Meiotic behavior in apomictic Brachiaria ruziziensis × B. brizantha (Poaceae) progenies

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Introduction

The extensive use of a few *Brachiaria* cultivars in the last three decades has placed Brazil as the major beef cattle exporter and the second largest beef producer in the world. However, there is a great demand for new cultivars to diversify the millions of hectares of *Brachiaria* pastures, and these must combine desirable agronomic characteristics such as those found in the two most widely used cultivars in the country: (i) the high production, nutritive value and resistance to spittlebugs (Homoptera: Cercopidae) as found in *B. brizantha* cv. Marandu, and (ii) the tolerance to acid soils as found in *B. decumbens* cv. Basilisk (Miles et al., 2004).

Obtaining spittlebug resistance was the major incentive for initiating Brachiaria breeding programs in America. Since no compatible sexual genotypes have been found within B. brizantha or B. decumbens, such combination of characters is possible only through interspecific hybridization. In the 1980s, the Embrapa Beef Cattle Center initiated a program of hybridization to better exploit the genetic variability found in some Brachiaria accessions. Interspecific hybridization is difficult to accomplish due to differences in ploidy levels and genetic affinity (Valle and Savidan, 1996; Valle and Pagliarini, 2009). Compatibility has been found among some species of Group 5 (Renvoize et al., 1996): B. brizantha, B. decumbens and B. ruziziensis, which form an agamic complex and produce fertile hybrids (Valle and Savidan, 1996; Valle and Milles, 2001).

Brachiaria breeding is a recent endeavor. Polyploid accessions with desirable agronomic characteristics are apomictic (Valle and Savidan, 1996), and sexuality is found only in diploid accessions. Ferguson and Crowder (1974) attempted interploidy crosses to no avail and thus

ABSTRACT: Hybrids combining desirable traits from divergent parents are the main objective of some *Brachiaria* (Syn. *Urochloa* P. Beauv.) breeding programs. There is great interest in the development of apomictic hybrid cultivars that combine desirable genes such as resistance to spittlebugs, high nutritive value, and tolerance to acid soils. Microsporogenesis of six apomictic progenies resulting from a tetraploid (2n = 4x = 36) cross between *B. ruziziensis* × *B. brizantha* was evaluated under light microscopy. Genetic recombination, ensured by multivalent chromosome association and crossing-over at prophase occurred in low frequency among progenies, and in one, recombination was almost nonexistent. The percentage of meiocytes with meiotic abnormalities among progenies ranged from 16.6 % to 85.6 %. Besides an observed irregular chromosome segregation typical of polyploid hybrids in these five progenies, putative meiotic mutations characterized as desynapsis and divergent spindle organization occurred in three progenies. These anomalies caused frequent fractionation of the genome into several microspores of different sizes. In *Brachiaria*, new cultivars must be apomictic to fix the genotype. However, *Brachiaria* is a pseudogamous apomict, and viable gametes are necessary to produce viable seeds. Considering meiotic behavior, only two progenies are promising for advancement in the breeding program.

Keywords: Brachiaria, microsporogenesis, breeding, pastures, apomixis

suggested doubling the chromosome number of sexual diploids to create a cross-compatible sexual tetraploid plant. This was accomplished a decade latter, in Belgium, when diploid accessions of B. ruziziensis were tetraploidized with colchicine (Gobbe et al., 1981; Swenne et al., 1981). These accessions continue to be the basis of the interspecific hybridization program both in CIAT (Miles et al., 2004) and in Brazil (Valle et al., 2009). Interspecific hybridization in Brachiaria was successfully accomplished between tetraploid accessions of B. ruziziensis \times B. brizantha or B. decumbens. Attempts to cross sexual B. decumbens \times B. brizantha were recently performed (Souza-Kaneshima et al., 2010). The single diploid accession of B. brizantha (Pinheiro et al., 2000) and some diploid accessions of B. decumbens (Simioni and Valle, 2009) were artificially tetraploidized by colchicine, creating new opportunities to recombine the genetic variability existing in each species.

The Brazilian program of interspecific hybridization in Brachiaria was initiated in 1988, when over 200 hybrids resulted from crosses between *B. ruziziensis* \times *B.* brizantha or B. decumbens. Several sexual and apomictic hybrids, mainly those resulting from *B. ruziziensis* \times *B.* brizantha, presented desirable traits such as resistance to spittlebugs, high nutritive value, and tolerance to acid soils. The sexual hybrids are maintained in an open polycross block, whereas the apomictic are under agronomic evaluation. In the genus Brachiaria, apomixis is pseudogamic, which means that viable male gametes are necessary to fertilize the secondary nucleus of the embryo sac to guarantee endosperm development to produce viable seeds (Valle and Savidan, 1996). However, polyploidy in Brachiaria causes abnormal meioses (Mendes-Bonato et al., 2002ab, 2006; Utsunomiya et al., 2005, Risso-Pascotto et al., 2006). Previous cytogenetic analyses of F, hybrids of *B. ruziziensis* \times *B. brizantha* hybrids (Risso-Pascotto et al., 2005a; Fuzinatto et al., 2007a, 2008, Adamowski et al., 2008) have revealed different types and frequencies of meiotic abnormalities, indicating that selection for meiotic stability is possible among them. We report herein an analysis of microsporogenesis in six apomictic progenies of a cross between a sexual tetraploidized accession of *B. ruziziensis* and a natural tetraploid of *B. brizantha*. Our goal was to select meiotically stable progenies for inclusion in our *Brachiaria* breeding program.

Materials and Methods

Hybrids (F, generation) were obtained in Campo Grande, state of Mato Grosso do Sul, Brazil (20°25' S, 54°46' W, 526 m a.s.l.), by crossing two accessions (R038 and B140) that exhibited desirable characters. R038 is an artificial tetraploid (2n = 4x = 36) that maintained sexuality after chromosome doubling in Belgium, and B140, the pollen donor, is a natural apomictic tetraploid (2n = 4x = 36 with high productivity and nutritive value. The latter is undergoing animal performance trials to be released as a new cultivar. Both accessions, the original diploid of B. ruziziensis (Zimbabwe, Africa, 20°28' S, 55°40' E, 520 m a.s.l) and B140 of B. brizantha (Rwanda, Africa, 20°26' S, 54°90' E, 1,030 m a.s.l.) were collected in Africa in 1984-85. Hybrids were planted in the field, and mode of reproduction was previously determined by examination of embryo sacs using interference contrast microscopy of methyl salicylate-cleared ovaries (Young et al., 1979). Among the obtained sexual and apomictic progenies, the six apomictic ones analyzed in the present paper (P05, P06, P07, P013, P014, and P020) were selected for use in our breeding program based on vigor and leafiness.

Inflorescences for meiotic studies were collected in each field-grown plant and fixed in a mixture of 95 % ethanol, chloroform and propionic acid (6:3:2) for 24 h, transferred to 70 % alcohol and stored under refrigeration until use. Microsporocytes were prepared by squashing and staining with 0.5 % propionic carmine. Meiotic abnormalities were observed in each phase of microsporogenesis. The number of cells analyzed per plant ranged from 958 to 1,717, according to the availability of inflorescences collected in each progeny. Photomicrographs were made with a Wild Leitz microscope using Kodak Imagelink – HQ, ISO 25 black and white film.

Results and Discussion

In the progenies here analyzed, bivalents predominated (Figure 1A), especially in P020, in which multivalents were rare. A maximum of three quadrivalents were observed in the other progenies. In plant breeding, the success of gene introgression via sexual hybridization depends on the phylogenetic relationships between species, opportunities for genetic recombination, and stability of the introgressed genes (Marfil et al., 2006). Among domesticated plants, cytological analyses are usually performed to evaluate the meiotic process in experimental hybrids. In Brachiaria, some studies were conducted in sexual and apomictic hybrids resulting from crosses between B. ruziziensis \times B. brizantha or B. decumbens (Risso-Pascotto et al., 2005a; Fuzinatto et al, 2007a; Adamowski et al., 2008; Felismino et al., 2010). Although these species belong to the same taxomonic group (Renvoize et al., 1996), the frequency of genetic recombination varies among them. A high frequency of multivalent chromosome association was found in three-way hybrids (Adamowski et al., 2008). One to three quadrivalents were found in the F, generations (Risso-Pascotto et al., 2005a; Fuzinatto et al., 2007a). The degree of genetic divergence in the hybrids is provided by chromosome pairing between the two genomes (Sundberg and Glimelius, 1991). A high pairing affinity of chromosomes indicates that interchange between gene pools of the genitors is possible (Zwierzykowski et al., 1999). Thus, a gene introgression can be expected among these progenies. Although the hybrids analyzed in the genus Brachiaria resulted from B. ruziziensis \times B. brizantha, genetic affinity in the genus Brachiaria has shown to be genome-specific (Risso-Pascotto et al., 2005a; Fuzinatto et al., 2007a; Adamowski et al., 2008; Felismino et al., 2010).

The degree of genetic divergence between genitors in a hybrid can be estimated not only by chromosome pairing, but also by the frequency of meiotic abnormalities (Rieseberg et al., 2000). In the six progenies, the mean percentage of abnormalities ranged from 16.6 % to 85.6 % (Table 1). Except for P020, all progenies presented irregular chromosome segregation, a typical abnormality of polyploids and widely reported in tetraploid Brachiaria hybrids (Risso-Pascotto et al., 2005a; Fuzinatto et al., 2007a; Adamowski et al., 2008; Felismino et al., 2010). Irregular chromosome segregation was characterized by precocious chromosome migration to the poles (Figure 1B), laggards (Figure 1C), and micronuclei (Figure 1D) in both meiotic divisions. Micronuclei remained in the microspores (Figure 1E, F) or were isolated in microcytes after abnormal cytokinesis (Figure 1E). In such cases, unbalanced microspores would be formed. The absence of irregular chromosome segregation in P020, coupled with the low frequency of the observed quadrivalent formation, suggest that the segregant parental genomes had low affinity, and the chromosomes associated preferentially as bivalents.

Two other meiotic abnormalities were recorded in some progenies: divergent spindles (P05, P07, and P020) and desynapsis (P05). Several putative meiotic mutations have been reported among *Brachiaria* accessions (Mendes-Bonato et al., 2002b, 2003, 2007; Risso-Pascotto et al., 2003, 2005b; Mendes-Vieira et al., 2005; Boldrini et al. 2006; Gallo et al., 2007; Adamowski et al., 2007; Calisto et al., 2008) and in hybrids (Mendes-Bonato et al., 2004, 2006b; Risso-Pascotto et al., 2005a; Fuzinatto et al., 2007ab; Adamowski et al., 2008; Felismino et al., 2008, 2010). These putative mutations are frequently be-

| | | P05 | | P06 | | P07 | | P013 | | P014 | | P020 | |
|-------|--|--------------|----------------------|-----------------|--------------------|-----------------|---------------------|--------------|----------------------|-----------------|--------------------|-----------------|---------------------|
| Phase | Abnormalities | No. of cells | Abn** cells | No. of cells | Abn. cells | No. of cells | Abn. cells | No. of cells | Abn. cells | No. of cells | Abn. cells | No. of cells | Abn. cells |
| | | | % | | % | | % | | % | | % | | % |
| MI* | Precocious migration Desynapsis | | 13.5 10.9 28.5 | | 18.5 | | 14.3 - 22.6 | | 24.2 | | 26.2 | | - 13.6 |
| | Divergent spindle Multiple spindle | 193 | 28.5 17.6 | 168 | - | 168 | - | 182 | - | 237 | - | 206 | - |
| AI | Laggards Divergent spindle Multiple spindle | 84 | 23.8 32.1 26.2 | 70 | 34.3 - - | 117 | 29.9 17.1 | 173 | 54.3 - - | 289 | 27.7 - - | 68 | - 11.8 - |
| TI | Micronuclei | 186 | 84.4 | 215 | 13.0 | 198 | 39.9 | 176 | 39.8 | 198 | 26.8 | 211 | 17.6 |
| PII | Micronuclei Abnormal cytokinesis | 76 | 72.4 | 162 | 13.6 | 184 | 4.9 58.1 | 123 | 29.3 | 190 | 14.2 | 152 | - 67.1 |
| MII | Precocious migration Multiple spindle Abnormal cytokinesis | 245 | 2.9 87.8 - | 81 | - - - | 131 | - - 87.8 | 40 | 42.5 - - | 259 | 14.3 - - | 120 | - 64.2 |
| All | Laggards Multiple spindle Abnormal cytokinesis | 68 | 5.9 77.9 - | 45 | 20.0 | 13 | - - 7.7 | 18 | 44.4 - - | 94 | 22.3 - - | 19 | - 21.1 |
| TII | Laggards Multiple spindle Abnormal cytokinesis | 157 | 3.18 91.7 | 81 | 9.9 - - | 92 | - | 42 | 45.2 - - | 183 | 26.2 | 121 | - 58.7 |
| Tetr | Micronuclei Microcytes Polyads | 454 | 5.9 3.3 80.0 | 185 | 6.5 4.3 13.5 | 352 | 9.9 21.6 36.1 | 70 | 55.7 37.1 45.7 | 267 | 28.5 9.7 9.0 | 137 | 10.4 6.7 10.4 |
| Total | - | 1463 | 85.6 | 1007 | 16.6 | 1255 | 53.1 | 824 | 46.7 | 1717 | 26.4 | 1034 | 33.2 |

Table 1 – Meiotic abnormalities and percentage of abnormal cells in apomictic progeny of *Brachiaria* hybrids.

*MI: metphase I; AI: anaphase I; TI: telophase I; PII: Prophase II; MII: metaphase II; AII: anaphase II; TI: telophase II; Tet: tetrad. **Abn: Abnormal.

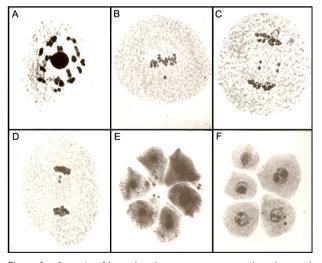


Figure 1 – Aspects of irregular chromosome segregation observed in the *Brachiaria* hybrid progenies. A) Diakinesis with 18 bivalents. B) Metaphase I with precocious chromosome migration to the pole. C) Anaphase I with laggards. D) Early telophase I with micronuclei. E) Polyad with microcytes. F) Pentad with micronucleus in one microspore.

ing expressed in different genotypes of *Brachiaria* suggesting that these genes are present in the gene pool of this genus. One of these genes is the 'divergent spindle'

(dv), reported in another *Brachiaria* hybrid resulting from a different cross between *B. ruziziensis* × *B. brizantha* (Mendes-Bonato et al, 2006b; Felismino et al., 2008). This gene affects spindle organization such that chromosomes do not converge to focused poles. In the mutants, bivalents are distantly spread over a large metaphase plate, and they fail to converge.

Depending on the distance between chromosomes at the poles, telophase I nuclei are elongated or the chromosomes are grouped into several micronuclei of different sizes in each pole (Figure 2A, B). In the presence of micronuclei, an abnormal cytokinesis occurs dividing the cytoplasm into three, four or more cells, according to the micronuclei number (Figure 2B, C, D). In each cell, the second division progresses normally (Figure 2C, D, E), generating polyads (Figure 2F). The percentage of cells expressing the divergent spindle phenotype was variable among progenies. Variations in penetrance and expressivity of dv were reported in maize and Brachiaria (Golubovskaya and Mashnenkov, 1981; Staiger and Cande, 1990; Shamina et al., 2000; Mendes-Bonato et al., 2006b; Felismino et al., 2008). This gene leads to genome fractionation, causing pollen sterility.

The other meiotic abnormality (desynapsis) was recorded only in P05. This abnormality was reported in one accession of *B. humidicola* (Calisto et al., 2008) and was correlated with abnormal cytokinesis. In P05, the expres-

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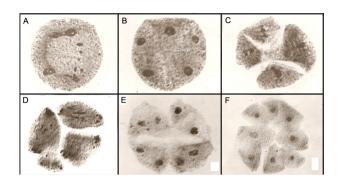


Figure 2 – Aspects of divergent spindle expression in the *Brachiaria* hybrid progenies. A) Telophase I with several micronuclei. B) Telophase I with five nuclei. C) Metaphase II after abnormal cytokinesis. D) Anaphase II. E) Telophase II. F) Octad of microspores.

sivity was different from the former. Chromosomes were associated in bi- or quadrivalents in diakinesis, but did not congregate at the metaphase plate. Chromosomes and chromatids separated precociously, so that 72 units could be counted (Figure 3A, B). Anaphase I did not occur, and the chromatids were grouped (Figure 3C) into several micronuclei of different sizes (Figure 3D). After this phase, abnormal cytokinesis occurred, dividing the cytoplasm into several cells with micronuclei (Figure 3E, F). After a new abnormal cytokinesis, polyads with unequal microspores (Figure 3G) generated meiotic products with pollen grains of different sizes (Figure 3H, I). Desynapsis is frequently reported in higher plants (Koduru and Rao, 1981). A similar phenotype was reported in Vaccinium darrowi (Qu and Vorsa, 1999). The direct consequences of mutations affecting synapsis are the production of abnormal gametes which are generally unable to function because of the unbalanced number of chromosomes.

Irregular chromosome segregation, desynapsis, and divergent spindles (leading to multiple spindle formation and abnormal cytokinesis) caused, to varying degrees among progenies, genome fractionation and pollen inviability. In the Brachiaria hybridization program, desirable combinations of agronomic characters and high seed set are required to meet the Brazilian demand for extensive pasture establishment, pasture renovation and export. Apomictic hybrids with putative mutations, such as divergent spindle and desynapsis, must be discarded from the breeding program to avoid compromising future generations. From our cytological analyses, P06 and P014, with a mean of 16.6 % and 26.4 % of cells with meiotic abnormalities, respectively, should remain in the breeding program. In these lines, abnormalities are generally limited to a low frequency of irregular chromosome segregation. Hence, genetic recombination of desirable genes is expected. Our cytogenetic studies of interspecific hybrids attest to their value in breeding programs. Time and effort can be saved when meiotic abnormalities are discovered early, and genotypes with potential cytogenetic problems are discarded.

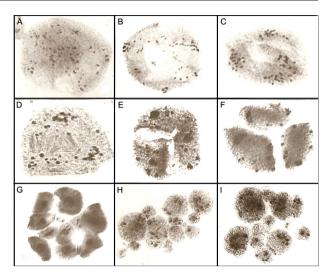


Figure 3 – Aspects of desynapsis in the progeny P05. A, B) Microsporocyte with 72 chromatids. C) Early telophase I with some groups of chromatids. D) Telophase I with several micronuclei of different sizes. E, F) Meiocytes in the second division with several micronuclei. G) Polyad of microspores. H, I) Pollen grains of different sizes resulted from the abnormal division.

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References

- Adamowski, E.V.; Boldrini, K.R.; Pagliarini, M.S.; Valle, C.B. 2007. Abnormal cytokinesis in microsporogenesis of *Brachiaria humidicola* (Poaceae: Paniceae). Genetics and Molecular Research 6: 616–621.
- Adamowski, E.V.; Pagliarini, M.S.; Valle, C.B. 2008. Meiotic behavior in three interspecific three-way hybrids between *Brachiaria ruziziensis* and *B. brizantha* (Poaceae: Paniceae). Journal of Genetics 87: 33-38.
- Boldrini, K.R.; Pagliarini, M.S.; Valle, C.B. 2006. Abnormal timing of cytokinesis in microsporogenesis of *Brachiaria humidicola* (Poaceae: Paniceae). Journal of Genetics 85: 225-228.
- Calisto, V.; Fuzinatto, V.A.; Message, H.J.; Mendes-Bonato, A.B.; Boldrini, K.R.; Pagliarini, M.S.; Valle, C.B. 2008. Desynapsis and precocious cytokinesis in *Brachiaria humidicola* (Poaceae) compromise meiotic division. Journal of Genetics 87: 1-5.
- Felismino, M.F.; Pagliarini, M.S.; Valle, C.B. 2008. A differential phenotype expression of a divergent spindle mutation in interspecific *Brachiaria* hybrids. Cell Biology International 32: 1459–1463.
- Felismino, M.F.; Pagliarini, M.S.; Valle, C.B. 2010. Meiotic behavior of interspecific hybrids between artificially sexual *Brachiaria ruziziensis* and tetraploid apomictic *B. brizantha* (Poaceae). Scientia Agricola 67: 191–197.
- Ferguson, J.E.; Crowder, L.V. 1974. Cytology and breeding behaviour of *Brachiaria ruziziensis*. Crop Science 14: 893-895.

- Fuzinatto, V.A.; Pagliarini, M.S.; Valle, C.B. 2007a. Microsporogenesis in sexual *Brachiaria* hybrids (Poaceae). Genetics and Molecular Research 6: 1107–1117.
- Fuzinatto, V.A.; Pagliarini, M.S.; Valle, C.B. 2007b. Evidence of programmed cell death during microsporogenesis in an interspecific *Brachiaria* (Poaceae: Panicoideae: Paniceae) hybrid. Genetics and Molecular Research 6: 208–215.
- Fuzinatto, V.A.; Pagliarini, M.S.; Valle, C.B. 2008. Evaluation of microsporogenesis in an interspecific *Brachiaria* hybrid (Poaceae) collected in distinct years. Genetics and Molecular Research 7: 424–432.
- Gallo, P.H.; Micheletti, P.L.; Boldrini, K.R.; Risso-Pascotto, C.; Pagliarini, M.S.; Valle, C.B. 2007. 2n gamete formation in the genus *Brachiaria* (Poaceae: Paniceae). Euphytica 154: 255–260.
- Gobbe, J.; Swenne, A.; Louant, B.P. 1981. Natural diploids and induced autotetraploids in *Brachiaria ruziziensis* Germain et Evrard: criteria of identification. Agronomia Tropicale 36: 339– 346 (in French).
- Golubovskaya, I.N.; Mashnenkov, A.S. 1981. Genetic control of chromosome segregation during the first meiotic division. Maize Genetics Cooperation News Letter 55: 78–81.
- Koduru, P.R.K.; Rao, M.K. 1981 Cytogenetics of synaptic mutants in higher plants. Theoretical and Applied Genetics 59: 197–214.
- Marfil, C.; Masuelli, R.W.; Davison, J.; Comai, L. 2006. Genomic instability in Solanum tuberosum × Solanum kurtzianum interspecific hybrids. Genome 49: 104–113.
- Mendes-Vieira, D.; Boldrini, K.R.; Mendes-Bonato, A.B.; Pagliarini, M.S.; Valle, C.B. 2005. Abnormal meiotic in *Brachiaria brizantha* (Poaceae) leading to microspore degeneration. Caryologia 58: 396–402.
- Mendes-Bonato, A.B.; Pagliarini, M.S.; Forli, F.; Valle, C.B.; Penteado, M.I.O. 2002a. Chromosome number and microsporogenesis in *Brachiaria brizantha* (Gramineae). Euphytica 125: 419–425.
- Mendes-Bonato, A.B.; Junqueira Filho, R.G.; Pagliarini, M.S.; Valle, C.B. 2002b. Unusual cytological patterns of microsporogenesis in *Brachiaria decumbens*: abnormalities in spindle and defective cytokinesis causing precocious cellularization. Cell Biology International 26: 641–646.
- Mendes-Bonato, A.B.; Risso-Pascotto, C.; Pagliarini, M.S.; Valle, C.B. 2003. Normal microspore production after cell fusion in *Brachiaria jubata* (Gramineae). Genetics and Molecular Biology 26: 517–520.
- Mendes-Bonato, A.B.; Pagliarini, M.S.; Valle, C.B. 2004. Abnormal pollen mitoses (PMI and PMII) in an interspecific hybrid of *Brachiaria ruziziensis* and *Brachiaria decumbens* (Gramineae). Journal of Genetics 83: 279–283.
- Mendes-Bonato, A.B.; Risso-Pascotto, C.; Pagliarini, M.S.; Valle, C.B. 2006a. Chromosome number and meiotic behavior in *Brachiaria jubata* (Gramineae). Journal of Genetics 85: 83–88.
- Mendes-Bonato, A.B.; Pagliarini, M.S.; Valle, C.B. 2006b. Abnormal spindle orientation during microsporogenesis in an interspecific *Brachiaria* (Gramineae) hybrid. Genetics and Molecular Biology 29: 122–125.
- Mendes-Bonato, A.B.; Pagliarini, M.S.; Valle, C.B. 2007. Meiotic arrest compromises pollen fertility in an interspecific hybrid between *Brachiaria ruziziensis* × *Brachiaria decumbens* (Poaceae: Paniceae). Brazilian Archives of Biology and Technology 50: 831–837.

- Miles, J.W.; Valle, C.B.; Rao, I.M.; Euclides, V.P.B. 2004. Brachiaria grasses. p. 745–760. In: Sollenberger, L., ed. Warm-season (C4) grasses. ASA/CSSA/SSSA, Madison, Wisconsin, USA.
- Pinheiro, A.A.; Pozzobon, M.T.; Valle, C.B.; Penteado, M.I.O.; Carneiro, V.T.C. 2000. Duplication of the chromosomes number of diploid *Brachiaria brizantha* plants using colchicine. Plant Cell Reports 9: 274–278.
- Qu, L.; Vorsa, N. 1999. Desynapsis and spindle abnormalities leading to 2n pollen formation in *Vaccinium darrowi*. Genome 42: 35-40.
- Renvoize, S.A.; Clayton, W.D.; Kabuye, C.H.S. 1996. Morphology, taxonomy, and natural distribution of *Brachiaria* (Trin.) Griseb. p. 1–15. In: Miles, J.W.; Maass, B.L.; Valle, C.B., eds. *Brachiaria: biology*, agronomy, and improvement. CIAT/EMBRAPA, Cali, Colombia.
- Rieseberg, L.H.; Baird, S.T.J.; Gardner, K.A. 2000. Hybridization, introgression, and linkage evolution. Plant and Molecular Biology 42: 205-224.
- Risso-Pascotto, C.; Pagliarini, M.S.; Valle, C.B. 2003. A mutation in the spindle checkpoint arresting meiosis II in *Brachiaria ruziziensis*. Genome 46: 724–728.
- Risso-Pascotto, C.; Pagliarini, M.S.; Valle, C.B. 2005a. Meiotic behavior in interspecific hybrids between *Brachiaria ruziziensis* and *Brachiaria brizantha* (Poaceae). Euphytica 145: 155–159.
- Risso-Pascotto, C.; Pagliarini, M.S.; Valle, C.B.; Jank, L. 2005b. Symmetric pollen mitosis I and suppression of pollen mitosis. II. Prevent pollen development in *Brachiaria jubata* (Gramineae). Brazilian Journal of Medical and Biological Research 38: 1603– 1608.
- Risso-Pascotto, C.; Pagliarini, M.S.; Valle, C.B. 2006. Microsporogenesis in *Brachiaria dictyoneura* (Fig. & De Not.) Stapf (Poaceae: Paniceae). Genetics and Molecular Research 5: 837–845.
- Shamina, N.; Dorogova, N.; Trunova, S. 2000. Radial spindle and the phenotype of the maize meiotic mutant, dν. Cell Biology International 24: 729–736.
- Simioni, C.; Valle, C.B. 2009. Chromosome duplicaction in Brachiaria (A. Rich.) Stapf allows intraspecific crosses. Crop Breeding and Applied Biotechnology 9: 328-334.
- Souza-Kaneshima, A.M.; Simioni, C.; Felismino, M.F.; Mendes-Bonato, A.B.; Risso-Pascotto, C.; Pessim, C.; Pagliarini, M.S.; Valle, C.B. 2010. Meiotic behavior in the first interspecific hybrid between *B. brizantha* and *B. decumbens*. Plant Breeding 129: 186–191.
- Staiger, C.J.; Cande, W.Z. 1990. Microtubule distribution in dv, a maize meiotic mutant defective in the prophase to metaphase transition. Developmental Biology 138: 231–242.
- Sundberg, E.; Glimelius, K. 1991. Effects of parental ploidy and genetic divergence on chromosome elimination and chloroplast segregation in somatic hybrids within Brassicaceae. Theoretical and Applied Genetics 83: 81–88.
- Swenne, A.; Louant, B.P.; Dujardin, M. 1981. Induction by colchicine of tetraploid accession of *Brachiaria ruziziensis* Germain et Evrard (Gramineae). Agronomia Tropicale 36: 134–141 (in French).
- Utsunomiya, K.S.; Pagliarini, M.S.; Valle, C.B. 2005. Microsporogenesis in tetraploid accessions of *Brachiaria nigropedata* (Ficalho & Hiern) Stapf (Gramineae). Biocell 29: 295–301.

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- Valle, C.B.; Savidan, Y. 1996. Genetics, cytogenetics, and reproductive biology of *Brachiaria*. p. 147–163. In: Miles, J.W.; Maass B.L.; Valle, C.B., eds. *Brachiaria*: biology, agronomy, and improvement. CIAT/EMBRAPA, Cali, Colombia.
- Valle, C.B.; Miles, J.W. 2001. Breeding of apomictic species. p. 137– 152. In: CYMMYT. The flowering of apomixis: from mechanisms to genetic engineering. CYMMYT/IRD, Mexico, DF, Mexico.
- Valle, C.B.; Pagliarini, M.S. 2009. Biology, cytogenetics, and breeeding of *Brachiaria*. p. 103–151. In: Singh, R.J., ed. Genetic resources, chromosome engineering, and crop improvement. CRC Press, Boca Raton, USA.
- Valle, C.B.; Jank, L.; Resende, R.M.S. 2009. Forage breeding in Brazil. Revista Ceres 56: 460-472 (in Portuguese).
- Young, B.A.; Sherwood, R.T.; Bashaw, E.C. 1979. Cleared-pistyl and thick-sectioning techniques for detecting aposporous apomixis in grasses. Canadian Journal of Botany 57: 1668–1672.
- Zwierzykowski, Z.; Lukaszewski, A.J.; Naganowska, B.; Lesniewska, A. 1999. The pattern of homeologous recombination in triploid hybrids of *Lolium multiflorum* with *Festuca pratensis*. Genome 42: 720–726.