The Echocardiography in the Cardiovascular Laboratory: A Guide to Research with Animals

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Abstract
The feasibility and potential for the morphological and hemodynamic investigation of the heart has been increasing the use of the echocardiography in the research setting. Additionally, the development of new technologies, like the real time 3D echocardiography and speckle tracking, demands validation throughout experimental studies before being instituted in the clinical setting.

This paper aims to provide information concerning the particularities of the echocardiographic examination in quadruped mammals, targeting the experimental research.

Introduction
Considering the low cost and noninvasive nature, the portability and potential of morphological and hemodynamic investigation of the heart, the echocardiography plays an important role in the clinical and experimental research setting.

The Swedish physician Inge Edler, considered as “The Father of the Echocardiography”, was the first to use this method in the experimental research. In the early 50’s, Edler demonstrated in calf hearts that the echo signals, firstly thought to be generated by left atrial anterior wall, in fact were from the anterior mitral valve leaflet. Since then, an increasing number of experimental studies using echocardiography has been published.

Recently, Real Time 3D Echocardiography (RT3DE), Tissue Doppler Imaging (TDI) and Speckle Tracking (STE) represent novel technology tools in the early diagnosis of dys synchrony, diastolic and systolic heart dysfunction, shedding light on scientific knowledge for clinical applications. Although cardiac magnetic resonance imaging is currently considered the reference technique for ventricle volumetry and calculation of the ejection fraction, several echocardiographic parameters can provide reliable information on ventricular dimensions and volumes, besides myocardial function in daily clinical practice.

This paper aims to review the technical aspects of echocardiography in the experimental laboratory, considering its advantages and limitations according to the animal species.

The Transthoracic Echocardiography (TTE) Planes and Animal Model
Nowadays, the transthoracic echocardiographic investigation in animals employed in experimental research is favored by the technological development of the equipment; however, it is crucial to select the right frequency transducer and have the knowledge on quadruped mammals peculiarities.

According to physical principles, low-frequency transducers (2.0 – 3.5 MHz) offer smaller image resolution; nevertheless, the ultrasound beam penetration is enhanced, which allows their use in animals with a bigger body surface area. Inversely, the high-frequency probes (4.0 – 8.0 MHz) do not enable larger penetrations, however, they are feasible for a better image resolution, being more adequate for smaller and/or younger animals. Regarding rabbits and small rodents (rats, mice, and guinea pigs), in order to obtain a satisfactory image resolution, the use of higher frequency transducer (10.0 – 15.0 MHz) is needed, because of their very small size. Therefore, once the frequency of the transducer is inversely proportional to the ultrasound beam penetration, the smaller the mammal, the greater the frequency of the probe.

The heart of quadruped mammals present the heart located in the middle part of chest, with the apex pointed to the left; also, the chest is generally keel-shaped, allowing to obtain images from both left and right sides of the thorax, as shown below.

In order to acquire images of optimal quality, the animal should have its fur chest removed, avoiding the presence of air between the transducer and the body surface, once it represents a barrier to the echo beam penetration. Besides that, sedation or anesthesia is crucial to the accuracy of the test; choosing the proper drug and dosage, according to the species, and the appraisal of possible alterations in the cardiovascular system should be scrutinized. The positioning varies according to the animal model: for dogs and pigs, left lateral decubitus is indicated; rabbits and little rodents should be kept in dorsal decubitus while ruminants need a customized stretcher due to their extremely kneel-shaped chest. (Figure 1)
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The use of echocardiography in cardiovascular research

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Figure 1 - Customized stretcher for animals with kneel-shaped chest, in order to obtain the parasternal echocardiographic views.

Table 1 - Rapid guide to the echocardiographic examination in animal cardiovascular research

<table>
<thead>
<tr>
<th>Animal</th>
<th>Transducer Frequency (MHz)</th>
<th>Animal Position</th>
<th>Main Echocardiographic Planes</th>
<th>Main Limitations</th>
</tr>
</thead>
<tbody>
<tr>
<td>Small rodents</td>
<td></td>
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<tr>
<td>Rabbits</td>
<td>10.0 – 15.0</td>
<td>Left lateral decubitus</td>
<td>PLAX, short-axis, apical</td>
<td>Elevated heart rate</td>
</tr>
<tr>
<td>Goats/Sheep</td>
<td>2.5 – 3.5 (adults)</td>
<td>Left lateral decubitus</td>
<td>PLAX, short axis</td>
<td>Chest shape</td>
</tr>
<tr>
<td></td>
<td>4.0 – 8.0 (young)</td>
<td>Customized stretcher</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dogs</td>
<td>2.5 – 3.5 (adults)</td>
<td>Left lateral decubitus</td>
<td>PLAX, short-axis, apical</td>
<td>---</td>
</tr>
<tr>
<td></td>
<td>4.0 – 8.0 (young)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pigs</td>
<td>2.5 – 3.5 (adults)</td>
<td>Left lateral decubitus</td>
<td>PLAX, short-axis, apical</td>
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</tr>
<tr>
<td></td>
<td>4.0 – 8.0 (young)</td>
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PLAX: paraesternal long-axis view.

Usually, the echocardiographic image acquisitions are the same as in humans, except for the ruminants, which do not have the apical approach. The heart of quadruped mammals, which occupies a medial position in the chest, and the apex pointed to the left and the diaphragm, confers two echocardiographic peculiarities. One of them is indeed a disadvantage: the impossibility of acquiring images of good quality through the subcostal approach, once the angle of insonation is not perpendicular to the heart (among the ruminants this is exacerbated by the presence of the pre-stomachs). The other is the feasibility of obtaining images both by left and right hemithorax.

Subsequently, a description of the approaches particularities according to the animal model; Table 1 provides a summarized guide related to each one of them and Figure 2 depicts the transducer position in relation to the chest of the animal. Of note, regarding the normal values of heart structures (diameters and volumes), they vary according to the species and size; therefore, the baseline echocardiography is focused on the myocardial function and valvular performance analysis before the experimental procedure, in order to confirm that the heart of the animals can be considered healthy.

Left parasternal approach

Feasible to obtain in dogs, pigs, rabbits and little rodents, at the third intercostal space.

Parasternal longitudinal plane (Figure 3A): allows visualization of the right ventricle (RV), left ventricle (LV), left ventricular outflow tract (LVOT), aorta (Ao) and aortic valve (AV), mitral valve (MV) and left atrium (LA).

Parasternal short-axis view: at the basal level it is observed the AV in the middle, the RV, right atrium (RA), tricuspid valve (TV) and the LA (Figure 3B). Tilting the probe towards the animal’s head, it is possible to obtain images of the right ventricular outflow tract (RVOT), pulmonary valve (PV) and pulmonary trunk (PT), the latter winging in the right and left pulmonary branches (Figure 3C). The short axes at the MV and papillary muscles level generally are not well visualized in this approach, because of the obliquely positioned heart.

Long-axis view of the right ventricular inflow tract (Figure 3D).

Right parasternal approach

Achievable in all animal models, at the third intercostal level. Of note, regarding the ruminants, this is the only reliable transthoracic echocardiography approach, allowing measurements that would not be feasible by the conventional views, as in the case of right ventricular mass calculation, where the long-axis view of the right ventricular inflow tract act as a surrogate for the apical 4-chamber view.

Parasternal longitudinal view (Figure 4A). In this view it is possible to visualize the same structures assessed through the left parasternal longitudinal approach (Ao, LA, LV and RV), besides the RA.

Left and right ventricular inflow tract long-axis view (Figure 4B) As mentioned above, this view might be used as a substitute for the apical view; however, care must be taken when considering the true apical region, which cannot be evaluated by this approach.

Parasternal short-axis view: obtained at the basal, MV (Figure 4C), and papillary muscles level (Figure 4D), by tilting
Figure 2 - Position of the transducer in relation to the chest of the animal in left (A) and right (B) hemithorax approach.

Figure 3 - Echocardiographic views from the left parasternal approach in a dog. Parasternal long-axis view (A). Parasternal short-axis view at the basal level (B and C) Long-axis view of the right ventricular inflow tract (D). RV: right ventricle; LV: left ventricle; Ao: aorta; LA: left atrium; RA: right atrium; PT: pulmonary trunk; RPA: right pulmonary artery; LPA: left pulmonary artery.
Figure 4 - Echocardiographic views from the right parasternal approach in a dog (A and C) and a goat (C and D). Parasternal long-axis view (A). Left and right ventricular inflow tract long-axis view (B). Parasternal short-axis view at the baseline (C) and at the papillary muscles level (D). MV: mitral valve; RV: right ventricle; LV: left ventricle; Ao: aorta; LA: left atrium; RA: right atrium.

Apical approach

Possible to obtain in dogs, pigs, rabbits and little rodents, at the fifth intercostal space, in the left hemithorax, at the ictus cordis (Figure 5). This plane is essential to perform the TDI analysis, since the Doppler sample volume has to be parallel to the myocardial wall\textsuperscript{13}.

The Transesophageal Echocardiography (TEE)

The TEE in the experimental lab is proving to be a reliable tool when TTE does not provide adequate images or when it is necessary to assess the heart structures during surgical procedure\textsuperscript{14}. It is feasible in a variety of animal models and, again, the drug used to achieve anesthesia must be chosen according to the species\textsuperscript{14-19}.

The images are very similar to those obtained in humans, composed by a transgastric view and three additional views within the esophagus: cranial, middle and caudal.

Transgastric

This view is very limiting, only approaching the LV in the short axis view.

Caudal Esophagus

Short axis view: provides images of the right and left ventricles at the MV level; soft anti-flexion movements allow the approach of the heart base, at the AV level, similar to the parasternal short axis view in the TTE.

Long axis view: in this plane it is possible to obtain images of the LV, LVOT and MV; rotating the probe counterclockwise allows imaging the left ventricular inflow tract, RA, atrial septum, and caval veins.

Middle Esophagus

Four-chamber view (Figure 6).

Long axis view: provides imaging of the LA, LV and aorta, like in the TTE apical longitudinal view.

Cranial Esophagus

Short axis view: aortic arch, LVOT and pulmonary valve.

Long axis view: the centrally-oriented beam allows imaging the ascending aorta and the LVOT; the clockwise rotation of the transducer enables imaging the RVOT and the PT.

Novel Technology

Over the past few decades, there has been a rapid and increasing development of novel technology and, usually, those tools need to be tested experimentally before they are introduced in the clinical scenario.
Contrast echocardiography aims to improve the endocardial border delineation and, associated with the stress echocardiography, it can be employed in the research setting for myocardial perfusion assessment.

Real time 3D echocardiography, 2D and 3D STE are promising novel tools for the evaluation and early diagnosis of cardiac diseases. The experimental validation of these techniques was important to support their use in the clinical setting, mainly considering the STE, once there is no true gold standard that might be applied with this purpose to humans: considering the torsional mechanics, the elegant study of Helle Valle et al., as well as the assays of Langeland et al., Amundsen et al., and Seo et al. regarding to the 2D and
3D myocardial strain, the technique was validated against sonomicrometry, which demands open chest surgery and the insertion of crystals in the myocardium. Table 2 shows the correlation and agreement between the gold standard and STE in these trials. However, in order to obtain reliable data, it is very important to proceed with the inhalation anesthesia to induce respiratory apnea while acquiring the images, once both the RT3DE and STE have their accuracy affected by the respiratory movements of chest. Another relevant issue concerns the suitable preset, since low temporal resolution, represented by the frame rate, can also interfere with the results.22.

**Final Considerations**

According to the best of the authors’ knowledge, the first systematic studies employing the echocardiography in the experimental laboratory were performed in the 1960’s. Lategola, in 1966, through a miniaturized ultrasonic transducer placed inside the aorta of dogs, evaluated the stroke volume invasively, in a prototype that, few years later, would be clinically applied, by means of the transthoracic Continuous Wave Doppler29. Two years later, Christiensen and Bonte showed, in dogs, the accuracy of the echocardiography to detect pericardial effusion. Since then, its importance has been increasing in the experimental investigation scenario, seeking for clinical applications30.

Unlike using other technologies, like magnetic resonance imaging, relevant information can be easily obtained by the echocardiography approach. According to the authors’ experience, the method is useful to follow-up myocardial hypertrophy and failure31,32,33.

The development of novel technologies, such as the ultrasound biomicroscopy, which uses frequencies from 30 to 100 MHz, allows the assessment of tiny structures – like the mice fetal heart in the uterus33-35. Certainly, it will be of extremely importance to shed light on issues of the cardiovascular system and to the development of microsurgeries in small animals. It represents a very appealing option in experimental research, since those animals are easily manipulated, have a better cost-benefit ratio, and can be maintained in small areas, enabling study larger samples in a shorter period of time.

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**Author contributions**

Conception and design of the research: Abduch MCD, Aiello VD; Acquisition of data, Analysis and interpretation of the data and Critical revision of the manuscript for intellectual content: Abduch MCD, Assad RS, Aiello VD; Writing of the manuscript: Abduch MCD, Assad RS, Mathias Jr. W, Aiello VD.

**Potential Conflict of Interest**

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**Study Association**

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**Table 2 - Experimental validation of speckle tracking against sonomicrometry**

<table>
<thead>
<tr>
<th>Study type (Reference #)</th>
<th>Parameter</th>
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<tbody>
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<td>2D STE (22)</td>
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<td>Basal rotation</td>
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<td>2D STE (27)</td>
<td>Longitudinal Strain</td>
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<tr>
<td>3D STE (28)</td>
<td>Area Strain</td>
<td>0.87</td>
</tr>
</tbody>
</table>

2D STE: two-dimensional speckle tracking; 3D STE: three-dimensional speckle tracking.

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**References**


