EXTENDED-SPECTRUM β-LACTAMASES IN KLEBSIELLA SPP AND ESCHERICHIA COLI OBTAINED IN A BRAZILIAN TEACHING HOSPITAL: DETECTION, PREVALENCE AND MOLECULAR TYPING

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ABSTRACT

His study was performed to compare the methods of detection and to estimate the prevalence of extended-spectrum β-lactamases (ESBL) among Klebsiella spp and E. coli in a university hospital in southern Brazil. We also used a molecular typing method to evaluate the genetic correlation between isolates of ESBL K. pneumoniae. Production of ESBL was investigated in 95 clinical isolates of Klebsiella spp and Escherichia coli from Hospital de Clínicas de Porto Alegre, using Kirby-Bauer zone diameter (KB), double-disk diffusion (DD), breakpoint for ceftazidime (MIC CAZ), increased zone diameter with clavulanate (CAZ/CAC) and ratio of ceftazidime MIC/ceftazidime-clavulanate MIC (MIC CAZ/CAC). Molecular typing was performed by DNA macrorestriction analysis followed by pulsed-field gel electrophoresis. The KB method displayed the highest rates of ESBL (up to 70% of Klebsiella and 59% of E. coli), contrasting with all the other methods (p < 0.05). The confirmatory methods (DD, MIC CAZ, CAZ/CAC and MIC CAZ/CAC) showed a range of ESBL production from 8 to 13% for E. coli and from 33 to 40% for Klebsiella species. Therefore, the KB method was useful only as a screening method as it provided several false positive results. Molecular typing of 17 ESBL K. pneumoniae indicated that the isolates had no clonal relation. We found a good correlation among the confirmatory methods for ESBL detection although the methods which evaluate inhibition of the β-lactamase by clavulanate appeared to be more specific. The high prevalence of ESBL Klebsiella in our hospital is probably due to individual selection of resistant strains rather than the transmission of a common strain.

Key words: ESBL, Klebsiella, resistance, molecular typing.

INTRODUCTION

One of the most prevalent mechanisms of bacterial resistance among Gram-negative bacteria is the production of β-lactamases. These enzymes comprise a family with high diversity and many schemes for their classification have been proposed (2). Extended-spectrum β-lactamases (ESBLs) are a group of clinically very important β-lactamases because they are able to hydrolyze the extended-spectrum cephalosporins (ceftazidime, cefotaxime, ceftriaxone) and monobactams (aztreonam). ESBLs are included in the 2nd group of Bush classification (2) and originally have derived from TEM and SHV enzymes although other classes of ESBL have been described recently (1). Interestingly, the mutations that expand the spectrum of these β-lactamases usually increase their sensitivity to β-lactamase inhibitors (clavulanate and sulbactam) (8,27).

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The prevalence of ESBL varies according to different regions or to different hospitals (6,26). Piddock et al. (24) studied more than 3,000 *Enterobacteriaceae* from 43 British hospitals, and found that only 1% of isolates produced ESBL. On the other hand, Lyon et al. (16) found an increase of overall resistance and specifically in ESBL production between 1990 and 1995 in isolates from blood culture in Hong Kong, and emphasize the importance to use appropriate screening methods in order to detect these enzymes since routine diffusion tests might give false negative results. Livermore and Yuan (14), found that 23% of *Klebsiella* species isolated from intensive treatment units of 35 European hospitals were ESBL producers. They also described that 33% of ESBL isolates were reported as susceptible to third generation cephalosporins and monobactams, and this pose as a problem, since treatment of ESBL producers with those agents were associated with failure (2,9). In Brazil there is scarce data about ESBL production in Gram-negative bacteria. In a study using nosocomial isolates, Gales et al. (7) found that 39% of *K. pneumoniae* isolated from São Paulo were ESBL producers.

Members of *Enterobacteriaceae* producing ESBL constitute a serious threat to current β-lactam therapy, because the *in vitro* detection of ESBL expression has proved to be troublesome and there is no consensus about the best method to detect ESBL. Practical methods such as double-disc test (5,16), E-test double strip (3,29) and the modified breakpoints for standard susceptibility tests (22,24) have been used by many studies.

Another important aspect is the epidemiological aspect of ESBL producers. Although Cotton et al. (4) had described an outbreak due to horizontal transmission between patients, ESBL can also arise from selection in individual microbiota. The use of cephalosporins, mainly ceftazidime, has been indicated as one of the major risk factors for acquisition of ESBL (7,18,21,25). Lucet et al. (15) pointed other risk factors for ESBL acquisition which are associated with the severity of illness of patient requiring long stay in intensive care units and presence of catheters.

This study was performed to compare the methods of detection and to estimate the prevalence of ESBL *Klebsiella* spp and *E. coli* in a university hospital in southern Brazil (HCPA). In addition we also used DNA macrorestriction analysis by PFGE to evaluate the genetic correlation between isolates of ESBL *K. pneumoniae*.

**MATERIALS AND METHODS**

Isolates of *E. coli* and *Klebsiella* spp were suspended in Mueller-Hinton broth (Oxoid, Basingstoke) to an optical density equal to 0.5 MacFarland standard and this suspension was used to inoculate Mueller-Hinton agar (Oxoid, Basingstoke) plates. The following methods were performed to detect ESBL production:

- **Kirby-Bauer zone diameter (KB):** isolates with inhibition zones of ≤ 22 mm to ceftazidime (CAZ) and/or ≤ 25 mm to ceftriaxone (CRO) and/or ≤ 27 mm to aztreonam (ATM) 30 µg disks (Oxoid, Basingstoke) on disk diffusion method were considered as suspicious for ESBL producers (19).

- **Double-disk diffusion (DD):** individual disks containing 30 µg of CAZ, CRO and ATM (Oxoid, Basingstoke) were placed onto the plate in a distance of 1.5 cm (edge to edge) from an amoxicillin/clavulanic acid disk (20/10 µg). An enhanced zone of inhibition between any one of the β-lactam disks and the disk containing clavulanic acid was interpreted as a positive result.

- **Ratio of ceftazidime MIC/ceftazidime-clavulanic acid MIC (MIC CAZ/CAC):** this method was performed using an E-test ESBL strip (AB Biodisk, Solna) carrying two gradients: on the one end ceftazidime (0.05 to 32 µg/mL), and on the opposite end ceftazidime (0.125 to 8 µg/mL) plus clavulanic acid in a fixed concentration (4 µg/mL). We considered a ratio of MIC CAZ / MIC CAC equal or greater than 8 as presumptive for ESBL production as proposed by the E-test ESBL strip manufacturer and some studies (29).

- **Breakpoint to ceftazidime (MIC CAZ):** this procedure used the result of the E-test strip as above and considered a MIC ≥ 2 µg/mL for ceftazidime as an indication of ESBL production (19).

- **Increased zone diameter with clavulanic acid (CAZ/CAC):** a ≥ 5 mm increase in zone diameter for ceftazidime disk (30 µg) added of clavulanic acid (10 mg) compared with a ceftazidime disk (Oxoid, Basingstoke) alone indicated ESBL (20).

The results of methods were compared by the Fisher’s Exact Test.

DNA Macrorestriction analyses of *Klebsiella pneumoniae* was performed using DNA digestion followed by pulsed-field gel electrophoresis as stated (10). Briefly, the bacteria was embedded in agarose (Gibco BRL, Grand Island) blocks, transferred to a lysis buffer containing proteinate K (Sigma, St. Louis) for 12 h, and digested with restriction endonuclease SpeI (Gibco BRL). Electrophoresis was carried out in a CHEF DR II apparatus (Bio-Rad, Richmond). The gels were run for 23 h, with switch time of 5 to 25 s. Lambda ladder (48.5 Kb, Sigma) was used as molecular weight marker. Fragments were stained with ethidium bromide (Sigma) and photographed. Visual comparisons were made and the criteria of Tenover et al. (28) was used to establish the relationship among the isolates.

**RESULTS**

A total of 95 clinical isolates of *E. coli* (39 isolates) and *Klebsiella* spp (47 *K. pneumoniae* and 9 *K. oxytoca*) recovered from 94 patients hospitalized in HCPA during a five months period (June to October, 1998) were used in this study. Overall, 20 samples
were obtained from surgical wounds, 14 from sputum, 13 from blood and the remain from other sites (body fluids and catheters). We were not able to interpret the results of MIC CAZ/CAC for two isolates of *K. pneumoniae* because their MIC for ceftazidime and ceftazidime plus clavulanic acid was above the maximum limit of the strip. These isolates were excluded from the general comparison between methods. According to the NCCLS breakpoints for CAZ, CRO and ATM by the disk-diffusion method (KB) the number of ESBL producers was very high among our isolates: up to 70% for *Klebsiella* sp and 59% for *E. coli*. Many isolates positive for ESBL by this method were not classified as positive by none of the other methods used in this study as one could expect for a typical screening method.

The confirmatory methods (DD, MIC CAZ and MIC CAZ/CAC) showed a range of ESBL production similar, from 8 to 13% for *E. coli* and from 33 to 40% for *Klebsiella* species (Table 1). The efficiency of screening for ESBL-producing isolates according to NCCLS (KB method) with disks of CAZ, CRO and ATM was evaluated for isolates of *K. pneumoniae* according to the other methods. In general, non-resistant isolates were not ESBL producers with the exception of three isolates which were positive for ESBL according to the other methods and susceptible to ceftazidime. It has to be mentioned that one of these isolates was positive only in the DD method. Considering ESBL production in *Klebsiella* species by methods which evaluate inhibition of the β-lactamase enzyme by clavulanic acid (double-disk method and MIC CAZ/CAC method) one will observe very similar results: 19 positive isolates in DD method and 18 isolates in MIC CAZ/CAC (Table 1). Detection of ESBL production in *Klebsiella* by DD and MIC CAZ indicated that 19 and 22 isolates respectively were ESBL positive (not statistically significant, p=0.6919). The number of isolates positive by MIC CAZ/CAC and MIC CAZ were also very similar (p=0.847) (Table 1).

The occurrence of ESBL in *E. coli* was not identical whether the DD and MIC CAZ/CAC methods were compared (Table 1).

**Table 1.** Positive results of methods for ESBL production in *E. coli* and *Klebsiella* spp.

<table>
<thead>
<tr>
<th>CAZ</th>
<th>No. of isolates</th>
<th>KB</th>
<th>DD</th>
<th>MIC CAZ/CAC</th>
<th>MIC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>38</td>
<td>19</td>
<td>18</td>
<td>22</td>
</tr>
<tr>
<td><em>Klebsiella</em> species</td>
<td>54</td>
<td>(70%)</td>
<td>(35%)</td>
<td>(33%)</td>
<td>(40%)</td>
</tr>
<tr>
<td><em>K. pneumoniae</em></td>
<td>45</td>
<td>31</td>
<td>17</td>
<td>16</td>
<td>20</td>
</tr>
<tr>
<td>(69%)</td>
<td>(38%)</td>
<td>(36%)</td>
<td>(44%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>K. oxytoca</em></td>
<td>9</td>
<td>7</td>
<td>2</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>(78%)</td>
<td>(22%)</td>
<td>(22%)</td>
<td>(22%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>E. coli</em></td>
<td>39</td>
<td>23</td>
<td>5</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>(59%)</td>
<td>(13%)</td>
<td>(8%)</td>
<td>(13%)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Kirby-Bauer disk diffusion; * Double-disk diffusion; * Ratio of ceftazidime MIC/ceftazidime-clavulanic acid MIC; * Breakpoint to ceftazidime.

Two isolates were positive by DD method and negative according to MIC CAZ/CAC. It is of importance to notice that these isolates displayed a difference between MICs of ceftazidime with and without clavulanic acid although this difference did not reach the cutoff ratio. Therefore, the isolates were not completely indifferent to the presence of the β-lactamase inhibitor.

Since January 1999 the NCCLS proposed a confirmatory test (CAZ/CAC) based on the measurement of the diameter of inhibition. We were able to apply the CAZ/CAC method to 80 isolates (39 *K. pneumoniae*, 8 *K. oxytoca* and 31 *E. coli*) and we found that 37% of *K. pneumoniae*, 25% of *K. oxytoca* and 16% of *E. coli* were positive with the confirmatory method.

Considering the high prevalence of *K. pneumoniae* ESBL in HCPA a genetic analysis by DNA macrorestriction followed by PFGE was performed in 17 isolates. Two isolates were obtained from the same patient during an interval of four days but all the other isolates were obtained from different patients. Nine of these isolates were recovered within a period of 25 days. Persistent degradation of DNA occurred in one sample. We were able to identify a great variety of profiles as none of the ESBL *K. pneumoniae* displayed the same pattern of macrorestriction (Fig. 1) with the exception of the isolates obtained from the same patient (data not shown).

**DISCUSSION**

The percentages of ESBL production in our hospital was highly variable whether all methods were compared in an equal hierarchy. The data obtained using NCCLS breakpoints (KB method) provided rates of ESBL production much higher (p <0.005) than those obtained by other methods. However, one should consider that the KB method, as proposed by NCCLS (19), has to be use only as a screening method and therefore cannot be directly compared to the confirmatory methods. As a matter of fact, none aztreonam non-resistant and ceftriaxone

**Figure 1.** Macrorestriction analysis of DNA by pulsed-field gel electrophoresis of 15 isolates of ESBL *K. pneumoniae* (λ = molecular weight marker of 48.5 kb).
non-resistant isolates were ESBL-producers. This indicates that NCCLS breakpoints were not necessary to detect ESBL using these disks, as it gave only false-positive results in our population. In the case of the ceftazidime disks we could reduce de NCCLS breakpoints by 2 mm and we would have detected all ESBL-producers. We, therefore, agree with Lee et al. (11) that adjusting breakpoints to detect ESBL in different countries or regions is important to avoid false-positive results and unnecessary work. In view of that, we will consider only the methods of DD, MIC CAZ/CAC and MIC CAZ to indicate the real rates of ESBL production in this study. The CAZ/CAC method also showed similar rates for ESBL although it was not used for all isolates tested by the other three methods.

The prevalence of \textit{K. pneumoniae} around the world varies greatly although it tends to be less than 25% in many countries (1,3,13,16,24). We, therefore, considered that the prevalence of ESBL was quite high among isolates of \textit{K. pneumoniae} in this study.

The difference, although not statistically significant, in range among the confirmatory methods could be attributed mainly to the fact that the methods are based in different characteristics of the enzyme. Thus, while the DD and MIC CAZ/CAC methods evaluate inhibition of the \(\beta\)-lactamase enzyme by clavulanic acid, the method of MIC CAZ take in account the raise of resistance. The latter determination is less specific for ESBL production since it may detect resistance due to other mechanisms such as AmpC production (1). Data obtained in this study for \textit{Klebsiella} spp corroborated that MIC CAZ is less specific for ESBL production. It has also to be considered that in some occasions the inhibitory effect of clavulanic acid was not clearly visualized in the DD method (5). Therefore, the interpretation of results in this method was somehow empiric and may depend on individual basis as there is no standard interpretative criteria which may difficult the use of this method in the routine laboratories. Another problem of this method, the distance between disks (1), can be illustrated by isolate number 15 which would be considered negative for ESBL production using DD method with disks 1.5 cm apart. This isolate was re-tested changing only the distance between disks (reduced to 1.0 cm) and this resulted in a clear inhibitory effect of the clavulanic acid which was barely seen before (Fig. 2).

To estimate the existence of an outbreak of ESBL \textit{K. pneumoniae} we performed macrorestriction analysis of DNA by pulsed-field gel electrophoresis. The data obtained indicated that the isolates had no clonal relation as each patient was harboring a distinct strain. Thus, the high prevalence of ESBL \textit{Klebsiella} in our hospital is probably consequence of individual selection of resistant strains rather than due to the transmission of a common strain between patients. This diversity in genotype was also described in Taiwan by Liu et al. (30) who suggested that the increase of ESBL \textit{K. pneumoniae} in their hospital was mainly due to dissemination, or mutation, in plasmids containing resistant genes. Conversely, in other centers the increase of ESBL \textit{K. pneumoniae} was correlated with a single, or few, clones (17,30).

The high levels of ESBL, mainly among isolates of \textit{K. pneumoniae}, in our hospital is worrying and warrants special attention by both the clinician and the microbiology laboratory. While the former has to re-evaluate the antibiotic policies, the laboratory must to be capable to readily identify these isolates.

RESUMO

\(\beta\)-lactamases de espectro ampliado em \textit{Klebsiella} spp e em \textit{Escherichia coli} obtidas em um hospital escola brasileiro: detecção, prevalência e tipagem molecular

Este estudo foi desenvolvido para comparar métodos de detecção e para estimar a prevalência de \textit{Klebsiella} spp e \textit{E.coli} produtoras de \(\beta\)-lactamases de espectro ampliado (ESBL) em um Hospital Universitário no sul do Brasil. A correlação genética, determinada através de método molecular de tipagem, entre as amostras de \textit{K. pneumoniae} também foi determinada. A produção de ESBL foi investigada em 95 amostras de \textit{Klebsiella} spp e \textit{E.coli} obtidas de pacientes no Hospital de Clínicas de Porto Alegre usando-se: medida do diâmetro a zona de inibição (KB), dupla-difusão de disco (DD), valores de concentração inibitória mínima da ceftazidima (MIC CAZ), aumento do diâmetro da zona de inibição com adição de clavulanato (CAZ/CAC) e a relação entre o MIC da ceftazidima/MIC ceftazidima com clavulanato (MIC CAZ/CAC). A tipagem molecular foi realizada utilizando-se o método de macrorestricção de DNA e eletroforese em campo pulsado (PFGE). O método KB apresentou as maiores taxas de produção de ESBL (> 70% para \textit{Klebsiella} e 59% para \textit{E.coli}) contrastando com os outros métodos (p< 0,05). Os métodos confirmatórios (DD, MIC CAZ e MIC CAZ/CAC) indicaram a produção de ESBL em 8 a 13% de \textit{E.coli} e em 33 a 40% para as espécies de \textit{Klebsiella}. Portanto, o método KB é útil apenas como método de triagem devido aos diversos resultados considerados falso-positivos. A
tipagem molecular realizada em 17 amostras de *K. pneumoniae* ESBL indicou não existência de relação clonal. Este estudo encontrou uma boa correlação entre os métodos confirmatórios de detecção de ESBL embora os métodos que avaliam a inibição da enzima pelo clavulanato pareçam ser mais específicos. A alta prevalência de *Klebsiella* ESBL em nosso hospital provavelmente se deve a seleção individual de cepas resistentes do que a transmissão de uma cepa comum.

**Palavras-Chave:** ESBL, Klebsiella, resistência, tipagem molecular.

**REFERENCES**