Biodegradation of Diesel Oil by Yeasts Isolated from the Vicinity of Suape Port in the State of Pernambuco -Brazil

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ABSTRACT

The aim of this work was to investigate the potential of the diesel oil degrading yeasts to use them in bioremediation of areas contaminated by this pollutant. The cultures, identified as Rhodotorula aurantiaca UFPEDA 845 and Candida ernobii UFPEDA 862, were selected at the initial stage. In the course of the biodegradation assays, C. ernobii degraded tetradecane, 5 methyl-octane and octadecane completely and decane (60.8%) and nonane (21.4%) partially whilst R. aurantiaca presented degradation percentages of 93% for decane, 38.4% for nonane and 22.9% for dodecane.

Key words: Biodegradation, yeasts, diesel oil

INTRODUCTION

Strategies for controlling environmental contamination by petroleum and its derivatives have been the subject of various studies over the past three decades. When a spillage occurs the first action is to remove the oily phase by mechanical or by physical-chemical means through the application of surfactants to disperse the layer of oil. Bioremediation is an alternative that has been used to eliminate or minimise the effects of pollutants by using microorganisms which have biodegradation potential (Atlas, 1995a). Diesel oil is a medium distillate of petroleum containing: n-alkanes, branched alkanes, olefins and small concentrations of aromatic polycyclic compounds (Baker and Herson, 1999). The Suape port terminal in the State of Pernambuco in Brazil is used by important distributors of diesel oil. The study of the biodegradation of fuels by native microorganisms is of extreme ecological importance, as the nearby beaches are subject to contamination by pollutants coming from the port complex. The objective of this work was to investigate the potential of the diesel oil degrading yeasts to use them in bioremediation of contaminated areas.

MATERIALS AND METHODS

Selection Test
Twenty-three strains of yeast, isolated from samples polluted by petroleum derivatives, collected from the Barra Lagoon, near the Suape port terminal, were evaluated for biodegradation of diesel oil. Hanson et al (1993) method was used for selection of the strains. This method consisted of incorporating into the medium an electron
acceptor such as 2,6-dichlorophenol-indophenol (DCPIP) to test the ability of the microorganism to utilize the hydrocarbon substrate by observing the colour change of DCPIP from blue (oxidized) to colourless (reduced). Each well in a microtitre plate was filled with 250µL Büshnell-Haas (BH) medium, 10µL of diesel oil and 25µL of each microbial suspension standardised at $10^8$CFU/mL.

### Identification of the Isolates
The selected yeasts were identified by their macroscopic, microscopic and physiological characteristics, as described by Lodder, (1970); Kreger-Van Rij, (1984) and Barnett et al., (1990).

### Acclimatation
The selected strains were initially submitted to acclimatation by varying the concentration of diesel oil from 2 to 12% for a period of 20 days, before being used in the biodegradability tests.

### Biodegradation Assay
The biodegradation assay was carried out over 15 days in six Erlenmeyer flasks of 500 mL (corresponding to 0, 3, 6, 9, 12 and 15 days of experiment, respectively) each flask containing 12mL of diesel oil, 68mL of the Bushnell Haas medium containing KH$_2$PO$_4$ (1g), K$_2$HPO$_4$ (1g), NH$_4$NO$_3$ (1g), MgSO$_4$ 7H$_2$O (0.2g), FeCl$_3$ (0.05g), CaCl$_2$ 2H$_2$O (0.02g), H$_2$O destilated 1L (Atlas, 1995b) and 20mL of acclimated inoculum under nonaseptic conditions. One sample from each assay was collected at three days intervals. No experiment replication was performed in the course of the assays. The pH and the biomass were measured while the evaluation of the biodegradation of the hydrocarbons of diesel oil was carried out only at the end of 15 days using gas chromatography connected to mass spectrometry.

### Biomass Measurement
The biomass was measured as a dry weight. Aliquots of 20mL were filtered through a microbiological membrane (0.45µm) and dried overnight (12h) at 80°C.

### Chemical Analysis
Chromatographic and spectrometric analyses were performed by GC-MS (Shimadzu™, Model 17A/QP5050A) using a DB-5 chromatographic column (5% diphenyl and 95% dimethylpolysiloxane) of size 30m x 0.25mm x 0.25 µm. The temperature was programmed to vary linearly from 40°C to 270°C at the rate of 10°C min$^{-1}$ and maintained for 22 min. Helium was the carrier gas and the interface temperature was 280°C. Injection of samples and control into a GC-MS system was carried out in triplicate. The highest resolution chromatographic peaks were scanned to find their corresponding mass-fragmentation profile. Compounds were characterized based on similarities between their mass spectrum and those presented by Wiley™ Compounds Library. Control peak-areas were used as a point of reference for the remaining compounds (100%) in the untreated system. Sample peak-areas were reported as a percentage of control peak-areas.

## RESULTS AND DISCUSSION

### Selection Test
Of the 23 strains tested, only two (UFPEDA 845 and UFPEDA 862) were selected because they completely discoloured the culture medium after 16 and 24h of incubation at 30°C, respectively. The other strains caused the culture medium to become colourless after periods of 48 h (8%) and 72 h (84%). Hanson et al. (1993), using a microtitre plate to test the potential of five isolated strains of bacteria to degrade crude oil, selected two isolates because they caused a change in the colour of the culture medium containing the DCPIP indicator after 12h of incubation, while the other isolates responded less quickly to the biological oxidation, showing a change in the DCPIP indicator only after 24h of incubation at 35°C. Another criterion used to determine the most probable number of microorganisms with the potential to degrade crude oil was suggested by Brown and Braddock (1990) who verify the rupture of the oily surface of the culture medium in microtitre plates. The rupture was attributed to the production of biosurfactants at the beginning of the biodegradation process.

### Identification of Yeasts
The UFPEDA 845 strain was identified as *Rhodotorula aurantiaca* and the UFPEDA 862 strain as *Candida ernobii*. Komagata et al. (1964), investigating 500 yeasts, found that 56 species assimilated hydrocarbons, most of them belonging to the *Candida* genus. Ahearn et al. (1971) cited
other genera that have the capacity to assimilate hydrocarbons, such as: *Rhodospiridium*, *Rhodotorula*, *Saccharomyces*, *Sporobolomyces*, *Trichosporum* and *Cladosporium*. Bento and Gaylarde (2001) suggested that the yeasts *Rhodotorula glutinis* and *Candida silvicola* were the main contaminants in diesel oil storage tanks.

**Biodegradation Assays**

Before proceeding to the biodegradation assays, the two selected strains were first acclimated using increasing concentrations (from 2% to 12%) of diesel oil as the only carbon and energy source for 20 days. During the acclimatation period, both selected yeast strains presented different growth, resulting in different initial biomass concentration in the biodegradation assays (0.3g/L for *Rhodotorula aurantiaca* UFPEDA 845 and 0.05g/L for *Candida ernobii* UFPEDA 862). According to Alexander (1994), on acclimatation, the level of assimilation of the pollutant may be higher or lower, but, if further pollutant is added, degradation of this increment will occur after a short or practically non-existent period of acclimatation. Several authors have pointed out that the adaptation of the microbial culture to the pollutant is preponderant in the biodegradation process. Microorganisms isolated from environments with a history of pollution by petroleum hydrocarbons have higher ability to degrade such pollutants (Englert and Kenzie, 1993; Kilbane II et al., 2000).

The biodegradation assays with 12% of diesel oil, carried out with the two selected strains (*Rhodotorula aurantiaca* UFPEDA 845 and *Candida ernobii* UFPEDA 862), were accompanied by measuring the pH and biomass over a period of 15 days, as seen in Figs. 1 and 2.

![Figure 1](image1.png)

**Figure 1** - Biomass and pH of *Rhodotorula aurantiaca* UFPEDA 845 over 15 days of degradation using 12% diesel oil as carbon source.

![Figure 2](image2.png)

**Figure 2** - Biomass and pH of *Candida ernobii* UFPEDA 862 over 15 days of degradation using 12% diesel oil as carbon source.
It should be noted that both strains showed pH values in the range of 6.6 and 7.0, near to neutrality, over the fifteen day assay, with a coincidental decrease on the pH values on the third day, which indicate greater production of organic acids as intermediate products of the biodegradation of the hydrocarbons. Leahy and Colwell (1990) observed that pH values between 6.0 and 8.0 were more appropriate to the action of microorganisms that degraded petroleum, although fungi were more tolerant of acidic conditions. Figs. 1 and 2 showed that there was an increase in the biomass concentration in both cultures of yeast. Fig. 1 showed that the biomass of *R. aurantiaca* UFPEDA 845 initially grew from 0.3 to 0.7 g/L over the first six days, then stabilized for three days, and then grew significantly (from 0.7 to 1.3 g/L) until the end of the experiment, suggesting a diauxic behaviour, which means two clear phases of growth separated by a period of adaptation to the new nutritional conditions. Given the great diversity of hydrocarbons in the diesel oil, it can be supposed that constituent of the oil responsible for the growth of the *R. aurantiaca* in the first phase was probably not that used in the second phase of growth. Fig. 2 showed that the *C. ernobii* UFPEDA 862 exhibited slow growth up to the ninth day, but the biomass subsequently grew exponentially until the twelfth day, maintaining by the end of the 15 days a maximum concentration of 0.33 g/L, which is four times less than that attained by the *R. aurantiaca* UFPEDA 845 in the same time (1.3 g/L). Espírito-Santo (2002), using agitated flasks with the Bushnell-Haas medium containing 5% (v/v) concentration of diesel oil, applied $10^7$ CFU/mL of inocula and observed a maximum of $10^8$ CFU/mL of fungi after 8 days, decreasing shortly thereafter to $10^5$ CFU/mL, at the end of the 20-day experiment. The evaluation of the biodegradability of diesel oil by the selected cultures was carried out only at the end of the assay (15 days), whose results in terms of percentages of degradation of aliphatic hydrocarbons present in the diesel oil are shown in Fig. 3.

Of the investigated aliphatic hydrocarbons, 10 were examined with greater precision by the methodology used in the GC-MS system. The yeast *R. aurantiaca* UFPEDA 845 showed higher percentages of degradation for decane, nonane and dodecane (93.0%, 38.4%, and 22.9%, respectively) while *C. ernobii* UFPEDA 862 completely degraded tetradecane, 5-methyl-octane and octadecane, and partially decane (60.8%) and nonane (21.4%).

![Figure 3](image_url) - Percentage of degradation of diesel oil hydrocarbons after 15 days of biodegradation assays, using the yeasts *Rhodotorula aurantiaca* UFPEDA 845 and *Candida ernobii* UFPEDA 862 with 12% of the oily source.
Richard and Vogel (1999), cultivated a consortium using diesel oil as the only source of carbon and energy and reported degradation of 90% of the fuel oil after 50 days of cultivation. Ruberto et al. (2003), working with a nutritional C:N:P ratio of 100:12:3, after 50 days of the process, obtained reductions in the mean concentration of hydrocarbons present in diesel oil of 35% (indigenous microflora) and 65% (psychrotolerant strain) compared to the initial concentration. Suparna et al. (2004), using diesel oil as the sole source of carbon and energy, tested the diesel oil biodegradation ability of cultures isolated from Arabian sea sediments. Under aerobic conditions, 39% loss of fuel was observed over 8 days where 80% of the loss was due to aliphatic constituents, and under anoxic nitrate reducing conditions the rate and extent of degradation was significantly lower (18%).

The results obtained for the yeasts \textit{R. aurantiaca} UFPEDA 845 and \textit{C. ernobii} UFPEDA 862 can be used to draw up a bioremediation protocol involving the optimisation of parameters, such as biostimulation and bioaugmentation for the recuperation of areas contaminated by diesel oil as well as by other oily sources from petroleum.

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