CHARACTERIZATION OF A NOVEL LIPASE FROM \textit{Bacillus} SP. ISOLATED FROM TANNERY WASTES

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ABSTRACT

Kinetics of a lipase isolated from \textit{Bacillus} sp. was studied. The enzyme showed maximum activity at pH 9 and temperature 60°C. The Michaelis constant (\(K_M\) 0.31 mM) obtained from three different plots i.e., Lineweaver-Burk, Hanes-Wolf and Hofstee, was found to be lower than already reported lipases that confirmed higher affinity of the enzyme for its substrate p-NPL (p-nitrophenyl laurate). \(V_{\text{max}}\) of the enzyme was found to be 7.6 \(\mu\)M/mL/min. Energy of activation calculated from Arrhenius plot was found to be 20.607 kJmol\(^{-1}\). Activation enthalpy (\(\Delta H^*\)) had negative trend and the value for the hydrolysis of p-NPL by the enzyme at optimum temperature was -2.748 kJmol\(^{-1}\). Activation entropy (\(\Delta S^*\)) and free energy of activation (\(\Delta G^*\)) of the enzyme were found to be 1.468 Jmol\(^{-1}\)K\(^{-1}\) and -3.237 kJmol\(^{-1}\), respectively at optimum temperature. Low value of \(Q_{10}\) (0.04788) shows high catalytic activity of the enzyme. Mn\(^{2+}\), Fe\(^{2+}\) and Mg\(^{2+}\) enhanced the lipase activity whereas Cu\(^{2+}\), Na\(^+\) and Co\(^{2+}\) inhibited the enzyme activity. However, the enzyme activity was not affected significantly by K\(^+\) ions. EDTA and SDS also significantly inhibited the lipase activity. Activity of the enzyme was increased in n-hexane while decreased with increase in concentration of acetone, chloroform, ethanol and isopropanol.

Key words: \textit{Bacillus} sp., Kinetic study, Lipases, p-Nitrophenyl laurate, Tannery wastes, Organic solvents

INTRODUCTION

Lipases (triacylglycerol hydrolases, EC 3.1.1.3) are hydrolases acting on the carboxylic ester bonds present in acylglycerols to liberate fatty acids and glycerol. These are among the most important industrial enzymes in terms of their versatility (1, 2). Thermostable and alkalophilic lipases have great potential to be used in detergent, food flavoring, leather processing, pharmaceutical, cosmetics, etc (3). Lipases remain enzymatically active in organic solvents (4, 5) that enhances their potential and flexibility as biocatalysts against a wide range of unnatural hosts (6). These enzymes are the most widely used biocatalysts in organic chemistry (1), thus find tremendous applications in organic synthesis (7, 8). The regio- and/or enantioselectivity of lipases makes them highly attractive source to work precisely in various esterifications,

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alcoholysis, aminolysis or transformation reactions (9). The enzyme catalyzed reactions in organic solvents have advantages over the reactions carried out in aqueous medium due to many properties including “molecular memory” (10), therefore, lipase-catalyzed ester hydrolysis in water is converted into ester synthesis in non-aqueous media (1).

Alkalophilic microorganisms are widely distributed in nature and the alkalophilic Bacillus strains are often good sources of alkaline extracellular enzymes (11, 12). Microbial lipases are usually extracellular enzymes, which are produced by various fungi, actinomycetes and bacteria (13, 14). Lipases are of significant importance in leather industry. Degreasing of leather during its processing is an important use of lipases in tanning industry in the process of bating. Removal of fat and protein debris by chemical processes is both polluting and laborious (15). Lipases can play distinct role in resolving such problems of leather industry and tanneries. Therefore, researchers are always in search of novel lipases with high catalytic rates from microbial sources. We isolated a Bacillus sp. strain FH5 from tannery wastes that was found to be a good producer of lipases (16). Two lipases were purified (17), however only one with Mr 62 kDa was characterized and reported. In this paper we report kinetic and thermodynamic characteristics of the other lipase (24 kDa) produced from this strain.

**MATERIALS AND METHODS**

**Microorganism and enzyme production**

Bacillus sp. FH5 isolated from tannery waste was grown in shake flask cultures in selective medium for the production of lipase (16). The lipases produced under optimum conditions were purified by acetone precipitation followed by chromatographic procedures i.e. gel filtration on Sephadex G-75 followed by ion-exchange chromatography on DEAE-cellulose (17). The lipase with Mr 24 kDa was subjected to further characterization.

**Lipase Assay**

The method of (18) was used for the determination of lipase activity. To 20 μL of lipase solution was added 880 μL buffer (0.1 M KH₂PO₄; 0.1% gum Arabic, 0.2% deoxycholate, pH 8.0). After three minutes of incubation at 37°C, 100 μL of 8 mM substrate (p-nitrophenyl laurate; pNPL) solubilized in isopropanol was added and re-incubated for 5 minutes at room temperature. The reaction was stopped by adding 0.5 mL of 3 M HCl and centrifuged at 10000xg for 10 minutes at room temperature. The 333 μL supernatant was mixed with 1 mL of 2 M NaOH and absorbance was recorded at 420 nm. One unit of enzyme activity was defined as the amount of enzyme that released 1 μmol of p-nitrophenol from pNPL in one minute.

**Effect of temperature on lipase activity**

The enzyme assay was performed as discussed above except that incubation was done at temperatures from 10-70°C in increments of 10°C.

**Effect of pH**

Optimum pH for lipase activity was determined covering the range (4-11) using buffers of different pH. The buffers were: pH 4 -6 (acetate); pH 7 (phosphate); pH 8 (Tris-HCl) and pH 9-11 (Glycine-NaOH).

**Effect of substrate concentration**

Lipase was assayed in reaction buffer (pH 8) with different concentrations (0.113-1.2 mM) of pNPL (p-nitrophenyl laurate) as a substrate. The values of $V_{max}$ (maximum velocity) and $K_m$ (Michaelis constant) were calculated from Lineweaver-Burk, Hofstee and Hanes-Woof plots.

**Activation energy and Thermodynamic parameters**

Activation energy (Ea) was determined from Arrhenius plot (19). The values for the activation enthalpy (ΔH°), free energy of activation (ΔG°) and the activation entropy (ΔS°)
were calculated according the following equations:

\[ \Delta H^* = E_a - RT \]
\[ \Delta G^* = -RT \ln K_c \]
\[ \Delta S^* = (\Delta H^* - \Delta G^*)/T \]

**Increase in reaction rate per 10 K rise in temperature (Q<sub>10</sub>)**

The value of activation every was also used to calculate the increase in reaction rate, Q<sub>10</sub>, for every 10 K increase in temperature i.e. from T<sub>1</sub> to T<sub>2</sub>, according to the following formula:

\[ \log Q_{10} = E_a/R \left[ 1/T_1 - 1/T_2 \right] \]

**Effect of metal ions and inhibitors**

The effect of metal ions viz Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Fe<sup>2+</sup>, Cu<sup>2+</sup>, Co<sup>2+</sup> and potential inhibitors EDTA (Ethylenediamine tetracetic acid) and SDS (sodium dodecyl sulphate) was studied by incubation of the enzyme in the presence of 1 mM metal ions or the inhibitor following the method of (19). The enzyme activities were determined by normal assay procedure as discussed above.

**Effect of organic solvents on lipase activity**

The enzyme was assayed in the presence of different percentages (10-100%) of organic solvents viz acetone, chloroform, ethanol, n-hexane and isopropanol using the assay procedure under the standard conditions as discussed above.

**RESULTS AND DISCUSSION**

Extracellular lipases were produced from a local isolate of *Bacillus* sp. and purified with the help of acetone precipitation and different chromatographic steps as reporter earlier (17). Two lipases with Mr 64 and 24 kDa were obtained, and the lipase with 64 kDa was characterized and discussed. The lipase with Mr 24 kDa was also characterized and discussed below.

**Effect of temperature on lipase activity**

The temperature activity profile of *Bacillus* sp. lipase is shown in Figure 1. The lipase had an optimum temperature of 60°C that is comparable to the previously reported optimum temperatures for lipases from two different strains of *Bacillus* sp. (20-22). The lipase from *Bacillus* sp. reported by (22) lost 50% activity after incubation for 65 minutes at 70°C. A lipase from *Bacillus coagulans* BTS-3 showed maximum activity at 55°C and was found stable up to 70°C (23). Our lipase was found resistant at high temperature and exhibited 50% activity at 70°C and might be highly useful for industrial use. It was superior to that from *Bacillus subtilis*-168 having 35°C as its optimum temperature and losing all its activity at 55°C (18). The optimum temperature for a lipase from *Bacillus subtilis* IFFI 10210 (24) was also lower (45°C) as compared to our lipase.

![Figure 1](image.png)

**Effect of pH on lipase activity**

The enzyme activity increased with an initial increase in pH and optimum activity was noted at pH 9 suggesting alkaline nature of the enzyme. Further increase in pH beyond optimum caused a rapid decrease in the enzyme activity (Figure 2). A pH optimum from 8-9 has previously been reported for lipases from some *Bacillus* species (22-25). However, lipase from *Bacillus* sp. is reported to (21) exhibit maximum activity at pH 5.6.
**Effect of substrate concentration**

The $K_M$ and $V_{\text{max}}$ for the lipase were determined using pNPL as a substrate. The $K_M$ value for the free enzyme, estimated from Lineweaver-Burk plot (Figure 3), Woolf-Augustsson-Hofstee plot (Figure 4) and Hanes-Woolf plot (Figure 5) were 0.345, 0.361 and 0.399 mM, respectively with pNPL as substrate. The $V_{\text{max}}$ values obtained from the three plots were 7.61, 7.64 and 7.7 µM/mL/min., respectively. The $K_M$ and $V_{\text{max}}$ for a 64 kDa lipase from the same strain were found to be 5.05 mM and 0.416 µM/mL/min., respectively. The $K_M$ and $V_{\text{max}}$ for a lipase from *Bacillus* sp. J33 were found to be 2.5 mM and 0.4 µM/mL/min., respectively, with pNPL as substrate (26). Similarly, a lipase from another *Bacillus* sp. had $K_M$ and $V_{\text{max}}$ 3.63 mM and 0.26 µM/mL/min., respectively. Whereas, a lipase from another *Bacillus* sp. exhibited $K_M$ and $V_{\text{max}}$ of 0.5 mM and 0.319 µM/mL/min. on the same substrate (22). Our lipase therefore, had better affinity for the substrate and better catalytic activity as compared to previously reported lipases.
Lipase from *Bacillus* sp.

Enthalpy of activation ($\Delta H^*$) was found to be negative and decreased with increase in incubation temperature (Table 1). This indicates alteration in the conformation of the enzyme (27). Negative value of $\Delta G^*$ reflects that the enzyme reaction was spontaneously favorable in forward direction, and the maximum hydrolysis of the substrate was taking place under the conditions tested. Entropy ($\Delta S^*$) of the enzyme was found positive with slight decrease in the temperature range of 10 and 80°C. The positive values of $\Delta S^*$ thermodynamically support the reaction. It further demonstrates an increase in disorderliness that might be observed due to unfolding of the enzyme structure with increase in temperature.

**Table 1.** Thermodynamic parameters of lipase isolated from *Bacillus* sp.

<table>
<thead>
<tr>
<th>Temp. (K)</th>
<th>$\Delta G^*$ (kJmol$^{-1}$)</th>
<th>$\Delta H^*$ (kJmol$^{-1}$)</th>
<th>$\Delta S^*$ (Jmol$^{-1}$K$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>283</td>
<td>-2.751</td>
<td>-2.332</td>
<td>1.479</td>
</tr>
<tr>
<td>293</td>
<td>-2.848</td>
<td>-2.415</td>
<td>1.476</td>
</tr>
<tr>
<td>303</td>
<td>-2.945</td>
<td>-2.499</td>
<td>1.474</td>
</tr>
<tr>
<td>313</td>
<td>-3.042</td>
<td>-2.582</td>
<td>1.472</td>
</tr>
<tr>
<td>323</td>
<td>-3.140</td>
<td>-2.665</td>
<td>1.470</td>
</tr>
<tr>
<td>333</td>
<td>-3.237</td>
<td>-2.748</td>
<td>1.468</td>
</tr>
</tbody>
</table>

Increase in reaction rate per 10°C rise in temperature ($Q_{10}$)

Increase in reaction rate for energy 10°C convert to K rise in temperature was calculated for the lipase of *Bacillus* sp. The $Q_{10}$ value obtained for the lipase was 0.04788. This value shows that there was on average, ~5% increase in the reaction rates of this enzyme when the temperature was increased from 20°C to 30°C convert to K. Lower $Q_{10}$ values demonstrate high catalysis. A distinctive feature of enzyme catalysis is that the $Q_{10}$ of a catalyzed reaction is lower as compared to the same reaction uncatalyzed.

**Figure 5.** Hanes-Woolf plot for lipase from *Bacillus* sp. Lipase assay was conducted at various substrate concentrations at pH 9.0 and temperature 60°C. The data were plotted according to Hanes-Woolf equation. Each value is average of three independent experiments.

**Figure 6.** Arrhenius plot for the determination of energy of activation of lipase produced from *Bacillus* sp.
Effect of metal ions and inhibitors

The effect of metal ions viz Na⁺, K⁺, Mn²⁺, Fe²⁺, Mg²⁺, Cu²⁺, Co²⁺ and two potential inhibitors EDTA and SDS was studied on the lipase isolated from Bacillus sp. The results are summarized in Figure 7. Mn²⁺, Fe²⁺ and Mg²⁺ enhanced the lipolytic activity indicating that these ions do not compete with the enzyme whereas Cu²⁺, Na⁺ and Co³⁺ inhibited the enzyme activity showing competitive inhibition rendering enzyme to a reduced catalytic activity. K⁺, on the other hand, had no significant effect on the enzyme activity. Mg²⁺ has been reported to increase the lipase activity from Bacillus sp. (23, 24). However, in contrast to our findings, Mn²⁺ is reported to have inhibited activity of the lipase isolated from Bacillus sp. (18, 23). Sugihara and Tominaga (21) found that a lipase from Bacillus sp. was inhibited by the addition of Cu²⁺, whereas other metal ions including Mg²⁺ and Fe²⁺ had no effect on the enzyme activity. According to (24) Fe²⁺, Cu²⁺ and Co²⁺ inhibited the lipolytic activity from Bacillus subtilis. EDTA and SDS has been reported to completely inhibit the lipolytic activity. The 64 kDa lipase reported earlier by our lab (17) was also inhibited by both of the inhibitors, although the inhibition was not as strong as for our lipase. SDS has been reported to completely inhibit the activity of the lipase isolated from Bacillus thermoleovorans ID-1 (28). However, contradictory reports have been reported for the effect of EDTA. In a few cases EDTA has stimulatory or no affect on lipase activity (29, 30), whereas in others it shows inhibitory effect (31). Strong inhibition of the lipase activity in our case shows that the enzyme requires metal ions that are chelated out with EDTA.

Effect of organic solvents on the enzyme activity

Utilization of enzymes, especially lipases, in organic solvents is gaining much industrial importance as the process leads to the development of products of high-added value (4, 32). We tested activity of the lipase in five different solvents i.e. acetone, chloroform, ethanol, n-hexane and isopropanol (Figure 8). The enzyme exhibited high activity in all the organic solvents except for chloroform. Maximum lipolytic residual activity was observed in n-hexane followed by isopropanol, acetone and ethanol. Except for n-hexane, the enzyme activity was decreased with increase in the solvent concentration. The lipolytic activity was enhanced at higher concentrations of n-hexane showing stimulatory effect of the solvent on the enzyme. Other solvents did not stimulate the lipolytic activity, however, high lipolytic activity was observed at lower concentrations of the organic solvents. Contradictory reports are found in literature regarding the effect of the organic solvents on lipase activity. A lipase from Bacillus sp. was stimulated in the presence of acetone while inhibited in n-hexane (21). On the other hand activity of the lipase from Bacillus sp. strain 42 was found to be enhanced in n-hexane (33). Lipase from Rhizopus oryzae was found stable for many days in different organic solvents, n-hexane being the most suitable (34).

Conclusively, 24 kDa lipase produced and isolated from the local isolate of Bacillus sp. exhibited favorable kinetics for its use in industrial and environmental applications.

Figure 7. Effect of metal ions and potential inhibitors on lipase activity. The metal ion or inhibitor was added at 1 mM concentration and the lipase activity was assayed under the standard conditions. The percent lipase activity for control was taken as 100%.
Figure 8. Effect of organic solvents on the activity of the lipase. The assay was carried out in the organic solvents at various concentrations under standard conditions.

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REFERENCES


