Persistent Sydenham’s chorea is not associated with sustained lymphocyte dysfunction

A coreia de Sydenham persistente não está associada à disfunção sustentada de linfócitos

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Sydenham’s chorea (SC) is the most common cause of chorea in developing countries1. Apart from chorea, SC patients can exhibit other motor symptoms such as dysarthria and decreased muscle tone. Psychiatric syndromes can also be present, including attention deficit hyperactivity disorder, and obsessive-compulsive disorder2,3.

SC is the late neurological manifestation of Group A beta-hemolytic streptococcal oropharynx infection and one of the major criteria for the diagnosis of rheumatic fever4. Despite the well-known relationship between streptococcal infection and rheumatic fever, the precise pathogenesis of SC remains a matter of debate. One of the most attractive hypotheses proposes that SC is an autoimmune disorder resulting from cross-reactive antibodies against basal ganglia neurons. Autoimmune mechanisms underlying SC pathophysiology are strongly supported by a range of evidence. Studies have reported increased levels of cytokines in serum and cerebrospinal fluid of SC patients in comparison with controls5,6, as well as the presence of circulating anti-basal ganglia antibodies (ABGA) in SC patients5,7.

Chorea and other motor symptoms of acute SC improve after immune-modulatory therapy8.

In vitro studies showed that SC autoantibodies may influence neuronal cell signaling, impairing CNS functioning9,10. Furthermore, rats receiving

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Criteria for rheumatic fever 14 after the careful exclusion of age ± SD, 24.0 ± 5.8). age and gender-matched healthy individuals (M/F, 8/4; mean RSC patients (M/F, 5/4; mean age ± SD, 25.4 ± 11.5) and 12 study: 5 PSC patients (M/F, 4/1; mean age ± SD, 24.7 ± 8.6); 9 Subjects metHoD comparison with control subjects.

SC is traditionally regarded as a self-limited disorder with spontaneous remission after a course of 6 to 9 months (remitted SC; RSC). Nevertheless, a significant number of SC patients remains with chorea on long-term follow-up (i.e. over 2 years), what is called persistent SC (PSC). Proposed mechanisms involved in the persistence of chorea include irreversible basal ganglia damage during the acute phase of the disease or sustained pathological immune response12.

B1 cells are a subset of B lymphocytes that produce antibodies that are frequently auto-reactive. The frequency of circulating B1 cells is very low in healthy subjects but tends to be elevated in patients with autoimmune diseases13. Considering the auto-reactive antibody-mediated hypothesis of SC pathogenesis, PSC may be associated with increased frequency of circulating B1 cells. Accordingly, the current study aimed at evaluating circulating lymphocyte subsets and their activation and functional states in RSC and PSC in comparison with control subjects.

METHOD

Subjects

Fourteen subjects over 16 years-old were recruited for this study: 5 PSC patients (M/F, 4/1; mean age ± SD, 24.7 ± 8.6); 9 RSC patients (M/F, 5/4; mean age ± SD, 25.4 ± 11.5) and 12 age and gender-matched healthy individuals (M/F, 8/4; mean age ± SD, 24.0 ± 5.8).

SC was diagnosed in patients fulfilling the modified Jones Criteria for rheumatic fever14 after the careful exclusion of alternative causes of chorea7. At diagnosis, all patients had ecocardiographic signs of carditis represented by mild mitral insufficiency. The definition for persistent SC was chorea lasting more than 2 years regardless of the use of antichoreic drugs22. Control subjects had no current clinical disease and/or previous history of rheumatic fever.

Flow cytometry analyses

Blood was aseptically collected in heparinized tubes. Peripheral blood mononuclear cells (PBMC) were obtained using a Ficoll-Hypaque (Sigma-Aldrich, St. Louis, MO, USA) gradient. Cells (2 x 10⁸) were stimulated with anti-CD3 monoclonal antibodies (1 mg/mL) (BD Biosciences, San Jose, CA, USA) and anti-CD28 monoclonal antibodies (0.5 mg/mL) (BD Biosciences) in RPMI 1640 (Sigma-Aldrich) supplemented with 5% heat-inactivated human serum (Sigma-Aldrich), 1 mM of L-glutamine and antibiotics 200U of penicillin (Sigma-Aldrich). Cultures were harvested following 18h of stimulation.

Cells were then stained with fluorescein isothiocyanate (FITC) and phycoerytrin (PE)-labeled antibody solutions for 20 min at 4°C. Then, PBMC were washed with 0.1% sodium azide PBS (Sigma-Aldrich), and fixed with 2% formaldehyde in PBS. The antibodies used for staining were specific monoclonal antibodies directed to CD4, CD8, CD5, CD19, CD25, CD69 and CD45RO surface antigens. All antibodies used were from BD Biosciences.

FITC and PE-labeled immunoglobulin isotype control antibodies were included in all experiments. The stained cells were acquired using a FACScan flow cytometer with an air-cooled argon laser (BD Biosciences). Analyses were performed using CellQuest (BD Biosciences) and FlowJo (Tree-Star Inc., Ashland, OR, USA) software, in order to perform the representative dot plots. Leukocytes were analyzed for the frequencies of surface markers expression. The frequency of positive cells was analyzed inside the lymphocyte gate. Limits for the quadrant markers were always set based on negative populations and isotype controls.

Results are shown as means ± standard deviations (SD). Differences among groups were tested using Kruskal-Wallis test with Dunn’s Multiple Comparison post-test. Statistical analyses were performed using GraphPad Prism (GraphPad Software, La Jolla, CA) with a significance level of α set at 0.05.

RESULTS

Initially we evaluated the frequency of B and T lymphocytes. We found no differences between RSC patients, PSC patients and controls regarding the percentages of B cells (CD19+) (Figure 1A: mean ± SD, RSC = 9.10 ± 4.33%; PSC = 10.57 ± 3.41% and controls = 8.51 ± 3.04%) and CD8+ T lymphocytes (Figure 1B: RSC = 0.54 ± 0.23%, PSC = 0.57 ± 0.25% and controls = 0.54 ± 0.21%). The percentage of CD4+ T lymphocytes (Figure 1C: RSC = 46.59 ± 10.84%; PSC = 50.73 ± 4.71% and controls = 41.91 ± 10.39%) and CD8+ T lymphocytes (Figure 1D: RSC = 16.82 ± 5.66%, PSC = 17.45 ± 5.12% and controls = 19.81 ± 8.63%) were also similar among the three evaluated groups.

To investigate lymphocyte activation and functional profiles, we analyzed the expression of the surface markers of recent and chronic activation (CD69 and CD25, respectively) in CD4+ and CD8+ T lymphocytes (Figure 2). Here again, we found no differences among the three evaluated groups of subjects (PSC, RSC and controls). The frequencies of CD4+CD25+ T cells (Figure 2A) were RSC = 3.30 ± 1.18%, PSC = 3.65 ± 1.06% and controls = 2.95 ± 1.16%. The percentages of CD8+CD25+ T cells are given in Figure 2B (RSC = 0.68 ± 0.37%; PSC = 0.40 ± 0.30% and controls = 0.76 ± 0.65%). Figure 2C shows the percentages of CD4+CD69+ T cells (RSC = 0.80 ± 0.38%; PSC = 0.90 ± 0.50% and controls = 1.07 ± 0.90%). Lastly, the percentages of CD8+CD69+ T cells are shown in
Figure 1D (RSC = 1.76 ± 1.42%; PSC = 0.80 ± 0.30% and controls = 1.91 ± 1.32%).

We also evaluated a surface marker of memory (CD45RO) in CD4+ and CD8+ T lymphocytes. No differences were found among groups. The percentages of CD4+CD45RO+ and CD8+CD45RO+ memory T cells are given in Figure 1E (RSC = 22.34 ± 10.16%; PSC = 24.32 ± 8.85% and controls = 21.02 ± 10.19%) and Figure 1F (RSC = 5.76 ± 4.53%; PSC = 3.85 ± 2.61% and controls = 4.79 ± 3.33%), respectively.

Additionally, we found no differences when analyzing PBMC cultured under the presence or not of polyclonal (anti-CD3/CD28) stimuli (data not shown).

**DISCUSSION**

We designed this study primarily to investigate a possible role of lymphocytes in the development and/or persistence of chorea in SC. We found no differences among groups in all of the evaluated immunological parameters. To the best of our knowledge, this is the first study assessing lymphocyte subsets in SC.

More specifically, we were interested in assessing CD4+ and CD8+ T lymphocytes as well as B1 cells in SC patients in comparison with asymptomatic subjects. While CD4+ T cells are involved in the orchestration of immune response, CD8+ T cells can promote target cells destruction\(^\text{15}\). Changes in lymphocyte profile may be associated with a series of autoimmune disorders. In the case of SC, this is particularly relevant for B1 cells that are involved in antibody-mediated autoimmune conditions\(^\text{13}\). Our results failed to demonstrate any significant difference between patients and controls in the composition of lymphocyte subsets, indicating that there is no evidence of sustained lymphocyte activation in PSC. Our work group has previously described a decrease in the percentage of CD14+ monocytes in the peripheral blood of SC patients. In addition, we observed reduced frequency of CD14+CD86+ and CD14+HLA-DR+ cells in SC, indicating that monocytes might play a role in the immune mechanisms underlying SC pathogenesis\(^\text{16}\).

Non-immune mechanisms may be associated with the pathogenesis of PSC. Basal ganglia structural damage may occur during the acute phase of SC, leading to persistent involuntary choreic movements in a subgroup of patients. In line with this hypothesis, neuroimaging studies have demonstrated that patients who remitted completely their choreic movements did not present lesions of the basal ganglia.

![Figure 1](image_url)

**Figure 1.** T and B lymphocyte frequencies of SC patients and controls. (A) percentage of B (CD19+) cells expression; (B) percentage of B1 cells expression (CD19+ CD5+); (C) frequency CD4+ T lymphocytes expression; (D) percentage of CD8+ T lymphocytes expression. RSC = remitted Sydenham’s chorea; PSC = persistent Sydenham’s chorea.
after the acute phase of the disease. Conversely, patients with persistent basal ganglia lesions were more prone to develop recurrences of chorea.

Altogether, our data suggest that the persistence of choreic involuntary movements in SC is not associated with persistent lymphocyte dysfunction. Specially, the percentage of B1 cells – a subset of B lymphocytes increased in autoimmune conditions – are not changed in RSC or PSC. Therefore, patients with persistent SC may not be candidates for immunosuppressive or immunomodulatory therapies.

Figure 2. Activation and functional state of lymphocytes of SC patients and controls. (A) percentage of CD4+ CD25+ T cells; (B) percentage of CD8+ CD25+ T cells expression; (C) frequency of CD4+ CD69+ T lymphocytes expression; (D) frequency of CD8+ CD69+ T lymphocytes expression; (E) frequency of CD4+ CD45RO+ memory T cells; (F) CD8+ CD45RO+ memory T cells expression.

RSC = remitted Sydenham's chorea; PSC = persistent Sydenham's chorea.


