Alcohol dependence induced in rats by semivoluntary intermittent intake

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Abstract

The objective of the present experiment was to assess ethyl alcohol (ETOH) dependence brought about by a semivoluntary intermittent intake regimen in rats. Male Wistar rats weighing 150-250 g at the onset of the experiment were assigned to the following groups: 0% ETOH (N = 11), 5% ETOH (N = 20), 20% ETOH (N = 20) and 40% ETOH (N = 18). ETOH solutions were offered at the end of the day and overnight from Monday to Friday, and throughout weekends, for 90 days. The concentration of the ETOH solutions was increased in a stepwise fashion allowing the rats to get used to the taste of alcohol. Reposition of pure water was permitted during 1-h water drinking periods in the morning. Daily volume intake (± SEM) averaged 25.4 ± 0.4 ml (0% ETOH), 23.8 ± 0.6 ml (5% ETOH), 17.6 ± 0.7 ml (20% ETOH) and 17.5 ± 0.6 ml (40% ETOH). ETOH consumption differed significantly (P<0.05) among groups, averaging 4.4 ± 0.2 g kg⁻¹ day⁻¹ (5% ETOH), 10.3 ± 0.3 g kg⁻¹ day⁻¹ (20% ETOH) and 26 ± 1.2 g kg⁻¹ day⁻¹ (40% ETOH). Furthermore, ETOH detection in plasma 10-12 h after offering the solution indicated that its consumption in the 40% ETOH group was sufficient to override its metabolism. Overt signs of ETOH dependence, such as increased thirst, hyperactivity, puffing, hair ruffling and startle responsiveness as well as reduced drowsiness, were significantly increased in the 20% and 40% ETOH groups compared to the 0% and 5% groups. Accordingly, the model described here proved to be a useful tool for the evaluation of subtle or moderate behavioral and physical consequences of long-term ETOH intake.

Key words
- Ethanol
- Alcohol
- Drug dependence
- Rat

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Experimental dependence on ethyl alcohol (ETOH) is usually induced by 3 methods of drug administration: involuntary, semivoluntary or voluntary (1). While the involuntary method refers to the forced drug administration by gastric intubation or by the intravenous, intraperitoneal and pulmonary routes (2), in the semivoluntary method ETOH is offered as part of a liquid or food diet, which is ingested spontaneously (3), and in the voluntary method the animals have to learn to self-administer the drug (4). Both advantages and disadvantages have been observed for each procedure. Thus, whereas the involuntary method is quite suitable for the evaluation of the development of tolerance and early signs of physical dependence (1,5), the procedure is rather ag-
gressive and often produces severe consequences. On the other hand, while the spontaneous oral intake of ETOH greatly favors the semivoluntary method, dehydration, which is not a common feature in clinical practice, may supervene as a consequence of long-term ETOH administration in which the drug solution is the only source of liquid. Finally, although the voluntary method is quite useful for the assessment of the reinforcing properties of ETOH, its higher cost and technical difficulties are the major drawbacks of its use. The aim of the present study was to establish a simple model of induction of ETOH dependence by means of a semivoluntary intermittent daily intake of different concentrations of the drug.

Two-month old male Wistar rats weighing 150–250 g at the beginning of the experiment were placed in individual cages with food ad libitum. Rats were randomly assigned to four groups: 0% ETOH (N = 11), 5% ETOH (N = 20), 20% ETOH (N = 20) and 40% ETOH (N = 18), according to treatment, i.e., tap water for the 0% ETOH control group and 5%, 20% and 40% ETOH solutions for the remaining groups. Rats were weighed twice a week and the solutions were offered at the end of the day and kept overnight from Monday to Friday, and throughout the weekends. The concentration of the ETOH solutions was increased in a stepwise fashion allowing the rats to get used to the taste of alcohol. Weekly concentration increments to 5%, 10% and 20% were used in the first three weeks for the 5%, 20% and 40% ETOH groups, respectively. The concentration of the 40% ETOH group was further increased in steps to 30% and 40% in the fourth and fifth week, respectively. Once the desired ETOH concentration was attained, the solution was kept unchanged up to the completion of the 90-day treatment.

Daily reposition of pure water was permitted during 1-h water drinking periods in the morning. Immediately thereafter, the ingested volume of water or ETOH solution was measured and behavioral responses indicative of physical dependence were recorded as present or not.

Mean weekly frequencies of behavioral responses were calculated for the following behavioral items: thirst (strong approach behavior to the bottle and vigorous drinking), hyperactivity (sudden quick movements during bottle replacement), startle response (sudden jump induced by finger snapping or key shaking near the cage), puffing (rapid respiratory movements), drowsiness (resting posture with eyes closed or semi-closed and apparent muscle relaxation as suggested by the lowering of the trunk and/or head and flexion of the limbs), and hair ruffling (pilomerection).

Daily ETOH consumption was later confirmed by means of a biochemical assay in a separate set of experiments. Thus, rats were submitted to the same alcohol intake regimen, i.e., 0% ETOH (N = 5), 5% ETOH (N = 6), 20% ETOH (N = 6) and 40% ETOH (N = 6), but only for 30 days. At the end of treatment, rats were anesthetized with ethyl ether and two blood samples were drawn from the tail artery, 2–4 h (1st sample) and 10–12 h (2nd sample) after withdrawal of the ETOH solutions. Blood samples were then centrifuged and plasma ETOH levels were determined by enzymatic assay (Sigma Diagnostics Alcohol Reagent, St. Louis, MO) using a spectrophotometer (Gilford Instruments, model 250, Oberlin, OH).

Alcohol intake and biochemical data were analyzed by two-way analysis of variance for repeated measures followed by the post hoc Tukey test and by the Student t-test of least square means, respectively. Weight gain was assessed by regression analysis. Behavioral scores were evaluated by nonparametric Kruskal-Wallis ANOVA followed by a post hoc nonparametric Tukey-like test (6).
Data are reported as the mean ± SEM and differences were considered to be significant at P<0.05.

Rats weighed 192.2 ± 17.6 g (0% ETOH), 180.5 ± 14.8 g (5% ETOH), 218.2 ± 13.6 g (20% ETOH) and 181.1 ± 4.8 g (40% ETOH) at the onset of the experiment and 395.5 ± 14.1 g (0% ETOH), 328 ± 15.2 g (5% ETOH), 354.2 ± 11.4 g (20% ETOH) and 289.4 ± 6.8 g (40% ETOH) at the end of the experiment. There was a significantly smaller weight gain in the 5% and 40% ETOH rats compared to the other groups (P<0.003) (Figure 1C). Nonetheless, linear regression analysis showed a significant weight gain for all groups (P<0.001) throughout the experiment: 0% ETOH (Y = 208.4 + 12.3x, r = 0.95), 5% ETOH (Y = 187.8 + 11.0x, r = 0.98), 20% ETOH (Y = 229.2 + 10.0x, r = 0.96) and 40% ETOH (Y = 191.2 + 7.2x, r = 0.97).

Daily volume intake averaged 25.4 ± 0.4 ml (0% ETOH), 23.8 ± 0.6 ml (5% ETOH), 17.6 ± 0.7 ml (20% ETOH) and 17.5 ± 0.6 ml (40% ETOH) (Figure 1A). The 5%, 20% and 40% ETOH groups drank smaller amounts in the first week than the 0% ETOH group (P<0.05). Compared to control rats, the 5% ETOH group showed a lower volume intake during the first six weeks (P<0.05), but did not differ from controls thereafter. The 20% and 40% ETOH groups drank smaller volumes than the 0% and 5% ETOH groups from the second week onwards (P<0.05). Moreover, compared to the 0% and 5% ETOH groups, a further decrease in volume intake was observed in the 20% and 40% ETOH groups after the increase of the alcohol concentration in the third and fifth weeks (P<0.05) (Figure 1A). ETOH consumption also differed significantly (P<0.05) among groups, averaging 4.4 ± 0.2 g kg⁻¹ day⁻¹ (5% ETOH), 10.3 ± 0.3 g kg⁻¹ day⁻¹ (20% ETOH) and 26 ± 1.2 g kg⁻¹ day⁻¹ (40% ETOH). Thus, ETOH consumption increased proportionally to alcohol concentration (P<0.05) (Figure 1B).

Daily evaluation of behavioral responses during the alcohol withdrawal period showed dose-dependent increases in thirst, hyperactivity and startle behavior and a dose-dependent decrease in drowsiness (P<0.05). Overt autonomic signs, i.e., puffing and hair ruffling, also showed a dose-dependent increase (P<0.05) (Table 1).

Compared to the corresponding background levels of the 0% ETOH group, there was a significant increase in plasma ETOH 10-12 h after the 40% solution was offered (P<0.01). Indeed, slightly increased levels were already detected 2-4 h after the presentation of the 40% ETOH solution (P<0.12) and 10-12 h after the presentation of the 5%
ETOH solution (P<0.13). However, no changes were observed for the 20% ETOH group (Table 2).

Alcohol concentrations of 5%, 20% and 40% are close to those usually found in beer, wine and spirits, respectively. These solutions were offered in a regimen similar to that observed in either episodic or heavy human drinkers. Moreover, in spite of the daily offering of tap water, ETOH consumption was quite high, ranging from 4.4 up to 26 g kg\(^{-1}\) day\(^{-1}\), a consumption similar to that used in many studies employing the semi-voluntary method in which the ETOH solution was the only available source of liquid (7). Alcohol intake of 11.1 g kg\(^{-1}\) day\(^{-1}\) after a 12-week treatment with 20% ETOH solution (8), or 8 g kg\(^{-1}\) day\(^{-1}\) after an 8-week treatment with 15% ETOH solution (9), or even 11.6 g kg\(^{-1}\) day\(^{-1}\) with a mild 9% ETOH solution after a 4-week treatment only (10), has been reported. Thus, in spite of the natural aversion of rats to the taste of alcohol (1), the present data suggest that rats on an intermittent alcohol intake regimen are able to drink large amounts of the solution. Furthermore, biochemical plasma analyses showed that the ETOH intake exceeded the rate of alcohol metabolism. Indeed, although the sampling time may not reflect the actual daily pattern of ingestion, plasma ETOH levels significantly higher than those for controls were found 10-12 h after offering the 40% ETOH solution. Plasma ETOH levels, however, were lower than the levels reported 15 min after acute intraperitoneal administration of 0.5 g/kg and 1 g/kg of ETOH, which produced plasma concentrations of 50 mg/dl and 107 mg/dl, respectively (1,11, 12).

Signs of physiological dependence on alcohol are often inferred from behavioral changes brought about by removal of the alcoholic solution (7,13). Accordingly, hyperexcitability of the central nervous system, including audiogenic convulsions, trembling, spasticity, hyperactivity and increased responsiveness to environmental stimuli such as startle, irritability and hypervigilance, as well as autonomic responses such as tachypnea and tachycardia, were commonly described during abstinence of rats chronically treated with ETOH (7). Nevertheless, the virtual absence of physical dependence is often alleged as a major drawback of the oral intake of ETOH (1). Although severe responses which follow involuntary administration (1,7,14) were not observed in the present study, overt signs indicating drug dependence were demonstrated for all groups, including increased puffing and hair ruffling in the 5% ETOH group. Nevertheless, the absence of heavy signs of dependence favors oral ETOH intake as a reliable model for

### Table 1 - Mean week frequency of behavioral responses recorded 1 h after the removal of the alcohol solution.

<table>
<thead>
<tr>
<th>Behavioral responses</th>
<th>0% ETOH</th>
<th>5% ETOH</th>
<th>20% ETOH</th>
<th>40% ETOH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thirst</td>
<td>6.1 ± 1.0</td>
<td>21.1 ± 1.9</td>
<td>64.9 ± 4.4*</td>
<td>75.2 ± 7.9*</td>
</tr>
<tr>
<td>Hyperactivity</td>
<td>5.4 ± 1.8</td>
<td>11.3 ± 1.9</td>
<td>26.3 ± 3.7*</td>
<td>48.2 ± 4.6*</td>
</tr>
<tr>
<td>Startle</td>
<td>54.8 ± 2.3</td>
<td>62.1 ± 3.7</td>
<td>78.7 ± 2.0*</td>
<td>87.6 ± 1.7*</td>
</tr>
<tr>
<td>Puffing</td>
<td>4.4 ± 1.4</td>
<td>18.5 ± 3.0*</td>
<td>19.7 ± 2.9*</td>
<td>45.9 ± 6.7*</td>
</tr>
<tr>
<td>Hair ruffling</td>
<td>70.0 ± 2.9</td>
<td>88.4 ± 3.1*</td>
<td>94.4 ± 1.4*</td>
<td>95.5 ± 1.5*</td>
</tr>
<tr>
<td>Drowsiness</td>
<td>21.5 ± 3.2</td>
<td>16.7 ± 2.2</td>
<td>8.3 ± 1.5*</td>
<td>4.6 ± 1.7*</td>
</tr>
</tbody>
</table>

Data are reported as means x 100 ± SEM. *P<0.05 compared to 0% ETOH; +P<0.05 compared to 0% and 5% ETOH (nonparametric Tukey test).

### Table 2 - Plasma alcohol levels of tail artery samples withdrawn 2-4 h and 10-12 h after offering the alcohol solution.

<table>
<thead>
<tr>
<th>Group</th>
<th>1st Sample</th>
<th>2nd Sample</th>
</tr>
</thead>
<tbody>
<tr>
<td>0% ETOH</td>
<td>2.2 ± 1.1</td>
<td>2.1 ± 1.0</td>
</tr>
<tr>
<td>5% ETOH</td>
<td>4.5 ± 1.8</td>
<td>7.4 ± 2.1</td>
</tr>
<tr>
<td>20% ETOH</td>
<td>5.5 ± 0.8</td>
<td>2.5 ± 1.7</td>
</tr>
<tr>
<td>40% ETOH</td>
<td>6.8 ± 3.3</td>
<td>11.1 ± 3.9*</td>
</tr>
</tbody>
</table>

Data are reported as means ± SEM (mg/dl). *P<0.01 compared to 0% ETOH (Student t-test of least square means).
Alcohol dependence in rats inducing mild or moderate alcohol dependence, a pattern most likely to be found in human drinkers. In conclusion, the present model proved to be a useful tool for the evaluation of subtle or moderate physical and behavioral consequences of long-term ETOH intake.

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References