The effect of methotrexate and azathioprine on the serum levels of IgA-\(\alpha_1\)-antitrypsin complex in juvenile chronic arthritis

J.K. Lacki\(^1,2\), K. Klama\(^1,2\), H. Michels\(^3\), H. Truckenbrodt\(^3\), S. Mackiewicz\(^2\) and W. Muller\(^1\)

1Hochrhein-Institute for Rheumatism Research and Prevention, 79713 Bad Säckingen, Germany
2Department of Rheumatology, Karol Marcinkowski University School of Medical Sciences, 61-626 Poznan, Poland
3Pediatric Rheumatology Center, 82467 Garmisch-Partenkirchen, Germany

Abstract

In the present study we investigated the influence of methotrexate (MTX) and azathioprine (AZA) on the serum levels of the IgA-\(\alpha_1\)-antitrypsin (IgA-AT) complex in patients with the systemic form of juvenile chronic arthritis (JCA). Fifty-six JCA patients (22 treated with MTX, 18 treated with AZA, and 16 not treated with any immunosuppressive agent) were enrolled in the study. MTX dosage ranged from 0.3 to 0.5 mg kg\(^{-1}\) week\(^{-1}\), while AZA was given daily at an average dose of 1 mg/kg. MTX was given for 13 months (SD = 7 months) whereas AZA for 11 months (SD = 6 months). The average value of the complex was higher in JCA patients than in both control groups (0.74 ± 0.73 U vs 0.37 ± 0.13 U (control children), P<0.001 and vs 0.23 ± 0.12 U (control adults), P<0.001). Values exceeding the normal range were found in twenty-two JCA patients (39.4%). Serum IgA-AT level was lowest in the MTX group compared to AZA and non-treated patients (0.56 ± 0.24 U, 0.76 ± 0.43 U, 0.95 ± 0.52 U, respectively, P<0.05). IgA values exceeding normal levels for age were found in 14% of the patients. A correlation between the levels of the IgA-AT complex and C-reactive protein (r = 0.43, P<0.01), \(\alpha_1\)-acid-glycoprotein (r = 0.45, P<0.01), \(\alpha_1\)-antichymotrypsin (r = 0.52, P<0.01), \(\alpha_1\)-antitrypsin (r = 0.40, P<0.01) and IgA (r = 0.56, P<0.01) was established.

Correspondence

J.K. Lacki
Department of Rheumatology
Karol Marcinkowski University School of Medical Sciences
ul. Winogrady 144
61-626 Poznan
Poland

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Key words
• Juvenile chronic arthritis
• IgA-\(\alpha_1\)-antitrypsin complex
• Methotrexate
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• C-reactive protein
• \(\alpha_1\)-Acid-glycoprotein
• \(\alpha_1\)-Antichymotrypsin

Introduction

Juvenile chronic arthritis (JCA), the most frequent connective tissue disease in children, is actually a collection of conditions that together constitute the major forms of chronic arthritis in childhood. The heterogeneous patterns of symptoms and clinical features make the uniform assessment of disease activity difficult. Fever, lymphadenopathy and rheumatoid rash are typical for the systemic onset of JCA. This form of JCA is usually accompanied by visceral involvement such as hepatosplenomegaly, pericarditis or other evidence of serositis. Arthritis can range from mild joint involvement to a chronic, erosive course (1). There is no single clinical or laboratory parameter that reflects
the inflammatory activity in JCA.

Increased levels of immunoglobulin A-\(\alpha_1\) -antitrypsin (IgA-AT) complex have been found in the serum of patients with rheumatoid arthritis (2-6), ankylosing spondylitis (7,8), and lupus erythematosus (9). In prospective studies of early rheumatoid arthritis the complex level at the onset of the disease was significantly higher in the group of patients who developed erosions (2,5). IgA-AT is a nonimmune complex formed by a disulfide bridge between an active thiol group available on the cysteine residue of the alpha heavy chains of IgA and a cysteine at position 232 of the \(\alpha_1\)-antitrypsin single polypeptide chain (4). Formation of the complex was proved to depend on the level of IgA (3,4,8). The pathological implications of the complex formation are still unknown.

The utility of methotrexate (MTX) and azathioprine (AZA) in the treatment of juvenile chronic arthritis has been shown in numerous studies (10-12). Although its mode of action is unknown, clinical improvement during MTX therapy appears a few weeks after the beginning of treatment and the number of side effects is relatively low. The aim of the present study was to estimate the influence of MTX and AZA on the levels of the IgA-AT complex in the systemic and polyarticular form of JCA and the relationship between the concentrations of the complex and its constituent components and other laboratory markers of inflammation for the disease.

Material and Methods

Patients

Fifty-six patients with the systemic and polyarticular form of JCA (25 females and 31 males, mean age 123 ± 69 months) were studied and their diagnosis was confirmed after at least 6 months of observation. All patients fulfilled the criteria of JCA (1). On the average, disease duration was 76 ± 62 months. Blood samples were obtained at the time of clinical examination. Serum samples were stored at less than -80°C. Twenty-two patients were treated with MTX, 18 with AZA, and 16 were not treated with any immunosuppressive drug. The clinical data are summarized in Table 1. Patients were randomly assigned to one of two immunosuppressive procedures. MTX dosage ranged from 0.3 to 0.5 mg kg\(^{-1}\) week\(^{-1}\), while AZA was given daily at an average dose of 1 mg/kg. MTX was given for 13 months (SD = 7 months) and AZA for 11 months (SD = 6 months). Sera from 16 children (9 females and 7 males, mean age 114 ± 54 months) with congenital malformations served as control. A second control group comprised 22 healthy adult volunteers (7 females and 15 males, mean age 25.6 ± 4.5 years). In all cases systemic connective tissue diseases were excluded.

Immunological methods

The IgA-\(\alpha_1\)-antitrypsin complex was evaluated by a double antibody enzyme immunoassay (EIA) on microtiter plates coated with monoclonal antibody against the IgA-AT complex. Briefly, sera diluted 1:100 were incubated at 37°C for 90 min, the plates were then emptied and washed 5 times. Bound IgA-AT complex was located by the addition of sheep anti-human IgA antibody and the second antibody, donkey anti-sheep IgG conjugated with peroxidase, was added. OPD peroxidase substrate tablets (Sigma Chemical Co., St. Louis, MO) served as substrate.
The plates were read with a spectrophotometer at 494 nm. The results were calculated using a calibration curve based on six concentrations of the reference complex and are reported as arbitrary units (U). Values less than 0.6 U were considered to be normal.

Kits were provided by Peptide Therapeutics Limited (Birmingham, England). C-reactive protein (CRP) was estimated by nephelometry and CRP values less than 5 mg/l were considered to be normal. α₁-Antitrypsin (AT), α₁-acid-glycoprotein (AGP), and α₁-antichymotrypsin (ACT) levels were measured by immunoelectrophoresis according to Laurell (13). The following range values were considered to be normal for adults: 2-4 g/l for AT, 0.45-1.4 g/l for AGP and 0.30-0.60 g/l for ACT (14). Children tend to have 80% of the adult values of AT, and 90% of AGP and ACT. IgA levels were estimated by radial immunodiffusion according to Mancini et al. (15) on NOR-Partigen IgA plates (Behring, Marburg, Germany).

Statistical analysis

Methods of descriptive statistics were used. Relationships between continuous variables were examined using the Spearman rank correlation coefficient. The chi-square test was used to evaluate the relationship between the complex, erythrocyte sedimentation rate (ESR), CRP and clinical manifestations of the disease such as fever and hepatosplenomegaly. The Mann-Whitney U-test was applied to determine differences between variables for the groups studied.

Results

The average value of the complex was higher in JCA patients than in both control groups (0.74 ± 0.73 U vs control children: 0.37 ± 0.13 U, P<0.001 and vs control adults: 0.23 ± 0.12 U, P<0.001). Values exceeding the normal range were found in 22 JCA patients (39.4%). We observed the lowest values of the IgA-AT complex in patients treated with MTX (Figure 1). The increased levels of the complex were correlated with the occurrence of fever (chi-square value with Yates correction = 11.31, P<0.001) and the number of affected joints (r = 0.35, P<0.01) but not with hepatosplenomegaly. Data for other systemic symptoms were not analyzed statistically because of the small number of patients. The levels of all acute phase proteins (APP) were significantly increased in the JCA patients compared to the controls (Table 2). IgA levels exceeding normal values were found in 8 patients (14.3%), who had increased serum levels of IgA-AT.

The level of the complex was correlated with the levels of its constituent components: IgA, r = 0.56, P<0.01, and AT, r = 0.40, P<0.01 in JCA patients. There was no correlation between IgA and AT in both control groups. Correlations between IgA-AT complex levels and the acute phase proteins CRP (r = 0.43, P<0.01), AGP (r = 0.45, P<0.01), and ACT (r = 0.52, P<0.01) in JCA patients were established.

Discussion

No studies on the IgA-AT complex in
JCA have been published previously. The present results show that patients with JCA display higher levels of IgA-AT complex than control children and control adults. These higher than normal values were found in 39% of JCA patients and most of them demonstrated an active form of the disease with high APP levels. Elevated levels of the complex have been detected in sera from rheumatoid arthritis patients at frequencies of 14 to 60% depending on the population studied (2,4). Increased levels of the complex were also found in 34-50% of patients with ankylosing spondylitis (7,8), and were correlated with “extraspinal” manifestations of the disease such as synovitis, anterior uveitis and increased values of ESR, CRP and IgA (8).

Changes in APP levels are of great value in estimating the intensity of inflammation in JCA (16,17) and rheumatoid arthritis (14,17,18). APP constitute a group of proteins synthesized mainly by the liver whose concentration increases or decreases in response to inflammation or injury (14,19). The correlation between complex level and clinical manifestation of the disease, such as fever and joint involvement, and the acute phase proteins suggests that the IgA-AT complex may be a part of the response to inflammatory stimuli in JCA patients. On the other hand, we found no correlation between the level of the complex and ESR. However, some findings suggest that ESR does not closely reflect the changes in disease activity in children with JCA (20). Due to the slow dynamics of fibrinogen synthesis and its long half-life, ESR increases slowly and persists longer and it is also affected by the immunoglobulin concentration which is relatively low in children.

We noticed that MTX-treated patients had significantly lower levels of IgA-AT complex compared to control groups, suggesting a relation with disease activity or a direct destructive action of MTX on the complex. There is evidence that some disease-modifying antirheumatic drugs such as gold salts, D-penicillamine, or sulfasalazine may destroy covalent linkage between IgA and

Table 2 - Levels of the IgA-AT complex, acute phase response markers and other laboratory parameters of juvenile chronic arthritis (JCA) patients undergoing immunosuppressive therapy.

<table>
<thead>
<tr>
<th></th>
<th>Control adults</th>
<th>Control children</th>
<th>JCA patients</th>
<th>MTX-treated JCA patients</th>
<th>AZA-treated JCA patients</th>
<th>Non-treated JCA patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>N = 22</td>
<td>N = 16</td>
<td>N = 56</td>
<td>N = 22</td>
<td>N = 22</td>
<td>N = 18</td>
<td>N = 16</td>
</tr>
<tr>
<td>ESR (mm/h)</td>
<td>6 (2-10)</td>
<td>8 (4-12)</td>
<td>31 (2.98)**</td>
<td>28 (2-66)</td>
<td>30 (3-98)</td>
<td>35 (2-72)</td>
</tr>
<tr>
<td>CRP (mg/l)</td>
<td>3 (0-5)</td>
<td>9 (1-17)</td>
<td>59 (3-175)**</td>
<td>52 (3-112)</td>
<td>60 (4-175)</td>
<td>67 (4-135)</td>
</tr>
<tr>
<td>AGP (mg/l)</td>
<td>0.47 (0.25-0.76)</td>
<td>0.89 (0.55-1.58)</td>
<td>1.62 (0.55-3.38)*</td>
<td>1.60 (0.89-3.05)</td>
<td>1.58 (0.55-3.38)</td>
<td>1.70 (0.67-3.25)</td>
</tr>
<tr>
<td>ACT (mg/l)</td>
<td>0.49 (0.29-0.65)</td>
<td>0.37 (0.28-0.53)</td>
<td>0.61 (0.22-1.39)*</td>
<td>0.60 (0.32-1.21)</td>
<td>0.62 (0.22-1.39)</td>
<td>0.63 (0.29-1.10)</td>
</tr>
<tr>
<td>AT (mg/l)</td>
<td>2.9 (2.1-3.9)</td>
<td>2.6 (1.8-3.7)</td>
<td>3.7 (2.5-7.8)*</td>
<td>3.6 (2.4-5.0)</td>
<td>3.8 (2.5-7.8)</td>
<td>3.7 (2.6-6.0)</td>
</tr>
<tr>
<td>IgA (g/l)</td>
<td>NA</td>
<td>1.48 (0.4-4.72)</td>
<td>1.77 (0.2-4.8)</td>
<td>1.50 (0.2-3.65)*</td>
<td>1.72 (0.72-4.8)</td>
<td>2.20 (0.49-4.74)</td>
</tr>
<tr>
<td>IgA-AT (U)</td>
<td>0.23 (0.12-0.48)</td>
<td>0.37 (0.17-0.57)</td>
<td>0.74 (0.26-2.73)*</td>
<td>0.56 (0.27-1.39)*</td>
<td>0.76 (0.26-2.36)</td>
<td>0.95 (0.29-2.73)</td>
</tr>
<tr>
<td>Hemoglobin (g/l)</td>
<td>13.4 (11.6-15.9)</td>
<td>11.8 (3.7-18.2)</td>
<td>11.9 (7.2-18.2)</td>
<td>12.6 (8.5-16.2)</td>
<td>10.9 (3.7-14.8)</td>
<td>12.1 (3.8-32.5)</td>
</tr>
<tr>
<td>Leucocyte (g/l)</td>
<td>7.8 (3.6-16.6)</td>
<td>10.8 (3.8-32.5)</td>
<td>11.8 (4.4-27.4)</td>
<td>8.4 (4.2-22.5)</td>
<td>12.1 (3.8-32.5)</td>
<td>NA</td>
</tr>
<tr>
<td>Platelets (g/l)</td>
<td>317 (146-415)</td>
<td>435 (178-998)</td>
<td>449 (178-998)</td>
<td>398 (192-742)</td>
<td>458 (236-766)</td>
<td>NA</td>
</tr>
</tbody>
</table>
antitrypsin (3,7,8). Our findings show that such an effect may be also elicited by MTX.

A recent study provided evidence that serum IgA is the component organizing the complex (21). The presence of high IgA levels in serum from patients with ankylosing spondylitis (7,8), rheumatoid arthritis (4), and JCA and its correlation with IgA-AT levels suggest that IgA and IgA-AT not only reflect disease activity but may contribute to the pathogenesis itself. The complex is probably produced within inflamed rheumatoid joints, since larger amounts of the IgA-AT complex are found in synovial fluid than in serum. The complex consumes large amounts of α1-antitrypsin which is a major antiprotease, and may elicit the release of lysosomal enzymes from macrophages by a process dependent on alternative complement pathway activation, thus destroying cartilage and bones and causing erosions. Accordingly, previous reports suggest that persistently increased complex levels might be a prognostic factor of erosion development in rheumatoid arthritis (3,6), but this hypothesis requires further study.

References