Vertical Alveolar Crest Bone Maintenance
Around Implants in Two-Stage Surgery:
An In Situ Study in Dogs

Mário Roberto LEONARDO¹
Alberto Tadeu Nascimento BORGES¹
Walter MARTINS-JÚNIOR²
Alexandra Mussolino de QUEIROZ¹
Sada ASSED¹

¹Department of Pediatric Clinics, Preventive and Community Dentistry,
Dental School of Ribeirão Preto, University of São Paulo, Ribeirão Preto, SP, Brazil
²Periodontics, Private Practice, Ribeirão Preto, SP, Brazil

The aim of this study was to evaluate in situ changes in the alveolar crest bone height around immediate implant-supported crowns in comparison to tooth-supported crowns (control) with the cervical margins located at the bone crest level, without occlusal load. In Group I, after extraction of 12 mandibular premolars from 4 adult dogs, implants from Branemark System (MK III TiU RP 4.0 x 11.5 mm) were placed to retain complete acrylic crowns. In Group II, premolars were prepared to receive complete metal crowns. Sixteen weeks after placement of the crowns (38 weeks after tooth extraction), the height of the alveolar bone crest was measured with a digital caliper. Data were analyzed statistically by the Mann-Whitney test at 5% significance level. The in situ analysis showed no statistically significant difference (p=0.880) between the implant-supported and the tooth-supported groups (1.528 ± 0.459 mm and 1.570 ± 0.263 mm, respectively). Based on the findings of the present study, it may be concluded that initial peri-implant bone loss may result from the remodeling process necessary to establish the biological space, similar to which occurs with tooth-supported crowns.

Key Words: immediate implants, alveolar bone crest, bone resorption, in situ evaluation.

INTRODUCTION

Immediate dental implant placement is a recent procedure to oral rehabilitation that has shown advantages, such as a smaller number of surgical procedures, lower cost and reduction of the edentulism period, compared to conventional technique (1). In addition, immediate implants have a better patient acceptance (2,3) for allowing a good preservation of both height and width of the alveolar bone crest (4), resulting in satisfactory esthetics and improved intermaxillary relationship (2,5). Maintenance of the alveolar bone crest dimensions has been observed around implants placed immediately after tooth extraction (5) as well as new bone formation in areas with alveolar infrabony defects (6). Also, cervical bone loss ratio is reduced around immediate implants placed in maxillary and mandibular posterior regions compared to delayed implants, mainly in poor bone quality areas (4).

The level of bone crest surrounding the implant is of utmost significance to determine osseointegrated implant success (7), as preservation of marginal bone height is highly important for long-term dental implant survival (8). Anatomic factors, such as the quality and architecture of bone tissue, as well as implant features, e.g., length, surface area, coating, implant timing (4) and occlusal load (9), may influence alveolar bone crest resorption.

Longevity is the most common parameter to evaluate the success of dental implants, and the maintenance of cervical bone around the implant is the main factor to determine such a positive outcome (10).
Schwartz-Arad et al. (4) have pointed out that the success of dental implants should be evaluated based on both clinical and radiographic parameters, which could indicate peri-implant bone loss.

The influence of the peri-implant alveolar bone crest level on the presence or absence of papillae between implants and adjacent teeth has been demonstrated (11). In this way, prediction of the amount of alveolar bone crest loss is important for a good esthetic outcome after placement of implant-supported crowns and dentures (12). This study evaluated \textit{in situ} changes in the alveolar crest bone height around immediate implant-supported crowns in comparison to tooth-supported crowns (control) with the cervical margins located at the bone crest level, without occlusal load.

\section*{MATERIAL AND METHODS}

\subsection*{Animal Procedures}

All animal procedures were performed according to the protocol reviewed and approved by the Ethics in Animal Research Committee of the University of São Paulo in compliance with the applicable ethical guidelines and regulations of the International Guiding Principles for Biomedical Research Involving Animals. The study was design according to the ISO 7405-1997 standard: Biological evaluation of dental materials (13).

Four 2-year-old adult mongrel dogs weighing 15-20 kg were used and received a standard diet with water \textit{ad libitum}. The animals were anesthetized with 1\% sodium thiopental (Thiopentax; Cristália Produtos Químicos Farmacêuticos Ltda., São Paulo, Brazil; 15 mg/kg body weight, i.v.) and preoperative periapical radiographs of the premolar region were taken in order to localize dental roots and nerve region. A total of 24 mandibular premolars were assigned to 2 groups: Group I was composed by 12 implant-supported crowns (after premolar extraction) and Group II was composed by 12 natural tooth-supported crowns. Figure 1 describes the clinical procedures and timelines of the study. Before the surgical/prosthetic procedures, antisepsis of the oral cavity was performed with 3\% hydrogen peroxide and 0.12\% chlorhexidine gluconate followed by rubber cup/pumice prophylaxis using sterile instruments.

In Group I, 3 mandibular premolars were extracted from each dog according to the following protocol: the first dog had the 2nd and 4th premolars extracted from the right side and the 3rd premolar extracted from the left side; the second dog had the 2nd and 4th premolars extracted from the left side and the 3rd premolar extracted from the right side; in the other animals, the sides were alternated in a randomized manner. After gingival flap elevation, the teeth was decoronated with a 29 mm carbide Zekrya bur (Dentsply/ Maillefer, Ballaigues, Switzerland) under sterile saline irrigation, followed by gentle luxation of the roots with a straight elevator and extraction with a \#151 forceps. For each dog, 3 titanium Branemark System TiUnite MK III implants (4.0 x 11.5 mm; Branemark System; Nobel Biocare AB, Gothenburg, Sweden) were inserted in the alveolar wounds immediately after extraction within the same session, according to the Nobel Biocare protocol.

For preparation of the receptor areas, a spear-shaped guide bur (Nobel Biocare AB) was used to mark the site of implant placement in the intraradicular septum of the extracted teeth followed by a helicoidal bur (2 mm in diameter; 13 mm long) (Nobel Biocare AB). The parallelism of each perforation was evaluated with a positioning device and a pilot bur (3 mm in diameter; 13 mm long) was used to enlarge its diameter from 2 to 3.

When necessary, the coronal portion of the perforation was widened in order to ensure that the implant platform and the alveolar bone crest were in the same level. All surgical burs (Nobel Biocare AB) were used under continuous and abundant saline irrigation and all procedures were performed at 2,000-3,000 rpm with an electronic torque controller (Torque Controller; Nobel Biocare AB, Gothenburg, Sweden). After preparation of the screw threads at 15-20 rpm, the implants were inserted and the cover screws were placed. A relaxing incision was made to allow the reposition of the flap over the implants and sutured with 4.0 silk sutures (Ethicon; Johnson and Johnson, São José dos Campos, SP, Brazil) and control periapical radiographs were taken.

After the surgical procedures, the animals were given a penicillin-based antibiotic (600,000 IU), an antitoxic immunostimulant (Mercepton; Bravet, Rio de Janeiro, RJ, Brazil) and an analgesic (Tramadol chlorhydrate; Anangor; Laboratórios Biosintética Ltda., São Paulo, SP, Brazil) at a dose of 3 mg/kg body weight, i.v., once a day for 7 days. Within this period, two biofilm control session with topical applications of 0.12\% chlorhexidine gluconate were performed. Soft food was given to avoid additional trauma to the surgical sites. Sutures were removed after 7 days under sedation.
with 0.2% Acepran (Acepromazine, Univer S/A, São Paulo, SP, Brazil).

Sixteen weeks after implant placement, the sites were surgically re-opened for placement of the healing abutments. At the 20th week after implant placement, which is the time necessary for peri-implant mucosa healing, impressions were made with addition-curing silicone (Monopren Transfer, Zhermack) and polysiloxane, condensation-type silicone elastomer (Speedex; Coltène, Altstatten, Switzerland) for fabrication of complete acrylic resin (Cesead 2; Kuraray Co. Ltd., Kurashiki, Japan) crowns, which were screwed onto the implants (n=12) with the gingival margins located at the level of the alveolar bone crest (Group I). The mandibular premolars (n=12) adjacent to the implants were prepared to receive complete metal crowns with beveled gingival margins located at the level of the alveolar bone crest. The crowns were made in gold alloy (Goldgato, CNG, Santo Amaro, SP, Brazil) and were cemented to the teeth with zinc phosphate cement (SS White, Rio de Janeiro, RJ, Brazil) (Group II). In both groups, the crowns were placed in infra-occlusion.

**In Situ Determination of Alveolar Bone Crest Height**

For *in situ* assessment of the alveolar bone crest height, the mandibles were dissected, reduced in volume, washed thoroughly in running water, and the gingival tissue was dissected with a scalpel blade. The height of the alveolar bone crest was measured directly on the specimens by the same calibrated examiners using a digital caliper (Mitutoyo Corporation, Tokyo, Japan).

In Group I (implant-supported crowns), this measurement corresponded to the distance between the implant platform and the top of the alveolar bone crest, while in Group II (tooth-supported crowns) this measurement corresponded to the distance between a reference point at the cervical margin of the crown and the top of the alveolar bone crest (Fig. 2). Two sites mesially (mesiobuccal and mesiolingual) and two sites distally (distobuccal and distolingual) to the teeth/implants were examined. The means of the linear measurements obtained by the 3 examiners in the mesial and distal regions of the teeth/implants were calculated.

![Figure 1. Summary of the clinical procedures and timelines of the study.](image-url)
Statistical Analysis

The in situ data (in mm) of the alveolar bone crest height 16 weeks after cementation of the implant-supported and tooth-supported crowns (38 weeks after the beginning of the study) were compared and analyzed statistically by Mann-Whitney U nonparametric test, for independent samples at 5% significance level. Data were expressed as mean ± standard deviation (SD).

RESULTS

Four specimens were lost during the study: 2 specimens from Group I due to lack of implant osseointegration and 2 specimens from Group II due to crown displacement. The results of the in situ assessment of the change in the alveolar bone crest height in both groups are presented in Table 1. The data correspond to the means of the 3 examiners in both examined regions (mesial and distal). There was no statistically significant difference (p=0.880) between Group I (1.528 ± 0.459 mm) and Group II (1.570 ± 0.263 mm).

DISCUSSION

Peri-implant bone loss is an important factor to determine the long-term success of dental implants (7). For this reason, this study investigated whether the placement of crowns retained by either immediate implants or natural teeth with margins located at the bone level and without incidence of occlusal load would induce changes in the alveolar bone crest height. The outcomes showed that the ability of immediate implants to maintain the alveolar bone crest height is similar to that of natural teeth prepared to support crowns.

Table 1. In situ assessment of the height (in mm) of the alveolar bone crest as measured 16 weeks after placement of the crowns (38 weeks after the beginning of the study). Data correspond to the means of the 3 examiners in the mesial and distal regions.

<table>
<thead>
<tr>
<th>Specimen</th>
<th>Group I (Implant-supported crowns)</th>
<th>Group II (Tooth-supported crowns)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1.085</td>
<td>1.452</td>
</tr>
<tr>
<td>2</td>
<td>1.040</td>
<td>1.645</td>
</tr>
<tr>
<td>3</td>
<td>1.030</td>
<td>1.510</td>
</tr>
<tr>
<td>4</td>
<td>1.010</td>
<td>1.355</td>
</tr>
<tr>
<td>5</td>
<td>1.605</td>
<td>1.500</td>
</tr>
<tr>
<td>6</td>
<td>1.545</td>
<td>1.595</td>
</tr>
<tr>
<td>7</td>
<td>1.810</td>
<td>1.190</td>
</tr>
<tr>
<td>8</td>
<td>2.135</td>
<td>1.480</td>
</tr>
<tr>
<td>9</td>
<td>1.890</td>
<td>2.140</td>
</tr>
<tr>
<td>10</td>
<td>2.130</td>
<td>1.835</td>
</tr>
<tr>
<td>Means ± SD</td>
<td>1.528 ± 0.459*</td>
<td>1.570 ± 0.263*</td>
</tr>
</tbody>
</table>

*Mann-Whitney U-test for independent samples (α=5%). There was no statistically significant difference (p=0.880) between the groups.
Alveolar bone crest loss in two-stage implant systems has been attributed to occlusal load, presence of microorganisms and implant microgap location (14). In the present study, the crowns were placed in infra-occlusion in order to minimize the influence of occlusal load. In addition, as peri-implant bone loss can often be associated with peri-implantitis (15), chemical control of biofilm after implant placement was carried out with topical applications of 0.12% chlorhexidine gluconate, which is considered as the gold standard of antimicrobial agents (16). According to Ricci et al. (17), rigorous control of dental biofilm can limit alveolar bone crest resorption in 2-stage implants.

Shibli et al. (18) reported that the presence of a microgap between the implant platform and abutment causes a biological condition in which the peri-implant bone tissue is remodeled as if it ‘seeking to delimit’ a space to accommodate the junctional epithelium and the connective tissue that are responsible for peri-implant marginal homeostasis. This space is similar to that of prosthetically restored natural teeth and is known as biological space. A physiological dimension seems to exist between the bone and the crown-implant interface that is established early and maintained over time (12).

Considering that in the present study alveolar crest bone loss, measured in situ, was very similar between implant-supported and tooth-supported crowns, it is possible to suggest that the location of the microgap at the alveolar bone crest level together with the presence of microorganisms caused bone resorption in an attempt to re-establish the biological space. Thus, implant placement had not a negative effect on alveolar bone crest. Similar results have been reported also using mandible dog as experimental model (19).

In contrast with some studies (4,20) that reported that implant placement induces alveolar bone crest loss, the results of the present study suggest that changes in the alveolar bone crest height is not caused by the implant itself, but it is rather induced by the location of the crown cervical margin at the bone crest level. It may be concluded that the peri-implant alveolar bone crest loss is due to need of bone remodeling for establishment of the biological space, similar to which occurs with natural tooth-supported crowns.

REFERENCES

10. Malmqvist JP, Senerby L. Clinical report on the success of
47 consecutively placed Core-Vent implants followed from 3 months to 4 years. Int J Oral Maxillofac Implants 1990;5:53-60.


Accepted April 17, 2008