Does acute alcohol intoxication interfere with colonic anastomosis wound healing? A rat model of nondestructive colon trauma

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ABSTRACT

PURPOSE: To evaluate the effects of acute alcohol intoxication on healing of colonic anastomosis.

METHODS: Thirty-six rats were allocated into two groups. Animals in the alcohol (A) were given 2 mL of ethanol diluted in 0.9% saline solution to a concentration of 40% by gavage immediately before anesthesia, whereas control (C) animals received 2 mL of 0.9% saline solution via the same route. A colonic anastomosis was then performed in all animals. On postoperative days 1, 3, and 7, anastomotic breaking strength was assessed and histopathological examination was performed. Change in body weight and mortality were also evaluated.

RESULTS: The median of anastomotic tensile strength on the postoperative day 1 was 0.09 Newtons for group A and 0.13 for group C. (p>0.05). The median of anastomotic tensile strength on the postoperative day 3 was 0.13 Newtons for group A and 0.17 for group C. (p>0.05). The median of anastomotic tensile strength on the postoperative day 7 was 0.30 Newtons for group A and 0.35 for group C. (p>0.05). There was no significant difference between the groups A and C, in the first, third or seventh POD (p>0.05), in any of the analyzed parameters. There were no statistical differences between groups in the weight. Three animals died, all from the group A.

CONCLUSION: Acute alcohol intoxication did not interfere with wound healing of colonic anastomoses, although it caused early postoperative mortality.

Key words: Wound Healing, Alcoholism, Ethanol, Tensile Strength, Rats.
Introduction

Preoperative assessment has become part of daily practice before surgery of the colon and other organs, but alcohol abuse has not received much attention. Acute alcohol exposure represents a risk factor for increased morbidity and mortality associated with surgical or traumatic injury in human patients. After colonic anastomosis surgery, the prognosis of the patient depends to a large extent on healing of the colonic wound.

Inappropriate wound healing of colonic anastomosis can lead to complications such as dehiscence and leakage, which can result in catastrophic consequences; the mortality rate following anastomotic leak ranges from 6.0–39.3%.

The overall leakage rate after colonic anastomosis usually ranges from 3.4% to 6% in abdominal trauma, the risk of this complication after anastomosis is higher, ranging from 7% to 14% in low-risk patients up to 40% in high-risk patients.

The prevalence of alcohol intoxication in trauma patients at the time of hospital admission ranges from 18% to 80%.

Higher blood alcohol levels have been found to correlate with more severe abdominal injuries, including colonic injuries, in trauma patients. However, the same study found that alcohol intake was not an independent risk factor for worse outcome, overall injury severity, or posttraumatic complications in patients with multiple trauma.

Primary repair is considered the optimal technique for nondestructive colon injury at most institutions, regardless of risk factors. We hypothesized that acute alcohol exposure shortly before surgery would be a risk factor for impaired wound healing of the colonic anastomosis, based on results from other experimental studies. However, in those studies, alcohol exposure followed different patterns.

The effects of alcohol abuse on patient prognosis and on experimental wound healing are highly controversial, because they depend on the type of exposure and on the presence of associated factors such as sepsis.

The purpose of this study was to evaluate the effects of acute alcohol intoxication on colonic anastomosis wound healing and postoperative outcomes in rats.

Methods

All procedures were performed in accordance with the ethical standards proposed by the Brazilian College of Animal Experimentation (COBEA). Approval was obtained from the Animal Research Ethics Committee of the University of Brasilia, Brazil, prior to the study.

This was a randomized, single-blind experimental study. Thirty-six male Wistar rats were randomly allocated into two groups of 18 animals each: an experimental group that received alcohol (A) preoperatively and a control group (C) that did not. Each group was subdivided into three subgroups according to the timing of euthanasia (postoperative day 1, 3, or 7), yielding groups 1A, 3A, 7A and 1C, 3C, 7C (Figure 1).

On the day of surgery, rats were fasted for one hour. Animals in group A received 2 mL of ethanol diluted in 0.9% saline solution to a concentration of 40%, given by mouth using the gavage method. This dose is equivalent to an intake of 480 mL of spirits (approximately 10 shots) in a young adult man weighing 75 kg. Half of the dose (1 mL) was given at once and the remaining 1 mL was given 1 hour later, also by mouth, immediately before anesthesia. Surgeons were blinded to group allocation, and all surgeries were performed by the same team. Control group animals received 2 mL of 0.9% saline solution using the same protocol.

Anesthesia was induced by intramuscular injection of xylazine (10 mg/kg) and ketamine (75 mg/kg). After induction, the abdomen was disinfected with iodinated detergent.

A midline abdominal incision approximately 4.5 cm in length, starting 1 centimeter cranial to the symphysis pubis, was made. A 1-centimeter segment of the left colon was resected, and an end-to-end anastomosis was performed with single-layer running sutures using 6-0 polypropylene. Abdominal wall closure was performed with running sutures, in two layers, using 3-0 polypropylene. Postoperative analgesia was provided with tramadol (0.72 mg/kg every 12 hours).

On postoperative day 1, 3, or 7, according to subgroup allocation, the animals were weighed and anesthetized. Colon specimens were collected and the animals were killed with an overdose of thiopental by intravenous injection.
Anastomotic tensile strength (ATS) was evaluated with a VersaTest motorized test stand (Mecmesin VersaTest, United Kingdom), coupled to a digital dynamometer to which the colonic anastomosis samples were attached (Figure 2). The maximum traction force that the tissue could withstand before rupture was measured in Newtons (N).

A sample of the anastomotic scar tissue was collected for histopathological analysis, fixed in formalin, and stained with hematoxylin and eosin. The amount of collagen, fibroblast, mononuclear and polymorphonuclear infiltration and neovascularization was graded qualitatively on a scale of 0 to 3, where 0 = absent and 3 = marked. Abscess, bacterial colonization, foreign body, crust, and fibrin formation were scored dichotomously as 0 or 1, meaning absent or present, respectively.

Results were analyzed in SPSS (Statistical Package for the Social Sciences) 18.0. Parametric and nonparametric tests were used as appropriate according to the nature of the variables. The paired-samples t-test was used for comparison of body mass, and the Kruskal–Wallis test for anastomotic tensile strength. Fisher’s exact test was used for statistical analysis of all histopathological variables. Significance was set at p<0.05.

Results

**Anastomotic tensile strength (ATS)**

Among animals killed on postoperative day 1 (blue boxes, Figure 3), group 1C (controls) had a median ATS of 0.09 N (interquartile range 0.15 N), whereas group 1A had a median ATS of 0.13 N and an undetermined interquartile range. There were no significant difference between these groups (p>0.05).

Among animals killed on postoperative day 3 (green boxes, Figure 3), group 3C had a median ATS of 0.17 N (interquartile range 0.17 N), and group 3A, a median ATS of 0.13 N (interquartile range 0.16 N). Again, there was no significant difference between these groups (p>0.05).

Among animals killed on postoperative day 7 (beige boxes, Figure 3), group 7C had a median ATS of 0.35 N (interquartile range 0.26 N), and group 7A, a median ATS of 0.3 N (interquartile range 0.33 N). There was no significant between-group difference (p>0.05).

**Histopathology**

There were no significant differences in any of the analyzed parameters between groups C and A, whether on postoperative days 1, 3, or 7 (p>0.05) (Table 1).
TABLE 1 - Sum of scores assigned for each histopathological parameter.

<table>
<thead>
<tr>
<th>Histopathology findings</th>
<th>A1</th>
<th>C1</th>
<th>A3</th>
<th>C3</th>
<th>A7</th>
<th>C7</th>
</tr>
</thead>
<tbody>
<tr>
<td>n (survived)</td>
<td>3</td>
<td>6</td>
<td>6</td>
<td>6</td>
<td>6</td>
<td>6</td>
</tr>
<tr>
<td>Collagen</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>6</td>
<td>6</td>
</tr>
<tr>
<td>Fibroblast</td>
<td>0</td>
<td>0</td>
<td>6</td>
<td>6</td>
<td>18</td>
<td>18</td>
</tr>
<tr>
<td>Mononuclear</td>
<td>0</td>
<td>0</td>
<td>6</td>
<td>6</td>
<td>12</td>
<td>12</td>
</tr>
<tr>
<td>Polymorphonuclear</td>
<td>5</td>
<td>8</td>
<td>14</td>
<td>12</td>
<td>13</td>
<td>15</td>
</tr>
<tr>
<td>Neovascularization</td>
<td>0</td>
<td>0</td>
<td>6</td>
<td>6</td>
<td>12</td>
<td>12</td>
</tr>
<tr>
<td>Abscess</td>
<td>1</td>
<td>1</td>
<td>4</td>
<td>3</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>Bacteria</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>FB</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>6</td>
<td>5</td>
</tr>
<tr>
<td>Crust</td>
<td>3</td>
<td>6</td>
<td>5</td>
<td>4</td>
<td>2</td>
<td>4</td>
</tr>
<tr>
<td>Fibrin</td>
<td>3</td>
<td>5</td>
<td>6</td>
<td>6</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

Body weight

Overall, the mean preoperative body weight was 310 ± 7.85 g, and the mean postoperative body weight, 291 ± 7 g. The only animals that did not lose weight postoperatively were those in group 1A. All other subgroups lost weight between postoperative day 0 and the time of euthanasia (Table 2).

TABLE 2 - Mean pre and postoperative body weight in each group.

<table>
<thead>
<tr>
<th>Body weight per group</th>
<th>Preoperative</th>
<th>Postoperative</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>1A</td>
<td>303</td>
<td>307</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>1C</td>
<td>337</td>
<td>322</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>3A</td>
<td>315</td>
<td>286</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>3C</td>
<td>310</td>
<td>284</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>7A</td>
<td>288</td>
<td>272</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>7C</td>
<td>302</td>
<td>281</td>
<td>&lt;0.05*</td>
</tr>
</tbody>
</table>

*Mortality rate

There were three deaths, all in group 1A, which corresponds to a 50% mortality rate in this subgroup and 16.6% if all alcohol-exposed animals are considered.

Discussion

Studies evaluating the effects of alcohol on trauma, colon surgery, and wound healing have reported conflicting findings, because the effects of alcohol intake depend on the pattern of exposure (chronic or acute), the amount ingested, the duration of intake, the time since alcohol exposure, the presence of alcohol withdrawal, and other associated factors. Epidemiological studies usually include patients with all types of exposures, and the outcome may depend on the prevailing type.

In this study, we analyzed the effects of acute alcohol intoxication on colon wound healing and prognosis in healthy rats, with no comorbidities, given a large amount of alcohol (in terms of ethanol intake per kg body weight) over a short time just before surgery. This model is representative of a substantial portion of human patients with colonic trauma, as alcohol is involved in up to 80% of incidents causing trauma and most of those patients are not chronic abusers, but rather were binge drinking. In the present study, this type of alcohol exposure did not affect colonic anastomosis healing.

Alcohol is one of most important risk factors for fatal injuries in trauma, contributing to one-third of all deaths from accidents. In traffic incidents, it is responsible for a twofold mortality rate, higher impact speed, greater injury severity, and higher preclinical mortality. In our sample, the mortality rate was 50% in group 1A—which corresponds to early and even preclinical death—versus zero in the control animals.

Experimental and epidemiological studies of chronic alcohol abusers have found alcohol to be deleterious to many aspects of the postoperative period, such as wound healing, reoperation rate, leakage, weight loss, and the metabolic response to trauma and surgery. Although this patient population clearly suffers the effects of alcohol abuse on wound healing, with higher mortality and morbidity rates, these effects are reversible after two weeks to three months.

Pereira et al. has shown that a model of chronic alcohol abuse in rats is associated with lower weight gain, impaired intestinal healing (as demonstrated by significantly lower bowel anastomosis breaking strength), and higher postoperative complication rates.

We did not find a higher risk of colon dehiscence in the treated group. Therefore, emergent primary repair of nondestructive colon injuries in patients with acute alcohol intoxication seems to be safe, corroborating the current practice of most hospitals.

Nevertheless, a previous study by our research group found that colonic anastomosis wound healing is impaired in rats.
with peritoneal sepsis and acute alcohol intake with the same alcohol exposure pattern used in the current study\textsuperscript{12}. This suggests that diversion may be required if the intoxicated patient has fecal contamination and sepsis, as occurs in destructive colon trauma, for instance\textsuperscript{12}. In our previous study, which included two groups (sepsis + alcohol and sepsis alone), the sepsis group exposed to alcohol exhibited poorer wound healing. Therefore, this type of alcohol exposure may act not as an independent risk factor, but as a cofactor.

**Conclusion**

In the exposure pattern used experimentally in this study, acute alcohol intoxication did not interfere with wound healing of colonic anastomoses, although it did increase early postoperative mortality.

**References**


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