Immune and hormonal activity in adults suffering from depression


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Abstract

An association between depression and altered immune and hormonal systems has been suggested by the results of many studies. In the present study we carried out immune and hormonal measurements in 40 non-medicated, ambulatory adult patients with depression determined by CID-10 criteria and compared with 34 healthy nondepressed subjects. The severity of the condition was determined with the Hamilton Depression Rating Scale. Of 40 depressed patients, 31 had very severe and 9 severe or moderate depression. 29 (72.5%) were females and 11 (27.5%) were males (2.6:1 ratio). The results revealed a significant reduction of albumin and elevation of α-1, α-2 and β-globulins, and soluble IL-2 receptor in patients with depression compared to the values obtained for nondepressed subjects (P<0.05). The decrease lymphocyte proliferation in response to a mitogen was significantly lower in severely or moderately depressed patients when compared to control (P<0.05). These data confirm the immunological disturbance of acute phase proteins and cellular immune response in patients with depression. Other results may be explained by a variety of interacting factors such as number of patients, age, sex, and the nature, severity and/or duration of depression. Thus, the data obtained should be interpreted with caution and the precise clinical relevance of these findings requires further investigation.

Key words
- Depression
- Immunity
- Psychoneuroimmunology
- Cytokines
- Acute phase proteins

Introduction

Many reports have considered a possible association between depressive disorders and immune and hormonal alterations, although the results obtained have not been consistent. The interactions between the central nervous system and the immune system in depression have a biological explanation supported by the monoamine hypothesis (1), dysregulation of hypothalamic-pituitary-adrenocortical activity (2), and the macrophage theory which proposes that an excessive secretion of macrophage monokines such as interleukin-1 (IL-1) may provoke the hormonal abnormalities linked to depression (3). The three hypotheses may be linked to one another (4).

A number of studies have shown that depressive symptoms are associated with an impairment of immune function and increased susceptibility to infections (5) and cancer (6).
and the progression of immunodeficiency virus (HIV) infection (7). At the cellular
level, some investigators have reported reduced mitogen-stimulated lymphocyte pro-
fileration (8-10) although others have not (11-13). An increase in total white cell num-
ber and abnormality in differential white blood cell count with increased percentage
of neutrophils and decreased lymphocytes have also been reported (4). However, most
of the cell enumeration studies on depressed patients have yielded an unchanged number
of white blood cells, neutrophils, lymphocytes, total T-cells or T cell subsets (14).
Impairment of natural killer cell activity has been reported by several investigators (7,15).

At the subcellular level, it has been re-
ported that serum and plasma concentrations
of immunoglobulins (IgA, IgM), C3 and C4
complement and acute phase proteins are
changed in depressed patients (4,16,17).
Positive acute phase proteins such as the
α-1 acid glycoprotein, α-2 globulins, C-re-
active protein and haptoglobin are increased
in depression, while negative acute phase
proteins, such as albumin and transferrin,
are decreased (18).

The assessment of the capacity of periph-
eral blood mononuclear cells of depressed
patients to produce cytokines has also yielded
controversial results (19). An increase in
plasma concentration and in vitro produc-
tion of IL-1β, plasma concentration of soluble
IL-6 receptor, soluble IL-2 receptor and trans-
ferrin receptor were significantly higher in
depressed patients than in healthy controls
(20). A significant reduction in IL-1β, IL-2
and IL-3-like activity was observed in un-
treated depressed patients when compared
to controls and the IL-1β and IL-3-like activity
synthesis was significantly increased af-
after drug treatment (14). Elevations in the
concentrations of cytokines IL-1, interferon
alpha (IFN-α) and tumor necrosis factor
(TNF) and reduction in IL-2 have been re-
ported in depressed patients (4). Increased
serum levels of pro-inflammatory cytokine
IL-1 and of IL-6 have also been reported
(21).

The hormonal abnormalities described in
depression are secretion of corticotrophin-
releasing factor and a neurally mediated
hyperresponsivity of the adrenal gland to
adrenocorticotropic hormone (ACTH) (22).
There is an increased number of ACTH se-
cretory episodes combined with an increased
magnitude of cortisol-secretory episodes. De-
pending on the severity of depression and
age, 20-50% of patients are defined as dexam-
ethasone nonsuppressors. The corticotrophin-
releasing factor system in association
with altered sympathetic activity may be
changed in depression, resulting in altered
ACTH-cortisol ratios (23). Conflicting re-
results about other hormones such as basal
prolactin and thyroid hormone have been
reported in depression (22).

The present study was undertaken to
evaluate the levels of immune and hormonal
components in adult ambulatory patients with
depression in comparison with nondepressed
subjects.

Subjects and Methods

Subjects

The study was conducted on 40 non-
medicated ambulatory adults with depre-
sive episodes and 34 healthy nondepressed
volunteers (control group). All subjects, pa-
tients and controls, were submitted to clini-
cal evaluation according to CID-10 criteria
(24) and the severity of depression was de-
termined using the Hamilton Depression
Rating Scale (25). The depressed patients
were divided into two groups: 31 patients
with very severe depression, with Hamilton
Depression Scale scores higher than 23, and
9 patients with severe or moderate depre-
sion with Hamilton Depression Scale scores
ranging from 19 to 22 for severe depression
and from 14 to 18 for moderate depression.
The control group presented scale scores of
7 or less. The depressed and nondepressed subjects were seen at the Ambulatório de Clínica Médica, Hospital de Clínicas, Universidade Estadual de Londrina (UEL), Londrina, PR, Brazil, during the period from July 1998 to March 2000. Subjects younger than 18 years and older than 60 years were excluded from both groups. All subjects were required to be in good health, defined as the absence of chronic diseases which affect the immune system, HIV, and an acute or inflammatory response, for at least 2 weeks before the study and not to be taking medications with known effects on the immune system. Subjects taking recreational drugs and with a recent history of shock, malnutrition, irradiation, fever or cancer treatment were also excluded. The research was approved by the Ethics Research Committee of the UEL and written informed consent was obtained from all depressed and nondepressed subjects participating in the study after the procedures were fully explained.

**Immunological evaluation**

Immune determinations including total white blood cell count, differential counts and serum protein fractions were performed using standard procedures. Serum immunoglobulins (IgG, IgA, IgM), C3 and C4 complement, and C-reactive protein were determined by the immunonephelometric method (Dade Behring, Marburg, Germany), and plasma concentrations of IL-1β, IL-6, TNF-α and the soluble IL-2 receptor were determined by an automated chemoluminescent immunoenzymatic method (Immulite®, Diagnostic Products Co., Los Angeles, CA, USA). The lymphocyte response to phytohemagglutinin stimulation was determined by standard methods (26).

**Hormonal evaluation**

The hormonal determinations included triiodothyronine (T3), thyroxine (T4), thyrotrophin (TSH), prolactin (PRL), cortisol (blood samples were collected at 8:00 am), and the dexamethasone suppression test (DST). An oral dose of 1 mg dexamethasone was given at 11:00 pm to patients and control subjects. The next day, serum cortisol levels were assayed in blood samples collected at 4:00 and 11:00 pm. All of these hormonal evaluations were made by radio-immunoassay using commercial products. The result of the DST was considered to be normal when the cortisol values were less than 5 μg/dl from 8 to 24 h after an oral dose of 1 mg dexamethasone given at 11:00 pm (27). The serum levels of dehydroepiandrosterone sulfate (S-DHEA), ACTH, and growth hormone were assayed by an automated chemoluminescent immunoenzymatic method (Immulite®).

Whenever possible, blood samples from patients and controls were collected in the morning between 8:00 and 10:00 am and processed together on the same day to control for day-to-day variation in the assay. The blood samples were identified by a sequential number with no difference between groups (patients and controls). All assays were performed using standard procedures according to manufacturer instructions.

**Statistical analysis**

Data were analyzed statistically by the F-test for analysis of variance (ANOVA), Kruskal-Wallis test, and chi-square test. When the results showed significant differences, the Tukey test was used to compare the means obtained for the groups assayed. The results are reported as means ± standard deviation (SD). A difference between the two groups was considered to be statistically significant when P<0.05. All levels of significance were two-tailed.

**Results**

The characteristics of adult patients with
very severe, severe or moderate depression and of nondepressed (control group) subjects were as follows. Of the 40 depressed patients, 29 (72.5%) were females and 11 (27.5%) were males. The female to male ratio was 2:6:1:31 patients (8 males and 23 females) aged 18-58 years (35.90 ± 11.20) presented very severe depression with mean Hamilton Depression Scale scores ranging from 23 to 50 (32.84 ± 6.75). The diagnosis of severe or moderate depression was made in 9 patients (3 males and 6 females) aged 19-57 years (36.44 ± 14.23) with Hamilton Depression Scale scores ranging from 18 to 22 (20.44 ± 1.59). Among the 34 nondepressed subjects (9 males and 25 females) aged 18-56 years (34.94 ± 11.78), the mean Hamilton Depression Scale score ranged from 0 to 7 (2.23 ± 2.24). The severity of depression among the groups was statistically evaluated by the Kruskal-Wallis test. The mean values, 2.23 ± 2.24, 20.44 ± 1.59 and 32.84 ± 6.75, were significantly different when compared by the Tukey test (P<0.05).

The results of the hematologic determinations on peripheral blood for leukocytes, neutrophils, typical lymphocytes, monocytes, eosinophils and basophils for the patients with very severe, severe or moderate depression did not differ significantly from the values obtained for the control group.

Table 1 shows the blood analyses. The albumin fraction percentage was lower in the very severely depressed patients when compared with the control group (P<0.05). The absolute and relative serum levels of α-1, α-2, and β-globulins were significantly higher in the very severely depressed patients than in the control group (P<0.05). The results of other biochemical parameters assayed such as mucoprotein, total serum proteins and γ-globulin did not differ significantly among groups.

Of the immune parameters evaluated, serum IgG, IgM, IgA, C-reactive protein, C3, C4, cytokines IL-1β, and TNF-α levels did not differ significantly between patients and controls. Serum IL-6 levels were lower in the severely or moderately depressed patients when compared to the controls (P<0.05). Serum soluble IL-2 receptor levels were lower in the severely or moderately depressed patients when compared with very severely depressed patients and the control group (P<0.05).

The lymphocyte stimulation response to phytohemagglutinin, expressed by the stimulation index, in the very severely, severely or moderately depressed patients and control group was 1.642, 1.014, and 2.129, respectively. Statistical analysis showed that the stimulation index was lower in the severely or moderately depressed patients when compared to the control group (P<0.05).

A subject was considered to be a nonsuppressor if the serum cortisol result obtained from a sample collected at 4:00 or 11:00 pm was similar to or higher than 5 μg/dl. The results of the DST revealed that the rate of nonsuppressors did not differ significantly between depressed patients and controls (21.1 and 36.7%, respectively). The serum levels of T3, T4, TSH, PRL, cortisol at 8:00 am, cortisol after DST (4:00 and 11:00 pm),

<table>
<thead>
<tr>
<th>Parameter (mean ± SD)</th>
<th>Control group (N = 34)</th>
<th>Severely or moderately depressed patients (N = 19)</th>
<th>Very severely depressed patients (N = 31)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mucoprotein (mg/dl)</td>
<td>0.86 ± 0.36</td>
<td>0.92 ± 0.20</td>
<td>0.99 ± 0.37</td>
</tr>
<tr>
<td>Total protein (g/dl)</td>
<td>7.10 ± 0.78</td>
<td>7.25 ± 0.69</td>
<td>7.50 ± 0.76</td>
</tr>
<tr>
<td>Albumin (%)</td>
<td>53.0 ± 7.36</td>
<td>47.40 ± 4.52</td>
<td>46.76 ± 5.93*</td>
</tr>
<tr>
<td>Albumin (g/dl)</td>
<td>3.79 ± 0.60</td>
<td>3.47 ± 0.35</td>
<td>3.43 ± 0.53</td>
</tr>
<tr>
<td>α-1 Globulin (%)</td>
<td>2.50 ± 0.92</td>
<td>3.05 ± 0.74</td>
<td>3.38 ± 1.09*</td>
</tr>
<tr>
<td>α-1 Globulin (g/dl)</td>
<td>0.18 ± 0.06</td>
<td>0.22 ± 0.04</td>
<td>0.24 ± 0.07*</td>
</tr>
<tr>
<td>α-2 Globulin (%)</td>
<td>8.67 ± 1.94</td>
<td>9.64 ± 2.12</td>
<td>10.07 ± 1.63*</td>
</tr>
<tr>
<td>α-2 Globulin (g/dl)</td>
<td>0.62 ± 0.16</td>
<td>0.71 ± 0.13</td>
<td>0.75 ± 0.15*</td>
</tr>
<tr>
<td>β-Globulin (%)</td>
<td>15.17 ± 2.65</td>
<td>14.55 ± 2.49</td>
<td>17.16 ± 2.86*</td>
</tr>
<tr>
<td>β-Globulin (g/dl)</td>
<td>1.09 ± 0.21</td>
<td>1.08 ± 0.21</td>
<td>1.28 ± 0.26*</td>
</tr>
<tr>
<td>γ-Globulin (%)</td>
<td>20.32 ± 4.94</td>
<td>24.91 ± 5.22</td>
<td>22.43 ± 3.92</td>
</tr>
<tr>
<td>γ-Globulin (g/dl)</td>
<td>1.46 ± 0.41</td>
<td>1.84 ± 0.43</td>
<td>1.68 ± 0.38</td>
</tr>
</tbody>
</table>

Data are reported as means ± SD.
*P<0.05 compared to control group (ANOVA test, Tukey test).
S-DHEA, ACTH, and growth hormone did not differ between patients and controls.

**Discussion**

The study evaluated the immunological and hormonal parameters of a group of very severely, severely or moderately depressed patients compared to nondepressed subjects. The female to male ratio of 2.6:1 was in keeping with the gender distribution found among patients with depression (28). Peripheral blood cell counts showed no differences in the number of white blood cells, neutrophils, lymphocytes or monocytes, in accordance with most of the cell enumeration studies in depressive disorders reporting an unaltered number of white blood cells, neutrophils, lymphocytes, total T-cells as well as T cell subsets, helper suppressor cell ratio, and B cells (14,29,30).

The significant reduction observed in serum albumin levels among very severely depressed patients, and a significant elevation in the serum levels of positive acute phase proteins among the very severely depressed patients when compared with the control subjects are consistent with most published reports (4,18,19,29,31). The findings that stressors may be able to activate the release of proinflammatory cytokines and an acute-phase response in the absence of an immune challenge further support the notion that the immune system may be recruited to participate in the behavioral response to stress, and therefore may contribute to the biochemical and molecular biological changes that characterize depression (21). The acute phase proteins are mediated by the pro-inflammatory cytokines, mainly by IL-1, IL-6 and TNF which in depressed patients seem to be correlated with the severity of the disease and the hyperactivity of the hypothalamic-pituitary-adrenocortical axis. Research on cytokine regulation in depression has been controversial. Some researchers have reported a reduced production of IL-1β (14) and IL-2 (4,14) by peripheral blood mononuclear cells in patients with major depression before treatment, reflecting an adaptive feedback mechanism for the prevention of pituitary ACTH hypersecretion. However, others have reported that depression may be associated with higher serum concentrations of IL-1, IFN-α and TNF (4), IL-1β, IL-6, and IFN-γ (32). An increase in plasma IL-6, soluble IL-6 receptor and soluble IL-2 receptor concentrations among depressed patients was reported in a study in which the authors stated that the increased production of IL-6 may represent a contributing factor to the various immune disorders encountered in major depression and perhaps to the pathophysiology or pathogenesis of this illness. However, these results were obtained among DST nonsuppressor depressed patients (20). The present study evaluated all depressed patients, DST suppressor and DST nonsuppressor. This difference could contribute to the disagreement of the results obtained. Another difference that could explain the results is that this study evaluated only ambulatory depressed patients, and not hospitalized depressed patients.

Mitogen-induced lymphocyte stimulation in the depressed patients assayed here confirmed the impaired cellular immune response, as revealed by the decreased lymphocyte proliferation in response to phytohemagglutinin, in agreement with previous studies (10,13,27).

The results obtained by T3, T4 and TSH evaluation were not significantly different among groups. The majority of depressed patients appeared to be euthyroid and changes in these measurements have been reported in a substantial minority of depressed patients. The results concerning basal PRL levels reported for depressed patients are conflicting. Basal PRL has been found to be low, normal and elevated and the underlying cause of the disturbances in PRL regulation in affective disorders is still unclear (22). As regards basal plasma cortisol and cortisol levels after
DST, the difference was not significant among the groups assayed. Only a 21.1% rate of nonsuppression was demonstrated among depressed patients evaluated in this study. According to previous studies, an impaired feedback inhibition resulting in an abnormal DST is registered in approximately 45% of patients with depression (22). However, the results obtained among the controls (36.7% DST nonsuppressor) are in agreement with reports about the hypothalamic-pituitary adrenocortical alteration in healthy controls at risk for depression. In healthy subjects who had never suffered from minor or major psychiatric illness, but whose family members were highly loaded with depression, several neurobiological signs of depression were found to be present, including the response to the DST-CRH test (23). These findings could suggest the usefulness of the DST results as a biological marker in those individuals from a high-risk population who present abnormal DST results. These subjects could be at higher risk for developing depression than those who have normal DST results.

Depression is by no means a homogeneous disorder, but is in fact a complex phenomenon which has many subtypes, and probably more than one etiology (32). A variety of interacting factors may have contributed to the results obtained in the present study such as patient age, dexamethasone suppressor or nonsuppressor status, whether the patients are experiencing acute depression or are in remission, the severity and/or duration of depression and whether patients are drug-free (and for how long), and the presence or absence of any kind of therapy (27,33,34). The difficulty to control some variables which could affect the parameters evaluated, such as tobacco use, alcohol and caffeine consumption, activity and exercise levels, may have affected the results (13,35,36). The fact that the control subjects were volunteers and self-selected shows that this sample does not represent the general population.

The results obtained added to other previous reports about the evidence of an association of immune abnormalities, both activation and depression, with depressive illness. The immune system activation would be characterized by the increased acute phase proteins and the immune system depression by the reduced mitogen-stimulated lymphocyte proliferation observed in the depressed patients evaluated. However, the conclusions reached in light of these results require a cautious interpretation and the precise clinical relevance of these findings requires further investigation.

References

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