Tracheal suture in rats with hypothyroidism. Wound healing study

Traqueorrafia em ratos com hipotireoidismo. Estudo da cicatrização

Elise Zimmermann¹, Jurandir Marcondes Ribas-Filho II, Osvaldo Malafaia II, Carmen Austrália Paredes Marcondes Ribas II, Paulo Afonso Nunes Nassif I, Edmar Stieven Filho III, Paulo Eduardo Przysiezny IV

¹Master, Principles of Surgery Post-Graduate Program, Evangelical Medical School, Curitiba - PR, Brazil.
²PhD, Full Professor, Post-Graduate Program, Evangelical Medical School, Curitiba - PR, Brazil.
³Master, Post-Graduate Program, Evangelical Medical School, Curitiba - PR, Brazil.
⁴Master, Buccomaxilofacial Surgery, Marilia University, Marilia -SP, Brazil.

ABSTRACT

Purpose: To evaluate the influence of hypothyroidism in tracheal wound healing process. Methods: A controlled study was designed with 48 male Wistar rats, divided into two groups: study and control groups. In the first one hypothyroidism was surgically induced and 10 weeks after, a tracheal opening followed by suture was performed in both groups, subdivided into 7, 14, and 21 days in accordance with the date of animals death. A laboratorial evaluation was performed to prove the decreased in thyroid function in the study group. Also a macroscopic evaluation through a stablished protocol and a microscopic analysis with Hematoxylin-eosin and Sirius-Red staining methods were done. Results: The laboratorial evaluation certified suppressed thyroid function in the study group. The macroscopic evaluation showed the presence of suture blockade in the study group in all the evaluated days. Microscopic analysis showed a prolongated inflammatory process and less collagen with delay in organization in the study group comparing to control group. All these data were statistic significant. Conclusion: Hypothyroidism had an influence in tracheal wound healing process, promoting delay in the inflammatory and organization processes and diminished tissue collagen quantity.

Key words: Wound Healing. Hypothyroidism. Trachea. Rats.

RESUMO


Introduction

Wound healing is a dynamic process that usually starts after tissue wound. It alternates anabolic and catabolic reactions administered by a great variety of cellular and molecular events that interact among themselves in order to start the wound healing process.

For didactic purposes, this reparation process may present three distinct superposed stages named: inflammatory stage, proliferative stage, and remodeling of the matrix formation stage. The retardation or failure in one of these stages may slow down or even hinder the process of wound healing. However, from the clinical point of view, the formation and matrix remodeling stage is the most important stage.

From the functional point of view, no matter what kind of tissue lesion, collagen is the most important component of tissue cicatrization. It is the standard, the quality and total quantity of collagen in the matrix that determine the resistance of the scar. In the course of cicatrization, these collagen fibers get thicker, ranging along the power lines of the wound in the process of remodeling and maturation, offering more resistance to the cicatrical tension.
Alterations in the cicatricial process have been described in patients who presented hypothyroidism gravis, resulting from total thyroidectomy or from cervical radiotherapy4,5.

Thyroid hormones have an important role in the growth and development of organs and tissues, essential to the maintenance of life quality6.

The hypofunction of the thyroid gland increases the chances of pre and post operatory complications7,8.

Thyroid hormones are associated with the proliferation and secretion of fibroblasts. The suppression of this hormone secretion disturbs the metabolic activation in tissues the in the collagen synthesis, extending from the inflammatory stage to the proliferative stage9.

The cicatrization process requires proteins and energy sources and it has been proved that the administration of thyroid hormone to animals suffering from hypothyroidism induced the synthesis of nucleic acid and proteins, besides increasing mitochondrial oxidative phosphorylation10. Studies with hypothyroid rats demonstrated that there is a compromise in the collagen synthesis decreasing its biosynthesis, collagen concentration, delaying its organization, solubility and metabolism11.

Tracheotomy is one of the most common surgical procedures performed on critical patients.

ICUs (Intensive Care Units) present a rate of complication that varies from 4 to 40%. Tracheal or sub-glottal stenosis is still a controversial subject and only a few studies present factors that could interfere in the answer to proposed therapies12,13.

This work aims at studying the effect of hypothyroidism on tracheal tissue cicatrization in rats.

Methods

This study was performed according to the Brazilian Law on experiments with animals and the normative document of the Brazilian College of Animal Experimentation. The experiment was approved by the Ethics Committee in Animal Research of the Institution. Forty eight male Wistar rats (Rattus norvegicus albinus, Rodentia mammalia) aged between 100 and 130 days, weighing between 288 and 444 grams, coming from the Animal House of the Technology Institute of Paraná were used in this study. They were kept in polypropylene boxes under constant temperature of 20 + 2 °C, at room humidity, with light cycles each 12 hours. The animals had free access to water and standard lab rat food.

The animals in the study group, after undergoing thyroid and parathyroidectomy, were fed with pentahydrated calcium lactate at 2% and water ad libitum.

The animals were identified with picric acid and divided in groups A (Control) and B (Study). Group A animals were submitted to simulate cervicotomy with digital manipulation of the thyroid gland, while Group B animals underwent thyroid parathyroidectomy.

Ten weeks after the surgical procedures, all the animals were submitted to tracheotomy and tracheal suture between the third and fourth tracheal rings. Animals in groups A and B were divided in six subgroups of eight animals denominated groups A1, A2, A3 and B1, B2, B3 to be euthanized, respectively after 7, 14 and 21 days after the tracheal procedure. The tracheal opening was performed under direct visualization, with the help of a surgical magnifying glass. A transversal incision, shorter than the tracheal diameter was made. Afterwards, a tracheal suture with separate 25 poliglecaprone stitches was made, followed by the approximation of the muscular plane with 4.0 poliglactina thread, the skin and subcutaneous tissue synthesis, with 4.0 nylon monofilament thread (Figure 1). After the surgical procedures, the animals were kept in individual cages in a heated room until they were completely recovered from anesthesia.

After each surgical procedure, 40mg/mL per 2mg/kg weight paracetamol was used as analgesic.

For euthanasia, the rats were placed in a closed bell jar for the inhalation of 97% sulfuric ether lethal vapor. At the moment of the animals’ death, a cardiac puncture with a 25 X 7 needle was made, withdrawing 3mL of blood in order to measure TSH ultra-sensitive, total thyroxine and ionized calcium (ion selective electrode method), taking as reference values the results of the control group analyses.

For the macroscopic investigation not only the presence of adherences but also the presence of abscesses, dehiscence and fistulas were considered. The clinical findings were transformed in numerical parameters. For the tracheal wound 0 (zero) was established, if it presented good healing (cicatrized wound without dehiscence); 1 (one) for doubtful healing (presence of abscess) and 2 for the presence of dehiscence, if it occurred and 1 (one) if it didn’t occur. It was also established 0 (zero) for the absence of fistulas and 1 (one) for the presence of fistulas.
After macroscopic evaluation, the entire trachea was carefully resected up to the carina. Specimens were kept in flasks containing 10% formaldehyde solution for at least 24 hours and then they were prepared for the histological study. They were kept in paraffin blocks and 4 micrometers thick histological cuts were made which were stained with Hematoxylin-eosin and Sirius-red. These samples were labeled with specific codes, known only to the team who performed the surgical procedure.

The histological cuts stained with Hematoxylin-eosin were intended to the evaluation of the type and intensity of the inflammatory reaction in the presence of fibrin leukocytic scab (fibrin and blood cells on the surface of the wound, clot dehydration); angiofibroblastic proliferation (granulate tissue formation, of the neo-vascularization and proliferation of fiberblasts); edge re-epithelization (moving of the epithelium over the wound); foreign body reaction (granulomatous aggregation with giant cells around the suture thread and the presence of microphages); micro-abscess (tissue necrosis in the presence of neutrophils and macrophages). These parameters were classified according to their intensity: absent, discrete and moderate and were scored in order to be transformed in quantitative variables.

For the evaluation of the area, density and quantification of types I and III collagens, the histological cuts were stained with Sirius-red. Samples were analyzed using the Image Pro Plus Application, connected to a Sony video camera and a BX 50 microscope, at magnification of 200, taking into consideration not only the total area (in micra) but also the collagen types I and III percentages. The thicker and strongly birefringent fibers were stained orange and red (type I) and the thinner and less birefringent fibers were stained yellow and green (type III). An average of these percentages was obtained in each slide and the total collagen value corresponded to the sum of the red and green areas.

Two distinct cuts (inferior and superior) 200 times augmented were analyzed and the best resolution of each polarized image with Olympus U-Pot lens, turned anti clock wise until polarization was selected. The representative values of the area (in micra) and of collagen I and III percentage were transported to Microsoft® Excel for statistical analysis.

Comparing control and study groups, in relation to the quantitative variables, Student’s t test was used for independent samples, taking into consideration the evaluation of the homogeneity of the variables. In relation to the nominal dichotomous variables, the groups were compared using Fisher’s exact test. The variables’ normality condition was evaluated using the Shapiro-Wilks test. Values from P<0.05 were statistically significant.

### Results

#### Laboratorial evaluation

The ultrasensitive TSH dosage in Group I presented values that varied between 38, 47 and 51,03µUI/mL, and the control group presented values between 1,75 and 2,02µUI/mL (Figure 2).
Total thyroxin evaluation (T4) analyzed, using the same method, varied in the study group between 0,29 and 0,43 mg/dL and in the control group between 2,50 and 3,70mg/dL (Figure 3).

The values of ionic calcium evaluation in the Study Group varied between 0,86 and 1,02 mmol/L and in the Control Group between 0,94 and 1,15 mmol/L.

The Study Group presented TSH levels above those of the reference values and T4 below them. The control Group maintained itself within normality. There was no significant difference between both groups in relation to ionic calcium values which proves hypothyroidism in the Study group and indicates that the calcium reposition was effective in this group.

**Macroscopic evaluation**

Considering wound and fistula variables, the macroscopic evaluation showed no significant statistical difference between both groups.

As to the blockade variable, it predominated in the Study Group during death, presenting statistically significant differences with P<0,05 (Figure 4).
Microscopic evaluation

Evaluation considering Hematoxilin-eosin coloration

Considering the analyzed variables, the proliferation of fibrin and leukocytes predominated in the study group at the 14th day of evaluation with $P=0.01$ (Figure 5). The other variables did not present statistically significant difference.

Evaluation considering Sirius-red coloration

The evaluation of the area showed the predominance of collagen in the Control Group compared to the Study Group in all evaluated phases, standing out the 1x1 evaluation on the 7th day, 3x3 on the 14th and 21st days, with $P<0.05$ (Figure 6).
The evaluation of the density variable indicated that there was a statistically significant difference between the groups at the 14th day in the 1x1 evaluation and on the 21st day in the 3x3 evaluation (Table 1).

<table>
<thead>
<tr>
<th>Period</th>
<th>Density</th>
<th>Group</th>
<th>Average</th>
<th>Median</th>
<th>Minimum</th>
<th>Maximum</th>
<th>SD</th>
<th>P* value</th>
</tr>
</thead>
<tbody>
<tr>
<td>7th day</td>
<td>3X3</td>
<td>Control</td>
<td>0.220</td>
<td>0.219</td>
<td>0.182</td>
<td>0.258</td>
<td>0.026</td>
<td>0.625</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>0.225</td>
<td>0.216</td>
<td>0.199</td>
<td>0.259</td>
<td>0.021</td>
<td>0.680</td>
</tr>
<tr>
<td>14th day</td>
<td>3X3</td>
<td>Control</td>
<td>0.209</td>
<td>0.224</td>
<td>0.164</td>
<td>0.246</td>
<td>0.029</td>
<td>0.034</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>0.184</td>
<td>0.186</td>
<td>0.159</td>
<td>0.217</td>
<td>0.019</td>
<td>0.161</td>
</tr>
<tr>
<td>21st day</td>
<td>3X3</td>
<td>Control</td>
<td>0.237</td>
<td>0.236</td>
<td>0.161</td>
<td>0.321</td>
<td>0.057</td>
<td>0.760</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>0.243</td>
<td>0.266</td>
<td>0.158</td>
<td>0.306</td>
<td>0.054</td>
<td>0.020</td>
</tr>
<tr>
<td></td>
<td>1X1</td>
<td>Control</td>
<td>0.280</td>
<td>0.281</td>
<td>0.219</td>
<td>0.351</td>
<td>0.049</td>
<td>0.020</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>0.325</td>
<td>0.322</td>
<td>0.276</td>
<td>0.413</td>
<td>0.044</td>
<td>0.020</td>
</tr>
</tbody>
</table>

(*) Student’s t Test for independent samples P<0,05

As to the evaluation of the relation collagen I and III, the predominance of collagen III was found in both groups at the 7th day evaluation. At the 14th and 21st days, collagen I predominated in both groups. However, on the 21st day it was noticed that the ratio collagen I: III was higher in the Control Group when compared to the Study Group. This information was statistically significant (P=0,007) (Table 2).

<table>
<thead>
<tr>
<th>Period</th>
<th>Collagen</th>
<th>Group</th>
<th>Average</th>
<th>Median</th>
<th>Minimum</th>
<th>Maximum</th>
<th>SD</th>
<th>P* value</th>
</tr>
</thead>
<tbody>
<tr>
<td>7th day</td>
<td>I</td>
<td>Control</td>
<td>27.91</td>
<td>26.66</td>
<td>14.98</td>
<td>43.16</td>
<td>10.16</td>
<td>0.548</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>25.96</td>
<td>26.71</td>
<td>17.98</td>
<td>31.63</td>
<td>5.61</td>
<td>0.548</td>
</tr>
<tr>
<td></td>
<td>II</td>
<td>Control</td>
<td>72.09</td>
<td>73.34</td>
<td>56.84</td>
<td>85.02</td>
<td>10.16</td>
<td>0.550</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>74.04</td>
<td>73.29</td>
<td>68.37</td>
<td>82.02</td>
<td>5.61</td>
<td>0.550</td>
</tr>
<tr>
<td>14th day</td>
<td>I</td>
<td>Control</td>
<td>60.72</td>
<td>61.85</td>
<td>47.50</td>
<td>73.34</td>
<td>9.25</td>
<td>0.550</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>58.77</td>
<td>59.74</td>
<td>51.86</td>
<td>66.98</td>
<td>5.34</td>
<td>0.550</td>
</tr>
<tr>
<td></td>
<td>II</td>
<td>Control</td>
<td>39.28</td>
<td>38.15</td>
<td>26.66</td>
<td>52.50</td>
<td>9.25</td>
<td>0.550</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>41.23</td>
<td>40.26</td>
<td>33.02</td>
<td>48.14</td>
<td>5.34</td>
<td>0.550</td>
</tr>
<tr>
<td>21st day</td>
<td>I</td>
<td>Control</td>
<td>64.89</td>
<td>61.99</td>
<td>47.60</td>
<td>84.62</td>
<td>11.40</td>
<td>0.007</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>54.65</td>
<td>56.11</td>
<td>42.57</td>
<td>62.40</td>
<td>6.77</td>
<td>0.007</td>
</tr>
<tr>
<td></td>
<td>II</td>
<td>Control</td>
<td>35.11</td>
<td>38.01</td>
<td>15.38</td>
<td>52.40</td>
<td>11.40</td>
<td>0.007</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Study</td>
<td>45.35</td>
<td>43.89</td>
<td>37.60</td>
<td>57.43</td>
<td>6.77</td>
<td>0.007</td>
</tr>
</tbody>
</table>

(*) Student’s t test for independent samples P<0.05

**Discussion**

Hypothyroidism affects people over 60 and influences protein synthesis. The delay in the cicatrical process due to hypothyroidism has been described since 1967 by Kivirikko and Kowalewsk and Yong who demonstrated that hypothyroidism decreases collagen synthesis and interfere with its metabolism. From then on, there has been a great interest in the study of the influence of hypothyroidism in the cicatrical process and other studies confirm its existence.
Because this work aims at evaluating the cicatricial process due to hormonal influence, we chose male animals in order to exempt a possible action of feminine hormones in the results. The literature registers descriptions of different methods to induce hypothyroidism in rats and, among them, the anti-thyroid drugs, either by irradiation or surgery. In the present study, we chose the surgical method to treat the animals in the study group because the animals presented an efficient and homogeneous condition of hypothyroidism, avoiding collateral effects of either the anti-thyroid drugs or of irradiation.

A period of ten weeks (time considered by most studies as effective to stop the effects of hormone action at the surgical site) was observed in order to certify the hypothyroidism condition. Other authors mention, among the main complications of thyroideectomy, the infection of the surgical wound, hemorrhage, the lesion of the laryngeal nerve and 20% transitory hypocalcemia and 2% permanent hypocalcemia. This may be due to the occurrence of unintentional lesion or surgical ablation of the parathyroid glands, causing hypoparathyroidism and thyrotoxic storm.

This possibility motivated calcium reposition in the animals subjected to thyroideectomy and a subsequent calcium laboratorial dosage in the sample. Chlorhexidine gluconate was used in the antisepsis because it does not interfere with thyroid metabolism.

Patients with severe hypothyroidism present chronic peripheral vasconstriction. This way, ketamine chloride was added to the process of anesthesia because it provides a positive inotropic effect, discretely elevates blood pressure and avoids some undesired effects of hypotention, such as the decrease of tissue perfusion that would hinder cicatrisation.

For tracheal opening, Manna’s experimental model confirmed by Mendez-Picon et al. was used. They described a microsurgical model of tracheotomy in adult rats in order to study three types of tracheal opening. They concluded that a transversal incision presented less morphological alterations when compared either with the longitudinal section or the excision of a segment.

As to the tracheal suture, a large bibliography on the subject was consulted, starting with Grillo that considered Vicryl® thread appropriate to tracheal sutures. Recently, Trimbos presented similar results comparing Vicryl® and Monocryl® threads in rat tracheal anastomosis. Monocryl® presented the best results of initial resistance, relatively quicker absorption and low inflammatory reaction. The microscopic analysis showed significant statistic difference between the groups only as to angio fibrin leukocyte proliferation which was longer and more intense in the study group. This demonstrates that hypothyroidism influences cicatrisation, prolonging the inflammatory phase.

The Sirius-red histological study centered in the collagen area and density and in the quantity of types I and III collagen, used a microscope with polarized lens. According to Shimizu, animals presenting hypothyroidism presented lesser connective tissue fibers than their healthy counterparts. During the proliferative phase, fibroblasts increased, with the predominance of the new tissue fibers than their healthy counterparts. During the proliferative phase, fibroblasts increased, with the predominance of the new collagen synthesis. The granulation tissue presented types I and III collagen; however, type I is the most abundant component. Mature dermis has 80% type I collagen and 20% type III (Zhang). Tissue resistance and integrity result from the power established among the collagenous fibers and they depend on their size and structure. Resistance depends more on type I collagen, the strongest fibers. Tissue resistance and integrity result from the power established among the collagenous fibers and they depend on their size and structure. Resistance depends more on type I collagen, the strongest fibers. Tissue resistance and integrity result from the power established among the collagenous fibers and they depend on their size and structure. Resistance depends more on type I collagen, the strongest fibers.

Macroscopic and microscopic analyses with hematoxilin-eosine and Sirius-red did not permit a thorough evaluation of the tracheal scar. This way, the tests were complementary as to morphological and functional aspects of cicatrisation, estimating not only the quality but also the resolution of the lesion.

**Conclusion**

Hypothyroidism influences the process of tracheal cicatrisation, prolonging the inflammatory phase, reducing collagen and delaying its maturation and organization.
References


Conflict of interest: none
Financial source: none

How to cite this article
Available from URL: http://www.scielo.br/acb