Bronchial hyperresponsiveness and analysis of induced sputum cells in Crohn’s disease

R.M. Bartholo1, C. Zaltman1, C. Elia1, A.P. Cardoso2, V. Flores2, P. Lago2, L. Cassabian2, F. Carvalho Dorileo2, and J.R. Lapa-e-Silva2

Abstract

With the aim of investigating the presence of latent inflammatory process in the lungs of patients with Crohn’s disease, 15 patients with Crohn’s disease were evaluated by spirometry, the methacholine challenge test, induced sputum, and skin tests for inhaled antigens. Serum IgE, erythrocyte sedimentation rate and hematocrit were also determined. The patients were compared with 20 healthy controls by the Mann-Whitney and Fisher exact tests. Their respiratory physical examination was normal. None had a personal or family history of clinical atopy. None had a previous history of pulmonary disease, smoking or toxic bronchopulmonary exposure. None had sinusitis, migraine, diabetes mellitus, or cardiac failure. Four (26.6%) of the patients with Crohn’s disease had a positive methacholine challenge test whereas none of the 20 controls had a positive methacholine test (P = 0.026, Fisher exact test). Patients with Crohn’s disease had a higher level of lymphocytes in induced sputum than controls (mean 14.59%, range 3.2-50 vs 5.46%, 0-26.92%, respectively; P = 0.011, Mann-Whitney test). Patients with Crohn’s disease and a positive methacholine challenge test had an even higher percentage of lymphocytes in induced sputum compared with patients with Crohn’s disease and a negative methacholine test (mean 24.88%, range 3.2-21.69%; P = 0.047, Mann-Whitney test). The simultaneous findings of bronchopulmonary lymphocytosis and bronchial hyperresponsiveness in patients with Crohn’s disease were not reported up to now. These results suggest that patients with Crohn’s disease present a subclinical inflammatory process despite the absence of pulmonary symptoms.

Correspondence
J.R. Lapa-e-Silva
Hospital Universitário
Clementino Fraga Filho
Av. Brigadeiro Trompowsky, s/n
Subsolo
Ilha do Fundão
21941-590 Rio de Janeiro, RJ
Brasil
Fax: +55-21-2290-3520
E-mail: jrlapa.ntg@terra.com.br

Research supported by FAPERJ and CNPq.

Received January 22, 2004
Accepted December 9, 2004

Key words
• Inflammatory bowel disease
• Crohn’s disease
• Bronchial hyperresponsiveness
• Lymphocytosis

Introduction

Many extraintestinal manifestations affecting various organs or organ systems have been described in association with inflammatory bowel disease (1) but clinically significant lung involvement is rare (2). However, pulmonary lesions have been reported in patients with Crohn’s disease, such as unexplained chronic bronchitis, bronchiolitis...
obliterans with organizing pneumonia, pleuroperticarditis (1), recurrent pulmonary edema (3), granulomatous (4,5) and nongranulomatous interstitial pulmonary disease (6), granulomatous lung masses (7), bronchiectasis (8), and granulomatous bronchiolitis (9). Tracheal involvement in Crohn’s disease is even more unusual and consists of mucosal inflammation (10-12). Drug therapy is another complicating factor, since sulfasalazine used in the treatment of Crohn’s disease can also cause reactions in the lung (13-15).

Crohn’s disease should be regarded as a multisystemic disorder that involves mainly the gastrointestinal tract. Although the overall prevalence of concomitant bronchopulmonary manifestations is only 0.4% (16), subclinical alterations in at least half of the adults with Crohn’s disease have been demonstrated (17,18) suggesting underlying bronchial inflammation. This pulmonary involvement can be reflected by an increased lymphocyte count in the bronchoalveolar lavage fluid (19,20) and/or lung function abnormalities (21,22).

Latent pulmonary involvement has been reported in a pediatric population with Crohn’s disease, despite a short duration of the disease and absence of smoking (22). An increased bronchial hyperresponsiveness in patients with inflammatory bowel disease with no bronchopulmonary symptoms and with normal baseline lung function has been reported (23). Recently, Mansi et al. (24) reported bronchial hyperresponsiveness to methacholine in children with Crohn’s disease with no evidence of airway disease.

Recent studies have suggested that the common mucosal immune system may be involved in various T-lymphocyte-mediated disorders. An increased frequency of asymptomatic bronchial hyperresponsiveness has been reported in patients with Crohn’s disease, and a subclinical accumulation of activated T lymphocytes has been demonstrated in the lung of patients with Crohn’s disease without respiratory symptoms (19,25). No study has demonstrated thus far the simultaneous presence of alveolar lymphocytosis and bronchial hyperresponsiveness to methacholine in respiratory asymptomatic patients with Crohn’s disease. In the present study, we compared bronchial responsiveness to methacholine and cellularity of the sputum of patients with Crohn’s disease and healthy controls.

Subjects and Methods

Subjects

The patients were recruited at the outpatient Gastroenterology clinic of the Clementino Fraga Filho University Hospital and Cardoso Fontes Municipal Hospital, Rio de Janeiro, RJ, Brazil. Fifteen patients with Crohn’s disease were included, comprising 7 females and 8 males, 18 to 53 years of age (mean 33.93). The following confounding conditions were excluded: a history of clinical manifestations of atopy, cigarette smoking, a previous history of pulmonary disease, bronchopulmonary exposure to toxic chemicals or fumes, sinusitis, associated diseases such as diabetes mellitus, cardiac failure, migraine or cystic fibrosis, and a family history of asthma. Physical examination and chest radiographs were normal. Baseline pulmonary function was normal. The diagnosis of Crohn’s disease was made by internationally recognized and accepted criteria (26). Duration of Crohn’s disease at the onset of the study ranged from 6 months to 30 years (mean duration 8.3 years). Three patients had systemic manifestations such as arthralgias and one had sacroiliitis. Crohn’s disease activity was measured by the Harvey-Bradshaw index (27). One patient had active disease (Bradshaw index = 7). Thirteen patients were treated with prednisone (12.5 to 40 mg/day) and 2 patients were treated with budesonide (enema). Associated drugs were metronidazole (3 patients), sulfasalazine (2 patients) and mesalazine (2 patients).
The control group consisted of 20 healthy volunteers, 12 females and 8 males ranging in age from 17 to 55 years (mean 27.6). None were cigarette smokers or had a personal or family history of asthma or atopy. Physical examination and chest radiographs were normal. Baseline pulmonary function was normal.

All patients and control subjects agreed to take part in the study and signed an informed consent form. The University Hospital Ethics Committee approved the study.

Atopy

Atopy was evaluated by skin prick tests for the following aeroallergens: *Dermatophagoides pteronyssinus*, *Dermatophagoides farinae*, *Blomia tropicalis*, and *Aspergillus fumigatus*, compared with histamine as positive control, and the diluent as negative control. The tests were considered positive when they induced a weal equal to or larger than 3 mm. No differences were observed between healthy controls and patients.

Blood tests

Erythrocyte sedimentation rate (Westergren method), serum IgE (nephelometry method) and hematocrit were determined to compare the two groups.

Bronchial responsiveness

Bronchial responsiveness was measured in the morning by methacholine stimulation. Methacholine bromide solutions (Sigma, St. Louis, MO, USA) were dissolved in 0.9% physiologic saline, stored at 4°C, and used within days of preparation. The vials were removed from the refrigerator 30 min before testing. Nose clips were used to prevent dilution of the inhaled solution with air through the nose during slow inhalation. The aerosols were delivered by a dosimeter (Spira Elektro 4; Hengityshoitokefkuf, Helsinki, Finland). The nebulizer was used with an output of 9 µl per 0.6 s (total delivery of 0.045 ml). After measurement of forced expiratory volume in 1 s (FEV1), subjects inhaled an aerosolized physiologic saline solution. FEV1 was measured again after saline inhalation and the value obtained was considered to be the control value. Provided this FEV1 value had not fallen to less than 95% of the baseline value, a methacholine challenge was carried out. Subjects successively inhaled a methacholine solution of 0.075, 0.15, 0.31, 0.62, 1.25, 2.5, 5.0, 10.0, and 20.0 mg/ml and FEV1 was measured 30 and 90 s after each concentration using a Collins GS spirometer (Louisville, CO, USA). The test was stopped when FEV1 fell to less than 80% of the control value recorded. In this case, the provocative dose of methacholine producing a 20% fall in FEV1 (PD20) was read from the log-response curve by linear interpolation. Bronchial hyperresponsiveness was defined as a methacholine PD20 of less than 7.80 µmol according to the European standardization for bronchoprovocation tests (28).

Induced sputum

Sputum was induced by the method of Pin et al. (29). The patients were pretreated with 200 µg inhaled salbutamol to inhibit airway constriction and an aerosol of hypertonic saline was then inhaled from an ultrasonic nebulizer. The concentrations of saline were 3, 4, and 5%, and the duration of inhalation was 7 min for each concentration. After each period of inhalation, the patient was asked to blow his nose, rinse his mouth, and swallow water to minimize contamination with postnasal dripping and saliva, and peak flow was measured in order to record any possible fall in this parameter. The patient was asked to expectorate sputum into a container. If peak flow fell 10% or more, inhalations were discontinued to prevent severe bronchoconstriction.

The expectoration was processed as soon
as possible and within 2 h. Sputum was considered adequate when it presented dense and viscid portions that macroscopically appeared to be free of salivary contamination. These portions were selected and placed in a 15-ml polystyrene tube (Falcon 2097 Becton Dickinson, Franklin Lakes, NJ, USA). The selected portion was treated with dithiothreitol (Sigma) diluted to 0.1% by the addition of distilled water to dissociate the disulfide bonds of the mucus. The mixture was vortexed for 15 s and gently aspirated in and out of a Pasteur pipette to ensure mixing. The polystyrene tube was rocked for 15 min in a bench rocker. Then, 4 additional volumes of phosphate-buffered saline were added to stop the effect of dithiothreitol on the cell suspension and the rocking was continued for 5 min. The suspension was centrifuged and the supernatant was aspirated and stored in 1.5-ml Eppendorf tubes (National Scientific Supply Co., San Rafael, CA, USA) at -70ºC for later examination of fluid-phase components.

The cell pellet was resuspended in 1 ml PBS and a total leukocyte count was performed with hemocytometer. Cell viability was determined by Trypan blue exclusion and the total and absolute numbers of cells of processed sputum were calculated. The cell suspension was adjusted to 1.0 x 10⁶/ml and cytospins were prepared (Cytospin 3 Shandon; Astmoor, Runcorn, England). Slides were airdried and stained with the Diff-Quick stain (Laborclin, Pinhais, PA, Brazil) and differential cell counts of at least 200 nonsquamous cells were performed.

**Statistical analysis**

The Mann-Whitney test was used for comparison between patients and controls, and the Fisher exact test was used when an expected value was less than 5. The level of significance was set at $P \leq 0.05$ in all analysis. The statistical package Epi-Info 6 was used for calculations.

**Results**

**Bronchial responsiveness**

Fifteen patients with Crohn’s disease without respiratory symptoms and 20 healthy controls participated in the study. Four patients with Crohn’s disease (26.7%) had a positive methacholine challenge indicating bronchial hyperresponsiveness. The measurements of PD20 in these patients were 1.85, 6.08, 7.48, and 7.48 µmol. The other 11 Crohn’s disease patients and the 20 healthy controls had negative a methacholine challenge, i.e., with PD20 above the cut-off point of 7.80 µmol ($P = 0.026$, Fisher exact test).

**Induced sputum**

Of the 15 patients with Crohn’s disease who participated in the study, one did not produce adequate induced sputum, as de-

<table>
<thead>
<tr>
<th>Group</th>
<th>% Lymphocytes (range)</th>
<th>% Macrophages (range)</th>
<th>% Eosinophils (range)</th>
<th>% Neutrophils (range)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crohn’s disease (N = 15)</td>
<td>14.59 (3.20-50.00)*</td>
<td>72.33 (45.31-82.35)</td>
<td>0 (0-13.04)</td>
<td>3.77 (0-13.04)</td>
</tr>
<tr>
<td>Control (N = 20)</td>
<td>5.46 (0-26.92)</td>
<td>79.80 (54.28-100)</td>
<td>0.13 (0-0.94)</td>
<td>6.33 (0-37.14)</td>
</tr>
</tbody>
</table>

* $P < 0.05$ compared to control (Mann-Whitney test).
fined above. The lymphocyte counts for these patients were significantly higher than for the control group (Table 1). Differential counts of induced sputum were not statistically different concerning other cell types. Patients with Crohn’s disease and a positive methacholine challenge test had an even higher percentage of lymphocytes in induced sputum compared with other Crohn’s disease patients but a negative methacholine test (mean 24.88%, range 12.87-50 vs 10.48%, 3.2-21.69%; P = 0.047, Mann-Whitney).

Discussion

The results of the present study demonstrated that bronchial hyperresponsiveness occurs in a high proportion (26.7%) of adult patients with Crohn’s disease compared to 0% of normal healthy controls. The PD20 values found in these patients, however, were higher than most of those seen in asthmatic individuals (30). This condition was observed despite the absence of clinical, radiological and functional evidence of airway disease, in agreement with data reported by Louis et al. (23) who observed bronchial hyperresponsiveness in 45% of the patients with intestinal bowel disease, including patients with ulcerative colitis. In 2000, Mansi et al. (24) reported bronchial hyperresponsiveness in 71% of children and adolescents with Crohn’s disease.

Bronchial hyperresponsiveness is a phenomenon related to multiple factors, including inflammation of the airways by several inflammatory cell types and their products, epithelial damage, microvascular leakage, and autonomic neural mechanisms. Conditions other than asthma can be associated with a positive pharmacologic challenge with methacholine or other agonists, such as allergic rhinitis, bronchiectasis, chronic obstructive pulmonary disease, cystic fibrosis, hypersensitivity pneumonitis, left ventricular failure, lung and heart-lung transplantation, sarcoidosis, tropical pulmonary eosinophilia, viral upper or lower respiratory tract infection within 6 weeks of testing, and rarely in normal subjects (31). The cut-off point of 7.8 µm for bronchial hyperresponsiveness used in this study was initially established by Woolcock et al. (32) in 1983 for the cumulative test using histamine and confirmed in a subsequent study (33) using methacholine. This value was maintained in the 1993 European standardization (28) and is used by most laboratories.

The present study revealed an increased percentage of lymphocytes in the differential counts of induced sputum in patients with Crohn’s disease compared with normal controls. Wallaert et al. (19) reported a high proportion of increased alveolar lymphocytosis in patients with Crohn’s disease free of clinical symptoms and with normal chest roentgenograms. The significance of the increased percentage of lymphocytes is not clear. It is currently admitted that a part of the immune system specific to the gastrointestinal tract is common to all mucosal surfaces (34). Therefore, the association of Crohn’s disease and lymphocyte alveolitis might be the result of an immunologic mechanism. The lymphocytes sensitized to antigens at one mucosal site could circulate and populate the mucosal surfaces of the lung (19). Muller et al. (34) showed that CD4 were vigorously activated in the intestinal mucosa of patients with active Crohn’s disease.

The proportion of lymphocytes in the induced sputum of patients with Crohn’s disease and a positive methacholine challenge was higher than in patients with Crohn’s disease and a negative methacholine challenge. This fact might represent the existence of a minimum inflammation level from which the methacholine challenge would become positive.

Despite the fact that two patients with a positive bronchoprovocation challenge were using sulfasalazine, a drug reaction was not probable. This would have resulted in an increased number of eosinophils in the induced
sputum or eosinophilia, which we did not observe. Another aspect was the presence of an interstitial infiltrate which was not found.

There is increasing evidence that subclinical Crohn’s disease-associated respiratory alterations can represent a problem in this group of patients. It is important to be alert to this clinical disorder and to try to detect it as early as possible in order to prevent future respiratory disturbances.

References


