Ocular manifestations of leishmaniasis in a cat: first case report from Brazil

[Manifestações oculares da leishmaniose em um gato: primeiro relato de caso do Brasil]


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ABSTRACT

An 8-year-old domestic short hair female cat initially presented with bilateral uveitis with pseudotumoral appearance. The patient tested negative for feline immunodeficiency virus (FIV), feline leukemia virus (FeLV) and Toxoplasma gondii. Histopathology of a granulomatous lesion on the upper left conjunctiva revealed amastigotes compatible with Leishmania spp. Aqueous humor was aspired and the diagnosis was confirmed after isolation of promastigotes cultivated in biphasic NNN medium and by positive polymerase chain reaction (PCR) for Leishmania infantum. Treatment with allopurinol (10mg/kg/BID/PO) was commenced and a natural insect repellent was prescribed. Six months of treatment with allopurinol associated with the initial topical medications helped to improve ocular signs. Leishmaniasis should be considered as a differential diagnosis in cats presenting uveitis with pseudotumoral appearance. To our knowledge, this is the first report of feline leishmaniasis with ocular manifestation in Brazil, in which diagnosis was confirmed by aqueous humor analysis.

Keywords: cat, uveitis, Leishmania infantum, paracentesis

INTRODUCTION

Leishmaniasis is a chronic zoonotic disease, which can potentially cause death (Pennisi et al., 2015). In the Americas, spreading occurs through sand flies of the genus Lutzomyia that transmit Leishmania infantum, a protozoan capable of causing the severe form of the disease (Almeida et al., 2012).

In Brazil, dogs were considered to be the only domestic reservoir of L. infantum until 2008...
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(Silva et al., 2008). However, natural infection of domestic cats with *Leishmania spp.* has been reported in 4/27 Brazilian states (Pennisi et al., 2015).

To date, *L. infantum* is the most frequently isolated species in Brazil identified in cats and in several countries throughout the world, such as Spain, Italy, Iran, Switzerland, Portugal and Greece (Pennisi et al., 2015).

Most cats affected by visceral leishmaniasis exhibit mucocutaneous lesions and lymphadenomegaly (Pennisi et al., 2015). Ophthalmic abnormalities secondary to leishmaniasis infection have been described in approximately one third of affected cats (Pennisi et al., 2015). Unilateral or bilateral uveitis is the most common ophthalmic finding in these cases, and generally exhibits a granulomatous pseudotumoral pattern that may progress to panophthalmitis (Leiva et al., 2005, Ritcher et al., 2014).

This report aimed to describe general and ocular clinical signs of a cat with visceral leishmaniasis, in which the ophthalmic diagnosis was confirmed by aqueous humor culture and PCR. In addition, to describe the evolution of the clinical signs during a 6-months period. To our knowledge, this is the first report of feline leishmaniasis with ocular manifestation in Brazil.

**CASE REPORT**

An 8-year-old female domestic shorthair cat was presented at the Ophthalmology Service of the Veterinary Teaching Hospital of the Federal University of Mato Grosso, Cuiabá-MT, with a history of bilateral photophobia, blepharospasm, and conjunctival hyperemia for approximately 3 months. The owner denied previous treatment. Deworming and vaccinations were updated. On physical examination, the patient exhibited good general condition, with no signs of malnutrition, skin disease, lymphadenomegaly, or abnormalities on abdominal palpation. All physiological parameters were within the normal ranges for the species.

On ophthalmic examination, the Schirmer tear test readings (Ophthalmos Ind. Farmacêutica, São Paulo, Brazil) were 8 and 10 mm/min in the *oculus dexter* (OD) and *oculus sinister* (OS), respectively. Menace response, pupillary and dazzle reflexes were normal. Slit-lamp biomicroscopy (SL-14, Kowa Company, Tokyo, Japan) revealed moderate conjunctival hyperemia, chemosis, and mucoid discharge in both eyes. The upper eyelid of the OS exhibited conjunctival thickening without ulcerations. Stromal keratitis was observed on the dorsal temporal region of the same eye, with loss of limbal structure in this area. Additionally, thickening of the iris on the dorsal temporal region of the OS was observed (Figure 1-a). The cornea of the OD exhibited mild edema and neovascularization; moderate aqueous flare, and thickening of the iris with a pseudotumoral appearance, which was more intense on the dorsal temporal region were seen (Figure 2-a). Intraocular pressure (IOP) was assessed with applanation tonometry (Tonopen-XL, Mentor O & O, Inc.) and was 11 and 12 mmHg for the OD and OS, respectively. After pupil dilation with 1% tropicamide (Mydriacyl®) no abnormalities were seen in the lens, vitreous, and the retina of both eyes. Fluorescein dye test was negative in both eyes. Ocular ultrasound (MyLab™Five Vet; Esaote Pie Medical, Genova, Italy) performed with a 10 MHz linear transducer revealed homogeneous hyperechoic thickening of the iris in the OD, and absence of alterations in the remaining intraocular and retrobulbar structures.

Results of complete blood cell count, serum levels of alanine aminotransferase (ALT), albumin, creatinine, and urea, were within the reference range for the species. Thoracic radiographs, abdominal ultrasound, and echocardiogram were unremarkable. Anti-*Toxoplasma gondii* specific IgG and FIV/FeLV rapid test (Alere TM®, Bionote, Gyeonggi-do, Korea) were negative.

Based on the clinical history and ocular examination that revealed anterior uveitis and thickening of the iris, uveal neoplasm was suspected, and anterior chamber paracentesis was suggested to confirm the diagnosis. Following general anesthesia with propofol, 0.2 mL of aqueous humor was collected from both of eyes. Aqueous humor cytopathology stained with modified Wrights stain revealed mixed inflammatory cell content but neoplastic cells were not detected.

Ocular therapy was initiated with 0.3% ketorolac trometamol (Acular®) and 1% prednisolone acetate (Predfort®) every 6h, for 10 days. In the first recheck the patient showed no signs of improvement of the ocular signs. A biopsy of the eyelid and palpebral conjunctiva of the OS, and a new anterior chamber paracentesis of both eyes was suggested for further investigation. In addition, blood sample was collected from the jugular vein, and bone marrow sample was collected from the sternum.

Since general anesthesia was required, the owner requested elective ovariohysterectomy. During the procedure, enlarged mesenteric lymph node was detected and a biopsy specimen was collected, placed in 10% neutral buffered formalin and submitted for histological analysis.

The sections of the eyelid and palpebral conjunctiva stained with H&E revealed the presence of inflammatory infiltrate on the superficial dermis composed mainly of histiocytes, with basophilic intracytoplasmic and extracytoplasmic amastigotes compatible with the protozoan *Leishmania* spp. (Figure 3-a). Examination of the mesenteric lymphnode sample stained with H&E revealed many macrophages containing multiple amastigotes that were also compatible with the protozoan *Leishmania* spp. (Figure 3-b).

In order to confirm the involvement of *Leishmania* spp. as a causative agent of anterior uveitis, samples of aqueous humor, bone marrow, and blood were seeded in biphasic NNN medium, and promastigotes were isolated from all samples. Additionally, PCR from samples of aqueous humor, blood, and bone marrow revealed the presence of *Leishmania* spp. DNA. Sequencing showed a 99% homology to a sequence of *L. infantum* in GenBank (accession number XR_001203206.1, FR799614.1, GQ332356.1, M81430.1) (Figure 4).

Once the diagnosis of leishmaniasis was established, treatment with allopurinol (10mg/kg/ BID/PO) was commenced and a natural insect repellent was prescribed. The previous ophthalmic treatment was continued, once improvement of the ocular signs were not observed until the second recheck, 10 days latter.

In the third recheck, 15 days from the beginning of the leishmaniasis specific treatment, the iris slown of the OD and the chemosis of the OS showed slighty improvement. Mild aqueous flare was still observed in both of eyes and IOPs were 13 and 12 mmHg for the OD and OS, respectively. Topical treatment was maintained for 3 more months, with ophthalmic evaluations performed every 15 days, once the owner reported that the patient exhibited blepharospasm and loss of appetite when the ocular medications were withdrawn. Following 30 days of treatment with allopurinol and topical treatment, the OD developed mild iris atrophy but remained calm (Figure 1-b), and the OS showed improvement of conjunctival thickening, loss of limbal delimitation, still exhibited intense corneal vascularization, and corneoscleral melanosis (Figure 2-b). At this time, IOP were 10 and 13mmHg for the OD and OS, respectively.

In the last recheck, which occurred 6 months later from the first appointment, the prednisolone acetate eye drops (Predfort®) was still being used every 12h. During this period, complete blood count revealed leukopenia (5000 mm$^3$) and thrombocytopenia (78,000/mm$^3$). Renal and hepatic function test results were within the normal range. Specific tests for leishmaniasis were performed: enzyme-linked immunoassay (ELISA) was non-reagent and indirect immunofluorescence (IFI) test was considered negative (<1:40). No signs of anterior uveitis were observed in both of eyes at this time and IOP was 11 and 12mmHg for the OD and OS, respectively.
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Figure 1. Ophthalmic findings of feline leishmaniasis. A- oculus sinister. Upper eyelid conjunctival thickening (black arrow). Note non-ulcerative keratitis with intense neovascularization (white arrow) associated with moderate uveitis. B- left eye after 3 months of ocular treatment. Note improvement of conjunctival thickening, loss of limbal delimitation, still exhibiting intense neovascularization, and corneoscleral melanosis (white arrow).

Figure 2. A - oculus dexter. Pseudotumoral appearance of the iris on the dorsal temporal region (black arrow). Note severe uveitis with conjunctival hyperemia, moderate flare, and miosis. B- right eye after 3 months of ocular treatment. Significant improvement of uveitis, remission of iridic neovascularization and development of mild iris atrophy (white arrow).

Figure 3. A- Histological section of palpebral conjunctiva of the oculus sinister. Presence of inflammatory infiltrate composed mainly of histiocytes in the superficial dermis of the eyelid, with basophilic intracytoplasmic and extracytoplasmic amastigotes compatible with the protozoan Leishmania spp. (arrow). B- Mesenteric lymph node containing a large number of macrophages with multiple intracytoplasmic amastigotes compatible with the protozoan Leishmania spp. (arrow).
Discussão

A possibilidade de encontrar amastigotes de Leishmania spp. em gatos através de citologia ou histologia de fígado, rins, linfonodos, estômago, intestino grosso, fígado, mucosa oral, exsudato nasal e tecidos oculares foi reportada (Leiva et al., 2005, Sobrinho et al., 2012, Pennisi et al., 2015). No entanto, em muitos gatos infectados por leishmaniose, os sinais clínicos correlacionados com outras causas incluem infecção retroviral (FIV e FeLV), tratamento imunossupressor e doenças concomitantes de apariência debilitante, como neoplasia maligna, diabetes melitus, toxoplasmose, e bartonelose (Hervás et al., 2001, Leiva et al., 2005, Sobrinho et al., 2012, Pennisi et al., 2015). Em estudos anteriores, os pacientes apresentaram outros sinais em adição aos sinais oculares, como estomatite, anorexia, dermatite, e alopecia em certas áreas do corpo (Hervás et al., 2001, Leiva et al., 2005).

Os amastigotos foram encontrados por citologia em amostras conjuntivais (Navarro et al., 2010) e corneos (Leiva et al., 2005), trato uveal (Hervás et al., 2001, Leiva et al., 2005, Sobrinho et al., 2012, Pennisi et al., 2015). Em detalhes, estas doenças foram excluídas pelo exame físico e pelas provas laboratoriais.

Nos casos descritos aqui, a manifestação ocular estava exclusivamente ophthalmica, principalmente uveíte e blefarite, tendo sido descritos na literatura (Verneuil, 2013, Ritcher et al., 2014). No entanto, em outros relatos, os pacientes apresentaram outros sinais em adição aos sinais oculares, como estomatite, anorexia, dermatite, e alopecia em certas áreas do corpo (Hervás et al., 2001, Leiva et al., 2005).

In contrast to other cases of uveitis caused by feline leishmaniasis, our patient did not develop increased IOP and secondary glaucoma (Hervás et al., 2001, Leiva et al., 2005).
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et al., 2001, Leiva et al., 2005, Verniuil, 2013, Ritcher et al., 2014). Pennisi et al. (2015) reported that ocular manifestations such as uveitis, blepharitis, and conjunctivitis have been described in cases of feline visceral leishmaniasis that usually exhibit other systemic clinical signs. However, in the present report, only bilateral uveitis unresponsive to anti-inflammatory therapy was observed. The patient did not exhibit any other apparent clinical sign 3 months after the diagnosis of leishmaniasis. Blepharitis and keratoconjunctivitis have also been described as the most frequent lesions in cats with leishmaniasis (Navarro et al., 2010, Migliazzo et al., 2015); however, granulomatous anterior uveitis with pseudotumoral appearance was the predominant clinical sign in this report and in others (Hervás et al., 2001, Leiva et al., 2005, Verniuil, 2013, Ritcher et al., 2014).

Long treatments with allopurinol or subcutaneous injections of meglumine antimoniate have been routinely used for cats with leishmaniasis despite the lack of information on the pharmacokinetics of these drugs (Pennisi et al., 2015). Although such agents are well tolerated, frequent monitoring of renal and hepatic function is imperative during management of the disease (Pennisi et al., 2015). Following 3 months of treatment with allopurinol and topical anti-inflammatory therapy, renal and hepatic functions of the patient were within normal ranges. Nonetheless, complete blood count revealed leukopenia and thrombocytopenia, which are commonly found in dogs but not in cats with this disease (Pennisi et al., 2015). In the last recheck, the patient tested negative to ELISA and IFI were <1:40, which is considered negative.

Since there are no studies regarding the use of vaccines for leishmaniasis in cats, the best prevention strategy for this species is the use of topical insecticides with repellent activity similar to those applied on dogs. Unfortunately, permethrin and deltamethrin should not be used in cats owing to their toxicity (Pennisi et al., 2015). In the case reported here, the patient was using a low toxicity natural citronella phlebotomine repellent formulated for animals, which is used as a topical repellent in humans; however, its efficacy still warrants further investigation in cats (Warburg and Faiman, 2011).

Allopurinol therapy has been reported to significantly aid in the remission of clinical signs of uveitis in a cat with leishmaniasis (Ritcher et al., 2014). In the report of Ritcher et al. (2014), the patient exhibited ocular improvement 2 months following allopurinol therapy and topical medication (prednisolone combined with dorzolamide and timolol). In the present report, OS of our patient did not show signs of improvement of uveitis, despite of maintenance of ophthalmic eye drops (ketorolac trometamol and prednisolone) combined with systemic allopurinol. In cats, there are no studies concerning the effects of allopurinol on intraocular inflammation caused by infection by Leishmania spp.

In conclusion, this report presents the first description of leishmaniasis with exclusively ocular clinical manifestation in a cat in Brazil. Furthermore, this is the first confirmed report of leishmaniasis in a cat by aqueous humor culture and PCR, which highlights the importance of these methods as supportive tools for the diagnosis of leishmaniasis – associated uveitis in cats.

REFERENCES


