EFFECTS OF PROTEIN AND VITAMIN B DEFICIENCY ON BLOOD PARAMETERS AND MYENTERIC NEURONS OF THE COLON OF RATS

Débora de Mello Gonçales Sant’Ana¹, Sônia Lucy Molinar², Marcílio Hubner Miranda-Neto³

ABSTRACT - The aims of this work were to evaluate the effects of the deficient ingestion of protein and vitamin B on the biochemical and hematologic parameters and on the NADH- and NADPH-diaphorase positive myenteric neurons. The control animals (n=10) received commercial chow and the experimental rats (n=10) received chow with protein level reduced to 8% during 120 days. At the time of killing blood was collected for assessment of the blood and hematologic parameters and the ascending colon for quantitative analysis of the neurons of the myenteric plexus. It was observed that the reduction of the protein level to 8% coupled to the reduction of the levels of vitamin B in adult rats neither led to qualitative or quantitative changes on red or white blood cells, nor decreased globulin levels, induced the formation of edema or gave rise to clinical signs typical of protein or vitamin B deficiency. On the other hand, the experimental protocol led to less weight gain, change on the body composition with fat deposition; decrease of the values of serum total protein and albumin; reduction of the area of colon and density of nitrergic and NADH-diaphorase myenteric neurons inferior to the expected.

KEY WORDS: enteric neurons, proteic desnutrition, ascending colon, hematology, vitamin B.

The utilization of animal models in studies of food restriction with known and controlled diets has as an advantage the improved knowledge about the many aspects of the human desnutrition⁴. Research on desnutrition aims at simulating natural conditions, which involve deficiencies not only of protein (20-60% of the amount needed) but also of vitamins and minerals, with increased ingestion of carbohy-

drates⁵. The evaluation of these moderate forms of desnutrition is important because protein deficiency itself leads to a limitation of caloric expenditure with pathological changes that progress through several steps. The rate of evolution of these changes depends on the organic stores of the nutrients and on the adaptative metabolic changes aiming at making up for the deficiency⁶.

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Studies in children with Kwashiorkor demonstrated a decrease in the rate of synthesis of plasma albumin and absence of changes in the synthesis of gama-globulins. In studies in rats it was demonstrated that when fed with a 0.5% protein diet, these animals had a decrease in the plasma levels of albumin and induction of edema. Other authors also found a significant decrease in the albumin levels in rats subjected to desnutrition. Experimental desnutrition is a fruitful area for the investigation of neuronal plasticity and the interdependence of neuronal and non-neuronal mechanisms. Quantitative and morphological evaluations of the neurons of the myenteric plexus in malnourished rats have been demonstrating a larger neuronal density in the small intestine and in the colon, both due to the smaller growth of the animals and their bowels, which lead to lesser neuronal spreading. Most of the authors reviewed offered to the animals diets with protein deficiency but included minerals and vitamins of the complex.

Because desnutrition is often multideficient, the association of protein deficiency with vitamin B deficiency being common, we carried out this work with the purpose of evaluating the results of the ingestion of a starch-rich, but protein- and vitamin B-deficient diet, on the biochemical and hematologic parameters and on the NADH- and NADPH-diaphorase positive myenteric neurons of rats.

**METHOD**

It was used 20 male *Rattus norvegicus* of the Wistar strain aging 90 days (298±66g body weight), which were kept and killed according to the rules of ethical conduct in animal experimentation. The animals were divided in two groups, control and experimental. They were kept in individual cages with constant temperature and light-dark cycles of 12-12 hours. Water and chow were offered ad libitum. The control group received NUVILAB® (Recommended by the National Council of Research and National Institute of Health – USA) chow with protein level of 22% and the experimental group received a modified chow with protein level of 8%, during 120 days.

The diet of the experimental group was obtained reducing the protein level of the NUVILAB® chow through the addition of corn starch and supplemented with a mixture of mineral salts, but without vitamin supplementation. This mode of preparation was based in models described in the literature.

The weight gain of the animals was accompanied weekly, and from the 60th to the 80th day of the experiment they were kept in metabolic cages for daily evaluation of food and water ingestion and body weight.

During the whole experimental period the characteristics of the fur, paws, face and motor behaviour of the animals were observed comparing control and experimental rats.

At the 120th day the animals were weighted and killed through excessive inhalation of ethyl ether. Next laparotomy was carried out and the colon was removed, which were measured in its length and width with a millimetered ruler.

The blood of 10 animals of each group was collected. Through the automatic enzymatic method with heparinized blood the dosage of plasma total protein and albumin was carried out and with the colorimetric method the concentration of globulins was assessed. Hemogram (eritrogram and leucogram) was made with the method of automatic cytometry by Scatter Laser and Electromagnetic. Hematocrit and hemoglobin concentration were also verified.

Retropertioneal fat of five rats of each group was removed and weight in analytical scales. The proportion of this fat relative to body weight was calculated.

The ascending colon of five animals of each group was filled with Krebs solution (pH 7.3), washed twice in the same solution (10 minutes each) and immersed for five minutes in 0.3% Triton X-100 solution dissolved in Krebs. They were washed twice more (10 minutes each) in Krebs solution and incubated for 45 minutes for the evidencing of the NADH-diaphorase enzyme. This solution contained in each dl: 25 ml of 0.5% stock solution of Nitro Blue Tetrazolium (NBT; Sigma, St. Louis, USA); 25 ml of 0.1M phosphate buffer, pH 7.3; 50 ml of distilled water and 50 mg of β-NADH (Sigma, Steinheim, Germany), according to the technique of Gabella. After incubation the segments were opened at the mesocolic margin and immersed in 10% buffered formol solution.

The ascending colon of four animals of each group were washed and filled with phosphate buffer (PBS; pH 7.4), fixed in 4% paraformaldehyde (Merk, Darmstadt, Germany) prepared in 0.1M phosphate buffer (PBS; pH 7.4) for 30 minutes, immersed in 0.3% Triton X-100 (Sigma, St. Louis, USA) dissolved in saline phosphate buffer (PBS, pH 7.4) for 10 minutes and then washed ten times (10 minutes each) in PBS and immersed in the incubation medium for the neuronal evidencing of the NADPH-diaphorase, during two hours. This medium contained in each dl: 25 mg of NBT; 50 mg of β-NADPH (Sigma, Steinheim, Germany), 0.3% Triton X-100 in 0.1M Tris-HCl buffer, pH 6.0 (GibcoBRL, N.Y. USA). After incubation, the segments were opened at the mesocolon insertion and washed three times in PBS for five minutes, and then immersed in 5% paraformaldehyde solution.

The whole-mounts were made under stereomicroscope with trans-illumination through removal of the tunica mucosa and the submucosa. Next they were dehydrated in ascending series of ethyl alcohol, diaphanized in xylene and mounted between slide and coverslip with Permount synthetic resin (Fischer Chemical, New Jersey, USA).
Quantitative analysis was carried out on both techniques using Olympus BX40 microscope under 40X objective. In each whole-mount 80 microscopic fields were counted. Half-seen neurons were counted in alternate fields. The area of each microscopic field was 0.224 mm².

It was calculated the mean, standard deviation and variation coefficient of all the data obtained. After realizing the variation coefficient that the dispersion of the data was small, and thus that the mean was an excellent representative of the values, means were compared using Student’s T test for non-paired data at the significance level of 5%.

RESULTS

At the beginning of the experiment, the mean weight of the animals in the control group was 302.67±20.44g and that of the rats in the experimental group was 296.09±34.96g. At the end of the experimental period the animals of the control group had a mean weight of 456.03±33.48g and those of the experimental group 388.86±46.3g. The difference between the final weights was statistically significant (t=2.74; c.v. =1.67; p<0.05).

In the control animals it was found a mean of 7.43±0.82g of retroperitoneal fat, which corresponded to 1.85±0.23 g/100g body weight. In the experimental animals a mean of 8.7±0.74g was found (t=2.74; c.v. =1.67), corresponding to 2.88±0.31g/100g body weight (t=6.06; c.v. =1.67).

Through the biochemical dosages of the blood of the control and experimental animals it was verified that there was a decrease in the mean value of total protein and serum albumin, as observed in Table 1; however, there were no significant changes in the values of hematocrit and hemoglobin, as well as in the counting of erythrocytes and leucocytes, as can be seen in Table 2.

During the whole experimental period the disnurtured rats showed the extremities of the limbs, fur and motor behaviour similar to those of the controls. During laparotomy, an increase of peritoneal fluid was not observed. In the experimental animals a visibly shorter and thinner colon was noted, being surrounded by adipose tissue as well.

In Table 3 it is observed the differences in the length, width and area of the colon of both groups.

The NADPH-diaphorase positive neurons of the experimental animals showed a density 3.7% greater than that of the control animals, while for the NADH-diaphorase positive neurons this difference reached 27% (Table 4).

<p>| Table 1. Mean blood values of total protein and fractions (g/100 ml) in the plasma of Wistar rats aging seven months from the control and experimental groups. Mean ± standard deviation. |</p>
<table>
<thead>
<tr>
<th>Group</th>
<th>Total Protein* (g/dl)</th>
<th>Albumin (g/dl)</th>
<th>Globulin (g/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (n=5)</td>
<td>6.52 ±0.4 a</td>
<td>3.97 ± 0.53 a</td>
<td>2.6 ± 0.49 a</td>
</tr>
<tr>
<td>Malnourished (n=5)</td>
<td>5.94 ± 0.32 b</td>
<td>3.22 ± 0.58 b</td>
<td>2.72 ± 0.31 a</td>
</tr>
<tr>
<td>Reference value**</td>
<td>6.3-8.6</td>
<td>3.3-4.9</td>
<td>2.4-3.9</td>
</tr>
</tbody>
</table>

*Dosages carried out with the Automatic Enzymatic Method. ** Source:21
Means followed by the same letter for each variable do not differ at the significance level of 5%.
Total Protein (t= 2.9; c.v. 1.86); Albumin (t= 2.25; c.v. 1.80); Globulin (t= 0.47; c.v. 1.86).

<p>| Table 2. Mean values of hemoglobin (g/100 ml), hematocrit (%), number of erythrocytes and leucocytes (million/mm²) of adult Wistar Rattus norvegicus of the control and experimental groups. Mean ± standard deviation. |</p>
<table>
<thead>
<tr>
<th>Group</th>
<th>Hematocrit (%)</th>
<th>Hemoglobin* (g/dl)</th>
<th>Erythrocyte Million/mm³</th>
<th>Leucocytes¹ Million/mm²</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (n=5)</td>
<td>47.2 ± 2.6 a</td>
<td>15.58 ± 0.876 a</td>
<td>8.06 ± 0.78 a</td>
<td>5.4 ± 1.69 a</td>
</tr>
<tr>
<td>Malnourished (n=8)</td>
<td>46.0 ± 2.4 a</td>
<td>15.11 ± 0.11 a</td>
<td>8.27 ± 0.36 a</td>
<td>5.8 ± 2.2 a</td>
</tr>
<tr>
<td>Reference value**</td>
<td>41-52</td>
<td>11.5 - 16.0</td>
<td>5.4 - 8.5</td>
<td>4.0 - 10.2</td>
</tr>
</tbody>
</table>

* Dosages carried out with Automatic Cytometry Method by Scatter Laser and Electromagnetic. **Source:21
Means followed by the same letter for each variable do not differ at the significance level of 5%.
Hemoglobin (t=1.46; c.v. 1.782); Hematocrit (t= 0.85; c.v. 1.782); Erythrocytes (t= 0.69; c.v. 1.782); Leucocytes (t= 0.398; c.v. 1.782).
Table 3. Mean lengths and widths of the total colon of adult rats from the control and experimental groups. Area of total colon based on the measures of length and width.

<table>
<thead>
<tr>
<th>Group</th>
<th>Lengths of total colon (cm)</th>
<th>Widths of total colon (cm)</th>
<th>Area of total colon (cm²) (lengths x widths)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (n=7)</td>
<td>18.34 ± 1.0 a</td>
<td>1.66 ± 0.21 a</td>
<td>30.1 ± 4.35 a</td>
</tr>
<tr>
<td>Malnourished (n=7)</td>
<td>13.6 ± 0.8 b</td>
<td>1.04 ± 0.13 b</td>
<td>13.78 ± 2.46 b</td>
</tr>
</tbody>
</table>

Means followed by the same letter for each variable do not differ at the significance level of 5%.
Lengths (t=11.0; c.v. 1.77); Widths (t=6.7; c.v. 1.77); Area (t=8.72; c.v. = 1.77) [14]

Table 4. Incidence of neurons in the myenteric plexus of the ascending colon of adult rats from the control and experimental groups evidenced by the techniques of NADPH-diaphorase and NADH-diaphorase in 80 microscopic fields (area of 17.92 mm²). Mean ± standard deviation.

<table>
<thead>
<tr>
<th>Group</th>
<th>NADPH-d (n=4)</th>
<th>NADH-d (n=5)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1609 ± 115. 6 a</td>
<td>1464.8 ± 209.2 a</td>
</tr>
<tr>
<td>Malnourished</td>
<td>1671 ± 729.4 a</td>
<td>2006.4 ± 489.1 b</td>
</tr>
</tbody>
</table>

Means followed by the same letter in the same column do not differ statistically at the significance level of 5%. NADPH-d (t= 0.16; c.v. 1.946); NADH (t= 2.25; c.v. = 1.86).
Means followed by the same symbol (•) in each row do not differ statistically at the significance level of 5%. Control (t= 1.23; c.v. = 1.89); Malnourished (t= 0.83; c.v. = 1.89). [14]

**DISCUSSION**

The addition of starch with resultant decrease of the protein and vitamin levels did not induce qualitative and quantitative changes in the red blood cells, which was showed through the counting of the erythrocytes, the hematocrit and the dosage of hemoglobin. Previously it was verified that the deficiency of vitamins from the complex B, which in humans lead to megaloblastic anemia, does not do it in rats. [20]

In this experiment, the value found for the total plasma proteins of the experimental animals (5.94±0.32 g/dl) was lower than the minimum reference value for rats (6.3-8.6 g/dl) but similar to that found in rats receiving diet with equivalent protein level but with vitamin B supplementation (5.88±0.31 g/dl) [17]. Decreases in the values of total protein were also found in rats fed with protein-free diet [6] and with diets containing 0 and 4% protein levels [22].

The mean value of serum albumin (3.22±0.58 g/dl) was slightly inferior to the reference value (3.3-4.9 g/dl) [11], this possibly prevented an apparent interference with the hemodynamic conditions of the animals. Significant reductions of serum albumin in rats in the absence of edema were also found with protein-free diet during 28-32 days [5] and during 84 days [8].

The formation of edema associated to a significant decrease of albumin was found with protein-free diet [22] through a 50% decrease in the amount of ration offered to the animals during gestation and the first six weeks after birth [21]. Many authors stress that the rats are not suited to reproduce this characteristic of Kvarshiokor, because they seldom develop nutritional edema [14].

As for the diet offered by Firmansyah et al. [21] it is implicit that the animals had their protein ingestion reduced to a half, while in the present experiment the decrease in the protein level of the diet was of 63%; despite this, the rats did not lose weight nor suffered a marked decrease in their albumin levels, possibly because proteins were spared and the major energy source was the oxidation of carbohydrates, which were added to the diet so as to keep caloric offer, while the experimental animals of Firmansyah et al. [21], being in a critical period of growth, used protein both structurally and as an energy source due to the lower carbohydrate ingestion, resulting in a reduced protein availability reflected in the tissue and circulating albumin levels.

Although total protein was reduced, the value for globulin (2.52±0.19 g/dl) was within the reference range for this species (2.4-3.9 g/dl) [21]. Normal values for globulin were also found in rats subjected to proteic deficiency (2.72±0.31 g/dl) [17].

The normality in the values for globulin and the absence of changes in the counting of white blood cells indicate that the imposed nutritional deficiency did not lead to immune depression; this observation is supported by the fact that the animals had no clinical signs of illness during the whole experimental period.

The importance of proteins in the processes of synthesis of substances for growth and tissue re-
newal was demonstrated by the lower weight gain and smaller bowels of the experimental animals, similar to the findings in rats with protein desnutrition.  

The calories from carbohydrates, both in rats and in humans, although fundamental for energy, are unique from the point of view of the processes of synthesis of plastic substances.

The weight gain verified during the experimental period was due primarily to fat deposition. This would be produced from the excess carbohydrate, because albeit these animals had smaller body weights the amount and proportion of retroperitoneal fat relative to the body weight were larger than in the controls. Thus, an implication of this diet is an alteration of the body composition with increased fat relative to lean mass.

When comparing the density of NADPH-diaphorase positive neurons between the groups, statistically significant differences were not found, yet, as the colon area of the experimental animals represented 45.8% of that of the controls, it was expected that the neuronal density were 54.2% greater, because the smaller the intestinal area, the lesser the neuronal spread. However, the density of these neurons was only 3.7% greater than that of the control animals. Therefore, about 50.5% of the nitricergic neurons and 27.2% of the NADH-diaphorase positive neurons of the ascending colon were not evidenced. Significant reductions in the number of myenteric neurons of malnourished rats were also verified in other investigations, and have been attributed to the nutritional deficiency to which the animals were subjected.

However, we cannot state that all the nitricergic neurons which were not seen have been lost, once their evidencing by the NADPH-diaphorase technique is made possible due to the formation of formazan granules from nitro blue tetrazolium, which functions as an artificial electron acceptor. In the specific case of the nitricergic neurons the production of nitric oxide from L-arginine depends on the energy supplied by the oxidation of NADPH, as catalyzed by the enzyme NADPH-diaphorase.

As the vitamins of the complex B participate of the pathways of carbohydrate utilization and niacin of the synthesis and composition of the adenine nucleotides, and as its ingestion was about 20% smaller than recommended and 75% smaller than the amount ingested by the control animals, the hypothesis can be put forward that in the neurons of the myenteric plexus a deficiency of these elements is taking place, with resultant reduction in the energy available to drive the formation of nitric oxide from L-arginine.

Despite niacin being possibly the major substrate of the electron-carrier substances (NAD+/NADH; NADP+/NADPH), we believe that a possible deficiency of L-arginine could be added as an explanation for the lesser evidencing of the NADPH-diaphorase positive neurons, because its decreased availability for the synthesis of nitric oxide would impair the evidencing of the NADPH-diaphorase positive neurons. On the other hand, lipogenesis, which depends on NADPH when taking place through the pentose-ribose phosphate pathway or on NADH and NADPH when through the glycolytic pathway, was maintained, as evidenced by the increased retroperitoneal fat.

In these animals we observed reduced serum albumin, which can be related to the reduced availability of essential amino acids. In pigs with limited protein ingestion it was verified an alteration in the ratio of non-essential/essential amino acids from the fall of plasma albumin onwards. Also in malnourished children it was found a decreased plasma concentration of essential and some non-essential amino acids, among which L-arginine. It should be stressed that arginine is considered an essential amino acid in growing and young rats, but not in adult animals. In humans with protein desnutrition arginine is reduced because it participates of protein synthesis and is used in the urea cycle as well.

Another evidence of the importance of the reduced protein intake for the decreased evidencing of the nitricergic neurons is the fact that the NADPH-diaphorase positive neurons suffered a less marked reduction, although their evidencing also depends upon an adenine nucleotide as energy substrate. It should also be considered that even the components of the vitamin B complex having been reduced due to the addition of starch, they were present in amounts sufficient to prevent clinical manifestations such as cutaneous, muscular, neurologic and hematologic changes, which are common in instances of marked deficit of vitamins from the complex B, although the animals could be in a pre-clinical step. In humans, for each case of extreme desnutrition there are 15-20 cases of pre-clinical alterations. In this period the energy stores decrease and biochemical changes begin to appear, and the functional and anatomical changes arise in the clinical period.

In summary, this experiment demonstrated that the decrease of the protein level of the diet to 8%...
coupled to a decrease in the level of vitamins from the complex B in adult rats does not lead to qualitative or quantitative changes of the blood cells both of the red and the white series, does not reduce the levels of globulin nor lead to edema or the appearance of clinical signs characteristic of protein and vitamin B deficits. On the other hand it causes: less weight gain; change in the body composition with increased fat deposition; decrease in the values of total protein and albumin; decrease in the colonic area, and evidencing of nitrergic and NADH-diaphorase positive neurons at densities inferior to those expected.

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