Tumor-Infiltrating Macrophage and Microvessel Density in Oral Squamous Cell Carcinoma

Deise Souza Vilas Bôas, Christina Maeda Takiya, Clarissa Araújo Silva Gurgel, Márcia Grillo Cabral, Jean Nunes dos Santos

Tumor-associated macrophages (TAM) are the main cellular component in stroma of many tumors and participate in tumor angiogenesis. The aim of present study was to compare the microvascular density (MVD) and infiltrating macrophage density (IMD) in oral squamous cell carcinomas (OSCCs) with different histological grades. A histomorphometric analysis was performed after immunohistochemistry using antibodies such as von-Willebrand factor and CD68. A significant difference in MVD was found between well and moderately differentiated OSCCs (p=0.05). TAM were largely present in all studied tumors and the IMD was not different among OSCCs with different histological grades (p=0.381). Significant correlation between MVD and IMD was not observed (p=0.870). In conclusion, these results suggest that TAM and angiogenesis have an influence at different histological grades of OSCC. However, the lack of correlation between MVD and IMD could suggest that angiogenesis does not depend on the number of macrophages present in OSCC, but their predominant phenotype. Further studies involving distinct phenotypes of macrophages should be done to better understand the influence of TAM on the tumor angiogenesis.

Introduction

The growth of oral squamous cell carcinoma (OSCC) occurs mainly as a result of the proliferation of malignant cells and the formation of supportive tissues such as new blood vessels. Due to this occurrence, a larger angiogenesis in OSCC is commonly associated with a more aggressive behavior of the tumor and a poor prognosis (1). In addition to blood vessel, the tumor microenvironment contains cells and chemical mediators that have also been the focus of studies due to their capacity of predisposing to tumor development and its dissemination (2-4).

Both physiological and pathological angiogeneses are characterized by the sprouting of new blood vessels from extensions of the existing vasculature mediated by the balance between proangiogenic and antiangiogenic factors. This regulation involves the interaction between endothelial cells, smooth muscle cells and pericytes, in addition to various stromal components that play an important role in the angiogenic process (5). In tumors, angiogenesis-regulating molecules are produced and disseminated mainly by the neoplastic cells themselves and by leukocytes, mast cells, fibroblasts or macrophages found in proximity to the tumor (3,6,7). Currently, anti-angiogenic therapy can influence the management of many tumors by different mechanisms (8).

Tumor-associated macrophages (TAM) are the main cellular component in the stroma of many tumors (3). Although macrophages can serve as negative or positive regulators of tumor growth (3,9), they have been the focus of studies due to their functions performed on behalf of the tumor, including the expression of growth factors and metalloproteinases, suppression of adaptive immunity and promotion of angiogenesis (3,9,10), demonstrating their importance for tumor progression. TAM are able to induce the formation of new blood vessels through the expression of potent proangiogenic molecules, including vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), transforming growth factor beta (TGFβ) and angiogenic chemokines such as monocyte chemotactic protein-1 (CCL2/MCP-1), whose levels are associated with the accumulation of TAM (3,10). In addition, TAM exert mobilization of angiogenic growth factors embedded in the extracellular matrix and interact with endothelial cells to improve blood vessel growth (4).

It is known that tumor microenvironment plays a key role in the tumor progression, including OSCC (11-13). Nevertheless, there are few studies focusing on angiogenesis and TAM regarding this cancer. Thus, the aim of the present study was to assess the microvascular density (MVD) and infiltrating macrophage density (IMD) at different histological grades of OSCC.

Material and Methods

Tissue Samples

After approval of the study by the Ethics Committee, OSCC biopsies from 25 patients were selected. The
anatomopathological records and clinical charts were obtained from the archives of two Surgical Pathology Services (Dental School, Federal University of Bahia and Dental School, Federal University of Rio de Janeiro). None of the patients had undergone chemo- or radiotherapy. After staining with hematoxylin–eosin, the material was reviewed by an experienced pathologist in order to classify the cases according to the World Health Organization International Histological Classification of Tumors (14).

**Immunohistochemistry**

Formalin-fixed and paraffin-embedded material was cut into 3-µm thick sections, deparaffinized in xylene and rehydrated in ethanol and water. For antigen retrieval, tissue sections were boiled in citrate buffer, pH 6.0, for 40 min in a water bath at 95–97°C. For immunohistochemical detection, antibodies to von Willebrand factor (Dako A/S, Copenhagen, Denmark; dilution 1:200) and CD68 (clone KP1; Dako A/S; dilution 1:100) were applied overnight using the EnVision™ System (Dako Corporation, Carpinteria, CA, USA). The immunohistochemical reactions were developed with diaminobenzidine (Dako Corporation) as chromogens to visualize the peroxidase activity and slides were counterstained with Harris hematoxylin.

**Microvascular Density and Infiltrating Macrophage Density**

For the analysis of immunostaining, the slides were examined and photographed using a Nikon microscope (Eclipse E800; Nikon, Tokyo, Japan). The observation was blind with respect to the clinical and histological outcomes. For the MVD, von Willebrand factor-positive staining microvessels were counted. The slides were first examined at low magnification (100x) for identification of highly vascularized areas (hot spots). Finally, images of 10 hot spots were acquired at 400x magnification and the MVD was calculated by counting the average number of microvessels in 10 fields (15) with tissue area measured in mm². Any positively stained individual endothelial cell or endothelial cell cluster that was clearly separate from adjacent microvessels, tumor cells or other connective tissue elements was considered as a countable microvessel. The presence of a lumen was not necessarily required for microvessel counting and large vessels with a muscle wall or a lumen larger than 50 µm were excluded (16). For the IMD, CD68-positive cells were defined as macrophages and analyzed as described by Ueno et al. (2). This method consists of quantifying positively stained cells by examination of the five most confluent fields (hot spots). Next, the mean of the three largest counts obtained for the five fields was calculated and the IMD was reported per tissue mm².

**Statistical Analysis**

Differences between groups were evaluated by one-way ANOVA followed by Newman-Keuls multiple-comparison test. The correlation between MVD and IMD was analyzed using the Pearson’s correlation coefficient. A p<0.05 value was considered to be statistically significant. The GraphPad Prism software, version 4.0 (GraphPad Software, Inc., San Diego, CA, USA) was applied for statistical calculation.

**Results**

The clinicopathological characterization of the OSCC tumors is shown in Table 1.

The mean MVD in the tumors was 48.38 microvessels/mm² (Fig. 1A–C). Comparison among tumors showed low vascularity in well-differentiated OSCC compared with moderately and poorly differentiated carcinomas (Table 2). CD68-positive cells were observed in all tumors. The mean IMD in the tumors was 91.47 cells/mm². These cells were present as clusters or distributed along the inflammatory

<table>
<thead>
<tr>
<th>Clinical variable</th>
<th>OSCC</th>
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<tbody>
<tr>
<td>Age</td>
<td>% (n)</td>
</tr>
<tr>
<td>&lt;60</td>
<td>52 (13)</td>
</tr>
<tr>
<td>≥60</td>
<td>44 (11)</td>
</tr>
<tr>
<td>Unknown</td>
<td>4 (1)</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>40 (10)</td>
</tr>
<tr>
<td>Male</td>
<td>60 (15)</td>
</tr>
<tr>
<td>Tumor site</td>
<td></td>
</tr>
<tr>
<td>Tongue</td>
<td>28 (7)</td>
</tr>
<tr>
<td>Floor of mouth</td>
<td>16 (4)</td>
</tr>
<tr>
<td>Vestibule of the mouth</td>
<td>16 (4)</td>
</tr>
<tr>
<td>Buccal mucosa</td>
<td>12 (3)</td>
</tr>
<tr>
<td>Gingiva</td>
<td>12 (3)</td>
</tr>
<tr>
<td>Hard palate</td>
<td>8 (2)</td>
</tr>
<tr>
<td>Retromolar area</td>
<td>4 (1)</td>
</tr>
<tr>
<td>Unknown</td>
<td>4 (1)</td>
</tr>
<tr>
<td>Histologic tumor grading*</td>
<td></td>
</tr>
<tr>
<td>Well differentiated</td>
<td>40 (10)</td>
</tr>
<tr>
<td>Moderately differentiated</td>
<td>36 (9)</td>
</tr>
<tr>
<td>Poorly differentiated</td>
<td>24 (6)</td>
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*According to the WHO International Histological Classification of Tumors.
infiltrate OSCC (Fig. 1D–F). The IMD was similar in all groups (Table 2). Pearson’s correlation test revealed no significant correlation between MVD and IMD regarding both markers (p=0.870).

Discussion

In the present study, the von Willebrand factor (vWF) was used as an endothelial cell marker. This protein is synthesized by endothelial cells and plays an important

Figure 1. Distribution of tumor microvessels and CD68-positive macrophages in the investigated tumors. Tumor microvessels in a well-differentiated (A, B) and a moderately differentiated (C) oral squamous cell carcinoma (OSCC). CD68-positive macrophages in a well-differentiated (D), a moderately differentiated (E) and a poorly differentiated (F) OSCC.
role in hemostasis, promoting the adhesion of the first platelets to the vascular subendothelium (17). This fact permits the interpretation of VWF as an angiogenesis marker (16,18,19). In addition, many investigators strongly advocate MVD as an important factor for establishing the prognosis of different cancers such as breast cancer (18) and oral cancer (16,19,20).

The findings of the present study were similar to those reported by Sharma et al. (19), who found significantly higher MVD in moderately differentiated OSCC than in well differentiated OSCC. This might contribute to the OSCC prognosis, as this lesion may be related to the degree of tumor vascularization (1,16). In this respect, an increase of angiogenesis in patients with OSCC may be related to high levels of interleukin-10 (IL-10), a regulatory molecule for angiogenesis in various cancers that leads to suppression of the immune response in patients (21).

Although the present results showed differences in MVD according to the degree of tumor differentiation, the literature has not yet reached a consensus regarding the influence of angiogenesis on the rate of recurrence, tumor invasion and regional metastasis. Fernandez et al. (22) investigated angiogenesis in OSCC of the tongue and found no evidence that tumor vascularization can be used as an independent factor to evaluate the possibility of recurrence or patient survival. In a similar study, Guttmann et al. (16) found a significant correlation between an increased number of microvessels and tumor invasion to the adjacent musculature and lymph node metastasis. Shpitzer et al. (1) counted vessels in OSCC of the tongue and established a positive correlation with the presence of metastases, indicating angiogenesis to be an independent factor for the prediction of lymph node metastasis and for the decision-making regarding the therapy to be used. According to these authors, patients with no clinical signs of lymph node involvement who present high vascularization rates should be submitted to adjuvant therapy and not only to surgical resection.

In addition to blood vessels, solid tumors contain a stromal cellular component that basically consists of fibroblasts and immunocompetent cells, especially macrophages and lymphocytes (3). However, the biological significance of the presence of macrophages has not been completely established, mainly because these cells often have contradictory functions, exerting pro and antitumor actions, depending on their polarization to a M1 or M2 macrophage. Thus, TAM have been described as M2-polarized macrophages exhibiting a protumoral phenotype (3). In addition, diverse studies report their influence on angiogenesis and lymphangiogenesis (4,7).

TAM are recruited as monocytes from blood in the direction of the tumor by molecules secreted by both malignant and stromal cells in tumors, including VEGF, colony stimulating factor-1 (CSF-1) and chemokines such as CCL2/MCP-1, placental growth factor (PIGF, PGF), macrophage inflammatory protein-1α (CCL3/MIP-1α), macrophage inflammatory protein-1β (CCL4/MIP-1β) and regulated on activation normal T-cell expressed and secreted (CCL5/RANTES) (3,10). Within the tumor, these macrophages undergo marked phenotypic change and express numerous chemokines and angiogenesis/lymphangiogenesis-promoting factors that contribute to the onset of tumorigenesis and metastasis, such as VEGF, TGF-β, PDGF, IL-8, IL-10, prostanoids, reactive oxygen species, in addition to enzymes related to the degradation of extracellular matrix (3,10).

A high frequency of macrophages as observed in the present investigation has also been reported in other studies and has been correlated with a poor prognosis in OSCC (11,12). Despite this aspect, a relationship between increased MVD and the presence of macrophages has been demonstrated in previous studies performed in different types of cancer (2,7,11,20,23). In contrast, in the present study, no correlation could be established between IMD and MVD. These results partially disagree with those reported by El-Rouby (20), who observed an association between an increase in the number of TAM and increased angiogenesis in oral verrucous cell carcinomas and OSCC, although, similarly to ours results, with no significant difference in the area percentage of TAM among different histological

<table>
<thead>
<tr>
<th>Density</th>
<th>Well differentiated OSCC (n=10)</th>
<th>Moderately differentiated OSCC (n=9)</th>
<th>Poorly differentiated OSCC (n=6)</th>
<th>p-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Microvascular density</td>
<td>$40.34 \pm 9.10^{b}$</td>
<td>$56.52 \pm 12.72^{b}$</td>
<td>$51.45 \pm 19.59$</td>
<td>0.046</td>
</tr>
<tr>
<td>Infiltrating macrophage density</td>
<td>$83.75 \pm 32.24$</td>
<td>$88.23 \pm 27.70$</td>
<td>$106.6 \pm 34.39$</td>
<td>0.381</td>
</tr>
</tbody>
</table>

OSCC: Oral squamous cell carcinoma. *Compared by one-way ANOVA and Newman-Keuls multiple-comparison test with statistically significant if p<0.05. Values are shown as the mean ± standard deviation.
grades of OSCC.

In this study, there was no significant difference in IMD between the OSCCs with different histological grades. A possible explanation could be the use of CD68, a pan-macrophage marker that does not distinguish the presence of M2-polarized macrophages that are much more relevant for an immunosuppressive cancer microenvironment and its progression (24). Similarly, no correlation was found between IMD and MDV. It is important to highlight that the potential to induce angiogenesis in OSCC is related not only to macrophages, but also other stromal cells as fibroblasts, mast cells and tumor epithelium (19).

In conclusion, angiogenesis and TAM play a key role in different histological grades of OSCC. There was a significant difference between well differentiated and moderately differentiated tumors regarding angiogenesis, which could indicate their influence in OSCC. However, the lack of correlation between MVD and IMD could suggest that angiogenesis does not depend on the number of macrophages present in OSCC, but their predominant phenotype. Further studies involving distinct phenotypes of macrophages should be performed for better understanding influence of TAM on tumor angiogenesis.

Acknowledgments

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