SCREENING FOR EXOPOLYSACCHARIDE-PRODUCING BACTERIA FROM SUB-TROPICAL POLLUTED GROUNDWATER

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(With 1 figure)

ABSTRACT

A selection of exopolysaccharide (EPS) – producing bacterial strains was conducted in groundwater adjacent to an old controlled landfill in the City of São Carlos (São Paulo, Brazil). The strains were isolated in P and E media under aerobic and microaerophilic conditions at 25°C. A total of 26 strains were isolated and based on the mucoid mode of the colonies, 6 were selected and their morphological, physiological and biochemical aspects were characterized. All strains presented pigmentation, ranging from yellow to orange and from pink to salmon, with a shiny glistening aspect in all tested media. Strains Lb, Lc and Lg, which excelled the others with regard to the mucoid mode of the colonies, were selected to be cultured in E medium with alternate sucrose and glucose as carbon sources in anaerobiosis at 25°C to analyze the production of EPS. Strains Lc and Lg were classified as being of order Actinomycetales, suborder Corynebacterineae. Lg strain was identified as Gordonia polyisoprenivorans and Lc strain did not correspond to a known description and therefore a more detailed study is under preparation. Considering all ecological aspects and the metabolic potential associated with the microorganisms of the environment studied, as well as the capacity to produce pigment and EPS, and the presence of G. polyisoprenivorans, a rubber degrader bacterium, the potential of the groundwater analyzed is evident as a source of microorganisms to be utilized in studies related to environmental remediation.

Key words: bacteria, exopolysaccharide, groundwater, landfill.

RESUMO

Seleção de bactérias produtoras de exopolissacarídeos de águas subterrâneas contaminadas de ambiente subtropical

Foi realizada a seleção de linhagens bacterianas produtoras de exopolissacarídeos (EPS) de águas subterrâneas adjacentes ao antigo aterro controlado da cidade de São Carlos (São Paulo, Brasil). O isolamento foi realizado nos meios P e E em condições de aerobiose e microaerofilia a 25°C. Foram isoladas 26 linhagens e, com base na mucosidade das colônias, 6 foram escolhidas e caracterizadas quanto a aspectos morfológicos, fisiológicos e bioquímicos. As colônias das linhagens apresentaram aspecto brilhante em todos os meios testados, com pigmentação variando de amarela a laranja e de rosa a salmão. As linhagens Lb, Lc e Lg, que se destacaram das demais pela mucosidade das colônias, foram selecionadas para cultivo em meio E suplementado alternadamente com glicose e sacarose como fonte de carbono, em anaerobiose a 25°C, para a verificação da produção de EPS. As linhagens Lc e Lg foram classificadas como sendo da ordem Actinomycetales, subordem Corynebacterineae. A
INTRODUCTION

Programs search for naturally occurring microorganisms that have better pollutant degradation kinetics, attack a wider range of pollutant compounds, and do so over a broader range of microbial conditions, are attempting to expand the range of microorganisms used for bioremediation of contaminated sites (Atlas, 1995).

Although technical difficulties in sampling the subsurface environments have long limited the ecology study of groundwater and until recently, most people thought that these environments were virtually sterile (Gibert et al., 1994), there is no longer doubt that a large number and diversity of viable organisms exist in the subsurface and that their metabolic activities contribute to the cycling of energy and materials in this environment (Fliermans & Balkwill, 1989; Butler et al., 1997). Screening programs of the subsurface environment showed that groundwater bacteria are potentially able to degrade many natural and xenobiotic compounds such as pesticides, fertilizers, and industrial chemicals which can reach aquifers making these microorganisms attractive to in situ bioremediation (Gounot, 1994). However, if we are to utilize in situ bioremediation, more basic research is needed to better characterize the microbial populations and activities in the subsurface (Gounot, 1994; Zheng & Kellogg, 1994).

On solid surfaces exposed to aqueous environments bacterial growth is seen as biofilms in which the microbial cells are associated with large amounts of exopolysaccharides (EPS) (Sutherland, 1990). Exopolysaccharides are essential to the biological success of most bacteria living within the biofilm in the varied natural environments in which they are observed since they can concentrate nutrients from water flow and protect the bacteria from antibacterial agents and from predators (Costerton, 1985). The EPS investigations are in an increasing demand since the biofilm degradative communities can be used for in situ bioremediation of contaminated sites (Lappin-Scott & Costerton, 1992).

Studies conducted on the old landfill in the town of São Carlos (SP, Brazil) revealed the presence of a pollution plume (Ellert et al., 1990) as well as the presence of an abundant microflora dominated by bacterial populations (Fusconi & Godinho, 1999). This paper deals with a search for indigenous exopolysaccharide-producing bacterial strains with potential usage in bioremediation programs through biofilms.

MATERIAL AND METHODS

Strains isolation and maintenance

The sampling for bacterial isolation from groundwater in the area of the old landfill of São Carlos (located in the recharge area for the Guarani aquifer) was carried out according to Fusconi & Godinho (1999). The heterotrophic bacterial strains were isolated at 25°C under aerobic and microaerophilic conditions in P medium (g/L) (Kölbel-Boelke et al., 1988): peptone, 1.0; glucose, 0.1; K,HPO₄, 3H₂O, 0.1; FeSO₄.x7H₂O, 0.02; and in E medium (Clark et al., 1981): a) basic medium (g/960 mL): KH₂PO₄, 2.7; K₂HPO₄, 13.9; yeast extract, 0.5; b) solution 1: 10% sucrose; c) solution 2 (Wolin solution) (g/L): EDTA, 0.5; MgSO₄.xH₂O, 6.14; MnSO₄.xH₂O, 0.56; NaCl, 1.0; CaCl₂.x2H₂O, 0.12; ZnSO₄, 0.1; FeSO₄.x7H₂O, 0.18; CuSO₄.x5H₂O, 0.02; AlK(SO₄)₂.x12H₂O, 0.02; Na₂MoO₄.x2H₂O, 0.01; H₂BO₃, 0.01; Na₂SeO₃, 0.007; NiCl₂, 0.002; d) solution 3: 3% (NH₄)₂SO₄; e) solution 4: 5% MgSO₄ solution. The solutions were autoclaved separately and added aseptically in the indicated order, 10 mL/L each, to the basic medium. Microaerophilic conditions were produced by a combination of iron and sodium bicarbonate (Jurgensen, 1981), and the pH of both media was...
7.0. After a maximum period of incubation of 7 days. The colonies were picked at random and replated for purification onto the respective media. Isolated strains were stored on nutrient agar at 4°C.

Selection of exopolysaccharide-producing bacteria

The selection of EPS-producing bacterial strains involved 5 steps:

**Step 1:** Strains whose colonies were found to be more mucoid in the isolation media (P and E media) were chosen. **Step 2:** The selected strains were phenotypically characterized according to Murray *et al.* (1994) and Smibert & Krieg (1994), involving the following tests: colonies and cells morphology, pigment production on nutrient agar, P medium, E medium and 50% BHA medium (Difco), catalase activity, acid production from glucose, relationship to oxygen, presence of spores and growth on different culture media (50% BHI, 50% BHA, solid E medium and solid and liquid P medium). The growth on liquid media was analyzed under aerobic and anaerobic conditions. For the anaerobic growth NaNO₃ (1 g/L) was added as the final electron acceptor and the atmosphere was exchanged through bubbling with nitrogen gas for 1 h for each liter of culture medium. **Step 3:** The strains were cultured in anaerobiosis, in media, which contributed to the production of EPS (E and BHA 50%). The production of EPS was identified with Alcian blue 8GX staining (Murray *et al.*, 1994) of cells from 40 h cultures on E medium under aerobic and anaerobic conditions. Another screening was performed based on the mucoid mode of colonies (Chan *et al.*, 1984) and EPS production. **Step 4:** The three strains, selected in step 3, were cultured in anaerobiosis at 25°C in a refrigerated orbital shaker (100 rpm). E medium was used with alternate, sucrose and glucose as carbon sources. According to Linton (1990) and Sutherland (1990), EPS production is affected by the cultivation conditions (i.e. substrate). The bacteria were cultured in 100 ml flasks, containing 50 ml of culture medium and 5% inoculum. Samples (2.5 ml) were taken during the cultivation for optical density determination and the production of EPS during the stationary phase was analyzed as described in step 3. Strains which exhibited better results relating to the mucoid mode pattern in solid medium and growth in E medium with at least one carbon source were selected. **Step 5:** Selected EPS-producing bacterial strains were identified by phylogenetic analysis at the Fundação Tropical de Pesquisas e Tecnologia André Tosello (Campinas, SP) and were stored for further investigations.

**RESULTS**

A total of 26 bacterial strains were isolated from groundwater samples in the landfill area of São Carlos City.

In step 1, 6 isolated bacterial strains with mucoid colonies were chosen (La, Lb, Lc, Lf, Lg and Lh) and the microbial characteristics are summarized in Table 1.

![TABLE 1](image)

**TABLE 1**

Microbial characteristics of bacterial strains isolated from polluted groundwater.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Cell morphology</th>
<th>Cell size (µm)</th>
<th>Gram staining</th>
<th>Catalase activity</th>
<th>Acid production with glucose</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Aerobiosis</td>
</tr>
<tr>
<td>La</td>
<td>coccobacillus</td>
<td>1.0 × 2.4</td>
<td>–</td>
<td>+</td>
<td>D</td>
</tr>
<tr>
<td>Lb</td>
<td>coccobacillus</td>
<td>0.8 × 2.1</td>
<td>–</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Lc</td>
<td>pleomorphic</td>
<td>1.0 × 3.0</td>
<td>+</td>
<td>+</td>
<td>_</td>
</tr>
<tr>
<td>Lf</td>
<td>rod</td>
<td>0.4 × 2.2</td>
<td>–</td>
<td>+</td>
<td>_</td>
</tr>
<tr>
<td>Lg</td>
<td>pleomorphic</td>
<td>1.0 × 2.4</td>
<td>+</td>
<td>+</td>
<td>D</td>
</tr>
<tr>
<td>Lh</td>
<td>rod</td>
<td>0.7 × 2.2</td>
<td>–</td>
<td>+</td>
<td>_</td>
</tr>
</tbody>
</table>

D: dubious; +: positive; –: negative.
The cells from strains La and Lb were coccobacillus and from strains Lf and Lh were rods; the strains Lc and Lg were pleomorphic and predominantly arranged in clusters while the other strains were isolated or paired cells. The results of the physiological and biochemical tests showed the absence of sporulating bacteria, the presence of facultative anaerobic metabolism and positive catalase activity in all strains. Only Lb bacteria produced acid in the presence and absence of oxygen. Acid production by the remaining strains occurred only under anaerobiosis. All strains presented pigmentation, ranging from yellow to orange and from pink to salmon, with a shiny glistening surface aspect in all tested media. La, Lb, Lc and Lg strains showed physiological versatility growing in all media and under all conditions under which they were tested (data not shown).

In step 3 production of EPS was observed in all strains, except La. The development of a mucoid aspect, indicating a possible EPS development by the strains, was more evident in strains Lb and Lc when cultured on E and 50% BHA media, and in strain Lg when cultured on E medium.

Due to the mucoid mode, strains Lb, Lc and Lg were selected and cultured in E medium supplemented with glucose or sucrose (step 4). The three selected strains grew with both carbon sources, being the highest optical density observed in Lg strain cultured in E medium with sucrose (OD = 1.185) and in E medium with glucose (OD = 1.170), followed by Lc strain cultured in E medium with glucose (OD = 0.792). The lowest growth was observed in Lc strain cultured in E medium with sucrose (OD = 0.200) (Fig. 1). The growth of Lc strain in E medium with glucose, considering the highest optical density obtained, was approximately four times higher than in E medium with sucrose. The production of EPS, tested with Alcian Blue, was positive for all tested strains and with both carbon sources (Table 2).

Lc and Lg strains, which exhibited better results relating to the mucoid mode pattern in solid medium and growth in E medium with at least one carbon source, were classified as being of order Actinomycetales, suborder Corynebacterineae. Lg strain was identified as *Gordonia polyisoprenivorans* and Lc strain did not correspond to a known bacterial description and therefore a more detailed study is under preparation.

**DISCUSSION**

The selection process of EPS-producing bacterial strains was fundamentally based on the development of mucoid aspect of strain colonies. This characteristic, according to Chan *et al.* (1984) and Junkins & Doyle (1992), who studied the production of EPS, respectively by *Pseudomonas aeruginosa* and enterohemorrhagic *Escherichia coli*, indicates a high production of EPS bacterial strains. Other studies in which the development of the mucoid mode was related to EPS production by bacteria have also been detected on *Pseudomonas*. Uhlinger & White (1983) and Robertson & Firestone (1992) selected strains on the basis of the mucoid aspect of bacterial colonies cultured on solid media. The former authors worked on *Pseudomonas* sp. isolated from soil and related EPS production to protection against desiccation, and the following authors studied the relationship between the physiological state and EPS production of *Pseudomonas atlantica*.

During the selection process the presence of pigmentation in all strains was detected. This was considered an important characteristic, since bacterial pigmentation in contaminated water has been attracting the attention of researchers, since it can give selective advantages which contribute to higher resistance and tolerance to microorganisms facing toxic substances (Nair *et al.*, 1992; Arrage *et al.*, 1993).

Lc and Lg strains excelled the other isolated strains in presenting Gram-positive pleomorphic cells arranged in clusters and, as Lb strain, presented better results with regard to the mucoid mode of colonies in E media and BHA 50%. When cultured in liquid E medium, the three strains (Lb, Lc and Lg) grew both with glucose and sucrose as carbon source, and a direct observation of the culture medium, during their cultivation, revealed a mucoid mode of growth in liquid media. This growth pattern both in solid and liquid media indicated a production of exopolysaccharide by bacteria, confirmed by the coloration with Alcian blue. Chan *et al.* (1984) observed the same growth pattern for *Pseudomonas aeruginosa* in liquid MVBM and BHI media and attributed this fact to the presence of cell embedded in a fiber EPS matrix, which promoted the adhesion between cells and cells to substrate (flask), forming microcolonies.
Fig. 1 — Growth of Lb, Lc and Lg strains with E medium supplemented with glucose (Eglu) and sucrose (Esuc). Growth under anaerobic conditions at 25°C in shaker at 100 rpm.
In groundwater, the presence of polymers on the surface of bacterial cells was detected by some investigators (Ghiorse & Balkwill, 1983; Hirsh & Rades-Rohkohl, 1983) who related it to the increased adhesion to soil particles and to a higher probability of nutrient retention in oligotrophic environments. Recent studies found out that the property of accumulating nutrients from the environment of EPS-producing bacteria is due to the presence of different reactant groups in the structure of biofilm and thus the same can be used to remove different organic pollutants, such as agricultural chemicals (e.g. the herbicide diclofop methyl) (Wolfaardt et al., 1994; Wolfaardt et al., 1998) and metals from the environment (Shen et al., 1993).

This work shows that groundwater adjacent to the old controlled landfill in the City of São Carlos can be a potential source of microorganisms for studies related to environmental remediation in the cited area, and that the bacteria that produce exopolysaccharide were selected successfully. Among the latter, it is important to emphasize the selection of *Gordonia polyisoprenivorans*, a species recently described by Linos et al. (1999). *G. polyisoprenivorans*, as other taxa of suborder Corynebacterineae (which Lc strain belongs to), has an important role in bioremediation and biodegradation of environmental pollutants, in particular for the capability to degrade rubber (Linos et al., 1999).

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REFERENCES


EXOPOLYSACCHARIDE-PRODUCING BACTERIA


