USE OF POLYGLYCOLIC ACID TUBE ASSOCIATED WITH FK506 IN REGENERATION OF PERIPHERAL NERVES

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SUMMARY
Extensive losses of neural tissue preclude the repair performed by means of primary anastomosis. In those cases, nerve autograft is considered as the treatment of choice. It drives the axonal growth and links the ends of distal and proximal stumps, reducing tension at the suture line, a factor that could inhibit neural regeneration(11). When using autografting, some factors must be considered: 1) it always produces morbidity of the donor area; 2) extensive neural tissue losses demand large amounts of autologous tissue, sometimes insufficient; 3) the use of synthetic materials reduces surgery time(2,3). Studies on large neural tissue losses and the need of bridges connecting proximal and distal ends have been conducted during the second half of Nineteenth Century(4). Some authors used other materials in replacement of nerve graft, such as vessels(5,6), fascia(7), plastic tubes(8), absorbable tubes, silicone tubes(9), muscle(10) and synthetic tubes(11). The use of absorbable tube made of polyglycolic acid has been extensively investigated in literature(12) and no statistically significant differences were observed between this approach and the nerve graft for peripheral nerves repair(13). Many substances have been assessed for peripheral nerves injuries treatment. Among those, the FK506 (Tacrolimus), an antibiotic isolated in 1984 for the bacteria Streptomyces tsukubaensis(14), used in primary immunosuppressive therapy, is the drug of choice as a replacement for cyclosporine in immunosuppression(15). Its major advantage consists of its high immunosuppressive potential associated to few side effects(16). In 1994, it has been shown that FK506 also increased the levels of neural regeneration “in vivo”(17) and “in vitro”(18). There are no studies in literature describing the use of neurotubes associated to this drug.

INTRODUCTION
Traumas and tumoral resections generally lead to large neural tissue losses, many times precluding primary anastomosis. In those cases, nerve graft is considered as the treatment of choice. It drives the axonal growth and links the ends of distal and proximal stumps, reducing tension at the suture line, a factor that could inhibit neural regeneration(11). Studies on large neural tissue losses and the need of bridges connecting proximal and distal ends have been conducted during the second half of Nineteenth Century(4). Some authors used other materials in replacement of nerve graft, such as vessels(5,6), fascia(7), plastic tubes(8), absorbable tubes, silicone tubes(9), muscle(10) and synthetic tubes(11). The use of absorbable tube made of polyglycolic acid has been extensively investigated in literature(12) and no statistically significant differences were observed between this approach and the nerve graft for peripheral nerves repair(13). Many substances have been assessed for peripheral nerves injuries treatment. Among those, the FK506 (Tacrolimus), an antibiotic isolated in 1984 for the bacteria Streptomyces tsukubaensis(14), used in primary immunosuppressive therapy, is the drug of choice as a replacement for cyclosporine in immunosuppression(15). Its major advantage consists of its high immunosuppressive potential associated to few side effects(16). In 1994, it has been shown that FK506 also increased the levels of neural regeneration “in vivo”(17) and “in vitro”(18). There are no studies in literature describing the use of neurotubes associated to this drug.

MATERIALS AND METHODS
Fifteen 8-week-old Lewis rats, weighing 200 g – 300 g have been used. With a microsurgical technique, 5 mm- extension defects were created on the sciatic nerve of the right foot. The animals were divided into 3 groups of five animals each. In animals from group A (control group), defect repair was performed by suturing the nerve segment removed (autograft), keeping the original direction. In animals from group B, a 10mm segment of a polyglycolic acid tube, with 2.3mm diameter (Neurotube, manufactured by Neurowen L.C.L., Baltimore/U.S.A.) was interposed between sectioned segments. In group C, a segment of polyglycolic acid tube with the same features as used in group B combined to subcutaneous injection of FK506 5mg/kg at the dorsal region of the neck, was interposed. Doses were initiated one hour after injury and then performed in a daily basis until the day of sacrifice(19) (Table 1).

For the surgical procedure, the animals were submitted to anesthesia with sodium pentobarbital (5m/kg) injected intraperitoneally.
Through the dorsal port on right foot, sciatic nerve was dissected and 5mm of nerve segment were removed (Figures 1, 2 and 3). In group A, the removed segment was sutured in its normal position with 4 epineurial separated stitches with mononylon 10.0 (Figure 4). In group B, the polyglycolic acid tube was positioned at 2.5 mm from each end, maintaining a 5-mm neural defect (Figures 5, 6 and 7). In group C, the same technique of polyglycolic acid tube interposition as in group B was used, and, one hour after injury, the FK506 (5 mg/kg) was subcutaneously injected at the dorsal region of the neck; a procedure repeated in a daily basis until the day of sacrifice (Figure 7).

The animals were supplied with water and food ad libitum and sacrificed 6 weeks after surgery for histological analysis and for counting the number of regenerated myelinated axons. The evaluation of the functional recovery was performed using a technique that analyzes the impression of posterior feet during walking ("walking track analysis") pre- and postoperatively, on the third week, and at the moment of sacrifice (6 weeks).

**RESULTS**

During the 6 weeks of the study, all animals remained healthy, with no infection on surgical wound or plantar neurodystrophic ulcers.

**Histological analysis**

Microscopic analysis of Group A (autograft) slides showed that the graft was well bounded by an epineurium formed by fusiform-profile cells. Internally to the epineurium, there was a large amount of myelinated axons, with variable diameters and homogeneously distributed. Tissue reaction around the graft was greater when compared to the other groups. An escape of regenerated fibers exceeding epineurium boundaries was detected in the 5 animals of that group. A subtle amount of axonal bundles was seen (smaller, if compared to groups B and C) and more signs of walleriana, if compared to groups B and C (Figures 8, 9 and 10).

Histological sections of the medial portion of regenerated nerve (osmium and toluidine blue), in 6 weeks postoperatively.

In Groups B (tAPG) and C (tAPG+FK506), histological findings were similar. In the 10 animals, tubes had a fusiform appearance, and contained tissue with a neural stroma pattern inside them, with a large amount of myelinated axons, of different sizes, grouped in minifascicles of variable sizes and heterogeneously distributed, presenting conjunctive tissue between them. Between the tube and the neural tissue, an intense neoangiogenesis was seen all around it, with blood vessels penetrating transversally to the "meshwork" of the polyglycolic acid tube. The presence of a large amount of neoangiogenesis was shown inside regenerated nerve, but this has not occurred in Group A (autograft). Groups B (tAPG) and C (tAPG+FK506) were different because there was less neoangiogenesis between the tube and the regenerated nerve in Group C (tAPG+FK506) compared to Group B (tAPG). There was less reactive fibrosis around the tubes when compared to Group A (autograft). No escape of regenerated fibers out of the tube was detected. (Figures 11, 12, 13, 14, 15 and 16).

There was no difference in fibers diameter and in the degree of myelinization presented by Groups A (autograft), B (tAPG) and C (tAPG+FK506).

**COUNT OF THE NUMBER OF REGENERATED MYELINATED AXONS**

The average number of regenerated myelinated axons, standard deviation and error for each group are represented on Graph 1 (average and standard error).

In Group A, an average of 7,225.6±617.5 regenerated myelinated axons were counted. In Groups B and C an average of 4,225.2±376.8 and 6,459.8±630.9 regenerated myelinated axons were observed, respectively. Variance analysis followed by multiple comparisons by the Tukey method (p < 0.05), was used for evaluating data. No statistically significant differences were seen between groups A and C regarding the number of regenerated axons after 6 weeks of nerve sectioning. Group B presented, in average, a higher and statistically significant number of regenerated axons.
myelinated axons when compared to groups A and C. Those data are summarized on Table 2.

**FUNCTIONAL STUDY**

Figure 17 shows the appearance of rats’ footprints pre- and postoperatively for the four groups. SFIs observed preoperatively were: in Group A, an average of 8.002±5.26; in Group B, 928±13.144; in Group C, 6,489±7.011. The statistical analysis by Kruskal-Wallis’ method, with a significance level (p<0.05), did not show statistically significant difference among the four groups for the sciatic function index (SFI) preoperatively.

Mean SFI (sciatic function index) postoperative values (immediate, 3 and 6 weeks), calculated for each group are presented on Table 3 (average and standard error).

Immediate postoperative SFIs were, respectively, in average: Group A (autograft) - 83.149±4.236; Group B (tAPG) - 73.378±1.109; Group C (tAPG+FK506) -81.050±6.002 and Group D (tAPG+graft) - 77.891±1.379.

SFIs of 3 weeks postoperatively were in average, respectively: Group A (autograft) - 71.017±6.240; Group B (tAPG) - 53.139±4.681; Group C (tAPG+FK506) -72.598±7.665 and Group D (tAPG+graft) - 69.486±0.810.

SFIs of 6 weeks postoperatively were in average, respectively: Group A (autograft) -51.052±4.994; Group B (tAPG) -44.658±2.870; Group C (tAPG+FK506) -53.811±3.648 and Group D (tAPG+graft) -50.298±1.585.

Using the same data presented by the groups regarding postoperative SFI and submitting them to statistical analysis by the variance method with repeated measure (time) and factor (treatment), followed by multiple comparisons by Tukey’s method and significance level of (p<0.05), a SFI variation was seen regarding postoperative time, as we will describe below:

In the immediate postoperative time, there was no statistically significant difference among the four assessed groups.

After 3 weeks postoperatively, there was a statistically significant difference between Group B (tAPG) and the other groups (Group A (autograft) and Group C (tAPG+FK506). There was no statistically significant difference between groups A and C. Those data are summarized on Table 4.

In the postoperative period of 6 weeks, there was no statistically significant difference among the four assessed groups.

**DISCUSSION**

In cases of peripheral nerves injury with tissue loss, where the extension of the defect precludes primary suture, the best method for repair seems to be the autograft[2,20]. Some factors regarding this topic are relevant, as follows: 1. the need to remove autologous tissue always produces morbidity to donor area; 2. large injuries require the removal of an extensive amount of autogenous tissue, sometimes unavailable; 3. the use of artificial materials does not require the time spent for removing the autograft[21]; 4. results achieved with the autograft are not fully satisfactory[22].

The interposition of tubular conductors as a bridge between stumps of an injured nerve is showing encouraging experimental and clinical outcomes. For fixing small defects, where the nerve distance is not sufficient to allow chemotactic and chemotrophic attraction exerted by the distal stump on the axonal growth region, the results achieved are comparable to those of the autograft[2,9,12,13,23]. Recent studies have been demonstrating the polyglycolic acid tube as an alternative for nerve graft[12,24,25]. Synthetic tubes made of absorbable material showed better results when compared to non-absorbable ones[26,27]. Absorbable materials cause little fibrotic reaction and the absorption of them do not preclude nerve regeneration and clinical outcomes. For fixing small defects, where the nerve distance is not sufficient to allow chemotactic and chemotrophic attraction exerted by the distal stump on the axonal growth region, the results achieved are comparable to those of the autograft[2,9,12,13,23].

The use of substances associated to tubing chambers shows an important role in the development of better functional results after peripheral nerve injury. The FK506 is a new immunosuppressive agent[15,28] 10 to 100 times more powerful than cyclosporine[29]. It presents fewer side effects[16] and is being used successfully in heart, kidney and liver transplants[15,30,31]. Some explanations have been suggested by biochemists for understanding neurons regeneration and growth process with the FK506. This antibiotic causes an immunosuppressive effect, inhibiting calcineurine activity (it plays an important role in the T-cells proliferation regulation). In addition, the FK506 bonds to the immunophilin FKBP-12 enhancing “Gap-43” (asso-
ciated growth protein 43) phosphorylation, activating it. "Gap-43" plays a relevant role in the axonal elongation process(18). Therefore, the FK506 could hasten axonal regeneration through a direct effect over axonal buds growth.

In group A, the use of an autograft is considered as the treatment of choice for the repair of neural defects with tissue loss(2,25). In group B, the polyglycolic acid tube has already been used as an alternative for nerve graft in primates when tissue loss is less than 30 mm(12,24,25,33). In group C, the tube was associated to a drug enhancing neural growth(17,34,35). The development of neuroma was seen only in rats submitted to autografts(36). No tube collapse was observed in groups B and C, although this complication has been described(2,12).

In group A, fibers out of the epineurium boundaries and neuroma formation along suture lines were observed. Those changes have previously been mentioned by other authors(9,36). No fibers out of the boundaries of the epineurium were seen in the groups using tubes.

Regarding histological aspects, groups B and C presented with similar patterns. It is known that FK506 increases axonal regeneration rates(17,19.35,37). When administered during different periods, it proportionally increases the number of myelinated fibers(19,37,38), more extensive axons(19,37), longer fibers(37) and a more advanced maturation period(17,37,38). Hypotheses intending to explain those properties are focused on a reduced level of regenerative buds destruction(37) and on an increase of the myelination process, leading to the formation of thicker fibers and an increased speed towards target organs(37,38).

Despite FK506’s properties related to neural regeneration described in literature, a great difference between groups B and C was not seen. The variation of the fiber diameter, distribution in variable-sized minifascicles heterogeneously located, neangiogenesis, degree of Walleriana degeneration, absence of escape, cicatricial reaction, and the amount of axons bundles behaved similarly regarding the presence of myelinated axons. The difference between groups B and C was a lower degree of neangiogenesis between the tube and the regenerated nerve in group C.

The average number of regenerated myelinated axons in group B was the lowest. The combination of FK506 with the polyglycolic acid tube increased the average number of regenerated myelinated axons, but did not show a statistically significant difference when compared to control group. This potential to increase the average number of regenerated axons corroborates literature findings(37,38).

Regarding functional evaluation, there were no differences between the immediate and the late postoperative periods. However, in the 3-week period, the SFI average for group B was the best, if compared to other groups, and there was no difference between groups A and C. The difference between functional findings and axons counts may be explained by the importance of re-established connections accuracy of the regenerated axons with target organs and not to the total number of fibers verified(40,41).

Expectations are indeed very good for further researches with the neurotubes, considering more adequate and cheaper materials to be used in la-

### Table 4 - Statistical analysis of 3-week postoperative SFI by the variance method with repeated measurement (time) and one factor (treatment) followed by multiple comparisons by Tukey’s method and significance level of (p<0.05).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>SE - IMM</th>
<th>SE - 3w</th>
<th>SE - 6w</th>
</tr>
</thead>
<tbody>
<tr>
<td>Autograft</td>
<td>4.236</td>
<td>6.240</td>
<td>4.994</td>
</tr>
<tr>
<td>TAPG</td>
<td>1.109</td>
<td>4.881</td>
<td>2.870</td>
</tr>
<tr>
<td>TAPG+FKN06</td>
<td>6.022</td>
<td>7.665</td>
<td>3.648</td>
</tr>
</tbody>
</table>

# denotes a statistically significant difference, showing a better SFI in group B.

### Table 3 - Postoperative SFI (immediate, 3 and 6 weeks) according to the kind of treatment employed.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Immediate</th>
<th>3 weeks</th>
<th>6 weeks</th>
<th>SE - IMM</th>
<th>SE - 3w</th>
<th>SE - 6w</th>
</tr>
</thead>
<tbody>
<tr>
<td>Autograft</td>
<td>81.419</td>
<td>71.015</td>
<td>51.052</td>
<td>4.236</td>
<td>6.240</td>
<td>4.994</td>
</tr>
<tr>
<td>TAPG</td>
<td>-73.378</td>
<td>-53.139</td>
<td>-44.658</td>
<td>1.109</td>
<td>4.881</td>
<td>2.870</td>
</tr>
<tr>
<td>TAPG+FKN06</td>
<td>-81.050</td>
<td>-72.598</td>
<td>-53.811</td>
<td>6.022</td>
<td>7.665</td>
<td>3.648</td>
</tr>
</tbody>
</table>

# denotes a statistically significant difference, showing a better SFI in group B.
REFERENCES


