarial activity was expressed as IC<sub>50</sub>, which was determined according to reported methodology<sup>10</sup> using data of inhibition of schizont maturation measured as described by Schlichtherle *et al.*,<sup>11</sup> and adapting recommendations for *P. falciparum* isolates.<sup>12</sup> Chloroquine phosphate and artemisinin (both from Sigma) were used as references.

## RESULTS AND DISCUSSION

Compound **1** was isolated as amorphous white solid and its mass spectrum showed molecular ions at *m/z* 188 and 190 in a 1:1 ratio, indicative of the presence of one bromine atom. The molecular formula was determined as C<sub>5</sub>H<sub>4</sub>NO<sub>2</sub>Br by HRESI-MS. The UV absorption [λ<sub>max</sub> 273 nm (log ε 4.10)] was attributed to a substituted pyrrole chromophore.<sup>13</sup> The bands at 3356, 3122, and 1649 cm<sup>-1</sup> in the IR spectrum suggested the presence of amine and carbonyl moieties. In the <sup>13</sup>C NMR spectrum, resonances due to a disubstituted pyrrole at δ 104.3 (s), 112.3 (d), 111.7 (d), and 129.3 (s) were observed as well as a carbonyl (COOH) at δ 164.1 (s). The <sup>1</sup>H NMR spectrum indicated resonances due to 2,5-disubstituted pyrrole protons at δ 6.26 (1H, d, *J* = 3.5 Hz, H-4) and 6.59 (1H, d, *J* = 3.5 Hz, H-3). The positions of the CO<sub>2</sub>H moiety at C-2 and the bromine atom at C-5 were in agreement with other 2,5-disubstituted pyrroles.<sup>14</sup> On this basis, compound **1** was concluded to be 5-bromopyrrole-2-carboxylic acid. On the other hand, compounds **2-8** were identified by a combination of spectroscopic methods (<sup>1</sup>H, <sup>13</sup>C 1D and 2D NMR, ESIMS) and comparison with the literature data.

Bromopyrrole alkaloids are known to be one of the most common metabolites contained in marine sponges<sup>15</sup> and they are widely distributed in the species belonging to the genera *Agelas*, *Axinella*, *Acanthella*, *Pseudoaxinyssa*, and *Hymeniacion*.<sup>16</sup> Oroidin (**5**), the first member of pyrrole-2-aminoimidazole alkaloids in this group, was isolated from the sponge *A. oroides*.<sup>17</sup> However, a revised structure was soon proposed.<sup>18</sup> It was not until 1981, during solidstate photodimerization studies of (-)-sceptrin, a related dimeric bromopyrrole alkaloid, that the final structure of **5** was confirmed by X-ray diffraction analysis.<sup>19</sup> Oroidin has been isolated from many *Agelas* sponges and species of other genera such as: *Axinella damicornis*, *Axinella verrucosa*, *Acanthella aurantiaca*, *Goreauiella sp* and *Pseudaxinyssa cantharella*.<sup>20</sup> Compound **4** (4,5-bromopyrrole-2-carboxylic acid) has been also isolated from many *Agelas* species and together with oroidin have been reported to exhibit significant biological properties.<sup>21</sup> Chanas *et al.*<sup>22</sup> suggested that *A. conifera*, *A. dispar*, *A. inaequalis*, *A. sceptrum* and *A. wiedenmmeri* share both metabolites as a common chemical defense against fish predators.

Bromoageliferin (**6**) and dibromoageliferin (**7**), dimers of oroidin, were isolated from *A. conifera* and *A. cf. mauritiana*.<sup>23</sup> These metabolites are potent actomyosin ATPase activators<sup>24</sup> and other significant biological effects have been reported.<sup>25</sup> Since then, these compounds have been isolated from many *Agelas* species and from sponges of other genera such as *Astroscera willeyana*<sup>26</sup> and *Stylissa caribica*.<sup>27</sup> Dibromosceptrin (**8**), another dimeric bioactive metabolite, belonging to the sceptrin family, was discovered in *A. conifera*.<sup>20</sup> This compound, as well as **6** and **7**, were found to be potent feeding deterrents.<sup>28</sup>

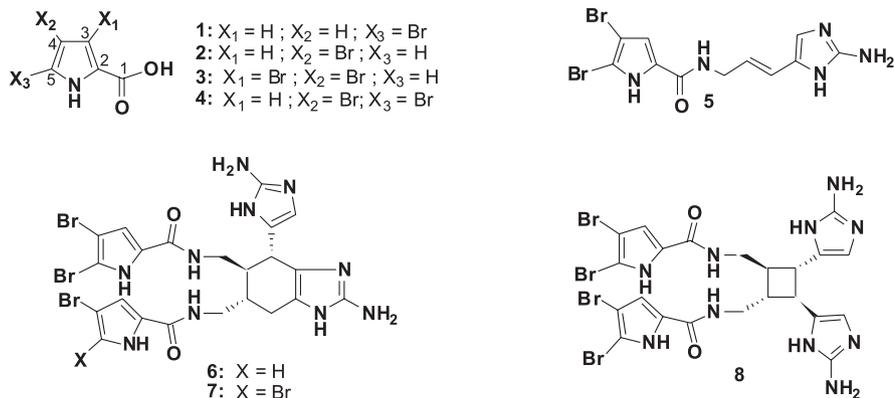


Figure 1. Structures of compounds **1-8** isolated from *A. cerebrum*

4-Bromopyrrole-2-carboxylic acid (**2**) and 3,4-bromopyrrole-2-carboxylic acid (**3**) were recently encountered in the Mediterranean sponge *Axinella verrucosa*<sup>29</sup> and the tropical sponge *Axinella damicornis*.<sup>6</sup> However, to our knowledge, this is the first report of the occurrence of both compounds in a sponge belonging to the *Agelas* genus. While compound **1** had been previously synthesized,<sup>28</sup> it is herein reported as a new natural bromopyrrole alkaloid isolated from this species. 5-bromopyrrole alkaloids are not commonly isolated from marine sources and there are only two examples of other new 5-bromopyrrole derivatives identified in the genus *Agelas*.<sup>30</sup> Then, in a sense, compound **1** may be useful as a chemotaxonomic marker for *Agelas cerebrum*.

Concerning bioactivity, the H<sub>2</sub>O/MeOH (1:3) fraction from which compounds **1-8** were isolated showed strong cytotoxic activity in an *in vitro* antitumoral assay against three human tumor cell lines (A549 lung cancer cells, HT29 colonic cancer cells, and MDA-MB-231 breast cancer cells) at values equal and greater than 1 µg/mL; however, no antitumor activity against the same cell lines was detected below 10 µg/mL for each isolated compound. Probably, undetectable quantities of a very potent antineoplastic substance justify the activity of the crude fraction, or the synergism of natural product mixtures.

The organic extract of *A. cerebrum* exhibited a moderate antimalarial activity, which was evaluated according to recommended endpoint criteria for natural complex mixtures,<sup>31</sup> with IC<sub>50</sub> value equal to 60,35 ± 10,6 µg x mL<sup>-1</sup> against *P. berghei*. Although this work revealed a different profile for bromopyrrole alkaloids isolated from *A. cerebrum* in comparison to that previously described for *A. oroides*, the presence of oroidin (**5**) and 4,5 dibromopyrrole-2-carboxylic acid (**4**) suggests that they are the main active principles in the antimalarial organic fraction. Both compounds were previously identified in *A. oroides* as devoid of any cytotoxicity against L6 cells, while they exhibited an IC<sub>50</sub> value of 3.9 mg/ml for **4** and a limited antimalarial activity for **5**, in the whole cell parasite assays.<sup>7</sup> Scepterin was evaluated against D6 and W2 strains by other authors and showed no activity.<sup>32</sup>

In conclusion, this chemical study underscores the presence of bromopyrrole alkaloids as the representative secondary metabolites of *Agelas cerebrum*, and contributes to the chemotaxonomy within the Agelasidae family. Studies are in progress in order to advance the evaluation of the *in vivo* antimalarial activity of bromopyrrole alkaloids from *Agelas cerebrum*.

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## REFERENCES

- Braekman, J. C.; Daloze, D.; Stoller, C.; van Soest, R. W. M.; *Biochem. Syst. Ecol.* **1992**, *20*, 417.
- van Soest, R.; Boursy-Esnault, N.; Janussen, D.; Hooper, J.; World Porifera database. In <http://www.marinespecies.org/porifera/porifera.php?p=taxlist.html>, accessed January 2010.
- Munro, M. H. G.; Blunt, J. W.; *Marine Literature DataBase (MarinLit)*, University of Canterbury, New Zealand, 2009.
- Costa-Lotufu, L. V.; Wilke, D. V.; Jimenez, P. C.; Epifanio, R. de A.; *Quim. Nova* **2009**, *32*, 703.
- Mayer, A. M.; Rodriguez, A. D.; Berlinck, R. G.; Hamann, M. T.; *Comp. Biochem. Physiol., C: Comp. Pharmacol. Toxicol.* **2007**, *145*, 553; Mayer, A. M.; Rodriguez, A. D.; Berlinck, R. G.; Hamann, M. T.; *Biochim. Biophys. Acta* **2009**, *1790*, 283.
- Hassan, W.; Elkhayat, E. S.; Edrada, R. A.; Ebel, R.; Proksch, P.; *Nat. Prod. Commun.* **2007**, *21*, 1149; Hertiani, T.; Edrada-Ebel, R.; Ortlepp, S.; van Soest, R. W. M.; de Voogd, N. J.; Wray, V.; Hentschel, U.; Kozzyska, S.; Müller, W. E. G.; Proksch, P.; *Bioorg. Med. Chem.* **2010**, *18*, 1297.
- Tasdemir, D.; Topaloglu, B.; Perozzo, R.; Brun, R.; O'Neill, R.; Carbal-leira, N. M.; Zhang, X.; Tonge, P. J.; Linden, A.; R ed. P.; *Bioorg. Med. Chem.* **2007**, *15*, 6834; Fattorusso, E.; Tagliatalata-Scafati, O.; *Mar. Drugs* **2009**, *7*, 130.
- Skehan, P.; Storeng, R.; Scudiero, D.; Monks, A.; McMahon, J.; Vistica, D.; Warren, J. T.; Bokesch, H.; Kenney, S.; Boyd, M. R.; *J. Natl. Cancer Inst.* **1990**, *82*, 1107.
- Janse, C. J.; Waters, A. P.; *Parasitol. Today* **1995**, *11*, 138.
- Huber, W.; Koella, J. C.; *Acta Trop.* **1993**, *55*, 257.
- Schlichtherle, M.; Wahlgren, M.; Perlmann, H.; Scherf, A.; *Methods in malaria research*, 3<sup>rd</sup> ed., Malaria Research and Reference Reagent Resource Center: Virginia, 2000.
- World Health Organization, Expert Committee. *In vitro micro-test (Mark III) for the assessment of the response of Plasmodium falciparum to chloroquine, mefloquine, quinine, amodiaquine, sulfadoxine/pyrimethamine and artemisinin*; CTD/MAL/97.20 Rev.2.2001.
- Araki, A.; Kubota, T.; Aoyama, K.; Mikami, Y.; Fromont, J.; Kobayashi, J.; *Org. Lett.* **2009**, *11*, 1785.
- Cimino, G.; De Rosa, S.; De Stefano, S.; Mazzarella, L.; Puliti, R.; Sodano, G.; *Tetrahedron Lett.* **1982**, *23*, 767; Nakamura, H.; Ohizumi, Y.; Kobayashi, J.; Hirata, Y.; *Tetrahedron Lett.* **1984**, *25*, 2475.
- Blunt, J. W.; Copp, B. R.; Hu, W.-P.; Munro, M. H. G.; Northcote, P. T.; Prinsep, M. R.; *Nat. Prod. Rep.* **2008**, *25*, 35.
- Faulkner, D. J.; *Nat. Prod. Rep.* **2000**, *17*, 7; Erpenbeck, D.; van Soest, R. W. M.; *Biochem. Syst. Ecol.* **2005**, *33*, 585.
- Forenza, S.; Minale, L.; Riccio, R.; Fattorusso, E.; *J. Chem. Soc., Chem. Commun.* **1971**, 1129.
- Garcia, E. E.; Benjamin, L. L.; Fryer, R. I.; *J. Chem. Soc., Chem. Commun.* **1973**, 78.
- Walker, R. P.; Faulkner, D. J.; van Engen, D.; Clardy, J.; *J. Am. Chem. Soc.* **1981**, *103*, 6772.
- Keifer, P. A.; Schwartz, R. E.; Koker, M. E. S.; Hughes, R. G., (Jr); Rittschof, D.; Rinehart, K. L.; *J. Org. Chem.* **1991**, *56*, 2965.
- Bickmeyer, U.; Drechsler, C.; Köck, M.; Assmann, M.; *Toxicol.* **2004**, *44*, 45.
- Chanas, B.; Pawlik, J. R.; Lindel, D.; Fenical, W.; *J. Exp. Mar. Biol. Ecol.* **1996**, *208*, 185.
- Rinehard, K. L.; *Pure Appl. Chem.* **1989**, *61*, 525.
- Kobayashi, J.; Tsuda, M.; *Tetrahedron* **1990**, *46*, 5579.
- Huigens, R. W.; Richards, J. J.; Parise, G.; Ballard, T. E.; Zeng, W.; Deora, R.; Melander, C.; *J. Am. Chem. Soc.* **2007**, *129*, 6966.
- Williams, D. H.; Faulkner, D. J.; *Tetrahedron* **1996**, *52*, 5381.
- Assmann, M.; van Soest, R. W. M.; Köck, M.; *J. Nat. Prod.* **2001**, *64*, 1345.
- Assmann, M.; Lichte, E.; Pawlik, J. R.; Köck, M.; *Mar. Ecol. Prog. Ser.* **2000**, *207*, 255.
- Aiello, A.; D'Esposito, M.; Fattorusso, E.; Menna, M.; Mueller, W. E. G.; Perović-Ottstadt, S.; Schröder, H. C.; *Bioorg. Med. Chem.* **2006**, *14*, 17.
- Iwagawa, T.; Kaneko, M.; Okamura, H.; Nakatani, M.; van Soest, R. W. M.; *J. Nat. Prod.* **1998**, *61*, 1310.
- Cos, P.; Vlietinck, A. J.; Vanden Berghe, D.; Maes, L.; *J. Ethnopharmacol.* **2006**, *106*, 290.
- Mohammed, R.; Peng, J.; Nelly, M.; Hamann, M. T.; *J. Nat. Prod.* **2006**, *69*, 1739.