Cytotoxicity of *Marchantia convoluta* leaf extracts to human liver and lung cancer cells

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Abstract

The cytotoxicity of three extracts (petroleum ether, ethyl acetate and n-butanol) from a plant used in folk medicine, *Marchantia convoluta*, to human non-small cell lung carcinoma (H1299) and liver carcinoma (HepG2) cell lines was tested. After 72-h incubation of lung and liver cancer cell cultures with varying concentrations of extracts (15 to 200 µg/mL), cytotoxicity was determined by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay and reported in terms of cell viability. The extracts that showed a significant cytotoxicity were subjected to gas chromatography-mass spectrometry analysis to identify the components. The ethyl acetate, but not the petroleum ether or n-butanol extract, had a significant cytotoxicity against lung and liver carcinoma cells with IC₅₀ values of 100 and 30 µg/mL, respectively. A high concentration of ethyl acetate extract (100 µg/mL) rapidly reduced the number of H1299 cells. At lower concentrations of ethyl acetate extract (15, 30, and 40 µg/mL), the numbers of HepG2 cells started to decrease markedly. Gas chromatography-mass spectrometry analysis of the ethyl acetate extract revealed the presence of several compounds such as phytol (23.42%), 1,2,4-tripropylbenzene (13.09%), 9-cedranone (12.75%), ledene oxide (7.22%), caryophyllene (1.82%), and caryophyllene oxide (1.15%). HPLC analysis result showed that there were no flavonoids in ethyl acetate extract, but flavonoids are abundant in n-butanol extract. Further studies are needed regarding the identification, toxicity, and mechanism of action of active compounds.

Key words

- *Marchantia convoluta* extract
- Cytotoxicity assay
- HepG2 and H1299 carcinoma cell lines
- Phytol
- Caryophyllene oxide

Introduction

Marchantiaceae plants are well-known traditional Chinese medicinal herbs extensively used to treat skin tumefaction, to protect the liver and to treat hepatitis, being also used as antipyretics (1-3). Large numbers of Marchantiaceae plants occur in Guangxi Zhuang Autonomous District such as *Marchantia polymorpha*, *M. convoluta* and *M. paleacea*. Many studies on the chemical constituents and bioactivities of *M. polymorpha* have been reported (4-13). These species grow together and are difficult to distinguish from one another because of their genetic similarity. *M. convoluta* is only found in
China (14) and is quite rare.

The major identified constituents of *M. convoluta* are flavonoids, triterpenoids and steroids (15-18). The flavonoids of *M. convoluta* consist mainly of quercetin, luteolin, apigenin, and their O- and C-glycosides (15-17). A high dosage of flavonoids from *M. convoluta* (20 and 40 µg/mL) can significantly reduce the activity of alanine aminotransferase and aspartate aminotransferase in the serum of mice with acute hepatic injury caused by CCl₄ and increase the contents of total protein and alkaline phosphatase, as well as inhibit the auricle tympanitis of mice caused by dimethylbenzene (1). Flavonoids from *M. convoluta* strongly inhibit colibacillus, typhoid bacillus, *Staphylococcus aureus*, *Bacillus enteritidis*, hemolytic streptococci type B, and *Diplococcus pneumoniae* and have antibiotic, anti-inflammatory and diuretic effects on mice (1).

In the present study, the effects of extracts of *M. convoluta* on human carcinoma cells were investigated. Non-small cell lung carcinoma cell line H1299 and the liver cancer cell line HepG2 were used. Some of the chemical constituents of petroleum ether and ethyl acetate extracts were also identified by gas chromatography-mass spectrometry (GC-MS).

**Material and Methods**

**General**

Ethanol, petroleum ether, ethyl acetate, and n-butanol, all of analytical reagent grade, were purchased from Tianjin Damao Chemical Reagent Factory (Dongli District, Tianjin, China).

**Plant material**

The leaves of *M. convoluta* were collected in Shangling City, Guangxi Zhuang Autonomous District, China, in August 2003. The specimen (No. 20041364) was identified by Zhou Zi-jing at the Biology Depart-
suspended in 10 mL of medium to prepare a single cell suspension. The density of viable cells was determined by Trypan blue exclusion in a hemocytometer and the preparation was then diluted with medium to yield previously determined optimal plating densities for H1299 and HepG2. The HepG2 and H1299 cells were seeded on 96-well plates at a final concentration of 5 x 10^4 cells per 100 µL medium per well 24 h before the assay. The cell suspensions were then incubated at 37ºC to permit cell attachment. After 24 h the cells were treated with the extracts. Each extract was initially dissolved in ethanol and 500 µg/mL of each extract was tested initially against both cancer cell lines. On the basis of the results obtained, extracts were considered to be active when they produced less than 50% survival after an exposure time of 72 h. The active extracts were further diluted in medium to produce 7 concentrations of 0, 15, 30, 40, 50, 100, 200 µg/mL of each extract and 100 µL/well of each concentration was added to the plates in six replicates. At the end of the exposure time the medium was removed and MTT assays were then performed using the cell titer kit™ (Promega Corp., Madison, WI, USA). Twenty milliliters MTT (5 mg/mL) in PBS was incubated with cells on a 96-well plate for 2 h at 37ºC. Subsequently, the medium containing MTT was removed, and 100 mL acidified isopropanol (0.04 mol/L HCl) was added. The absorbance of each sample was measured at 570 nm using a microplate reader (Bio-Rad Laboratories, Orlando, FL, USA, model 3550). The data were normalized (A570 nm) and the mean absorbance was plotted against drug concentration. Three replicate plates were used to determine the cytotoxicity of each extract.

Gas chromatography-mass spectrometry

GC-MS analysis was performed using an Agilent (Palo Alto, CA, USA) system consisting of a model 6890 gas chromatograph, a model 5973 mass selective detector, and an Agilent ChemStation data system. The GC column was an HP-5 ms fused silica capillary column (30 m x 0.25 mm, 0.25 µm). The carrier gas was helium at a flow rate of 1.0 mL/min. Oven temperature was 60ºC for 5 min, and then increased to 250ºC at a rate of 5ºC/min. Injector and detector temperatures were 250 and 265ºC, respectively. One microliter of diluted solution was injected into the GC/MS apparatus with a split ratio of 1/60. The ionization energy was 70 eV with a scan time of 1 s and mass range of 40-540 amu. The percentages of the compounds were calculated by the area normalization method without considering response factors. The components of the oil were identified by comparison of their mass spectra with those of a computer library (NIST database/ChemStation data system) (19).

High-performance liquid chromatography

The chemical constituents of three extracts were investigated by high-performance liquid chromatography (HPLC) performed according to the literature (17). HPLC analysis was performed using equipment from Shimadzu (Shimadzu, Kyoto, Japan): a Shimadzu LC-2010A liquid chromatograph, a Shimadzu SPD-M10A Diode Array Detector and a Shimadzu Class-vp V6.12 SP4 offline processing system. Flavonoids were analyzed using a Kromasil RP-C18 column (250 x 4.6 mm ID, 5 µm; Hanbon Science & Technology Co., Ltd., Huaiyin, China). The mobile phase consisted of a mixture, methanol-acetonitrile-acetic acid-phosphoric acid-H₂O (200:100:10:10:200, v/v) and the solution was degassed by suction-filtration through a nylon membrane. The detecting wavelength was 352 nm. The flow rate was 0.60 mL/min and the sensitivity was set at 0.05 AUFS. The quantity volume of injecting sample was 6.0 µL. The HPLC system was operated at ambient temperature (28 ± 1ºC).
Statistical analysis

Data are reported as the mean ± SD for at least three determinations. Statistical analysis was performed using the Student t-test, with the level of significance set at P < 0.05.

Results

IC₅₀

The inhibitory effects of the extracts were determined by exposure of the human non-small cell lung carcinoma cell line H1299 and the human hepatocellular carcinoma cell line HepG2 to increasing concentrations of all extracts in a stepwise manner for 72 h. The concentration of each extract which reduced cell survival by 50% (IC₅₀) was determined from cell survival curves and the results are presented in Table 1.

As indicated in Table 1, treatment of H1299 and HepG2 cells with ethyl acetate extract resulted in loss of cell viability, whereas both cell lines were more resistant to the petroleum ether and n-butanol extracts, as shown by the respective IC₅₀ of H1299 and HepG2 cells.

Effect of the ethyl acetate extract on cell proliferation

The effect of all extracts on different cell lines was studied by measuring cell numbers by the MTT assay after treatment of the cultures with each extract for 72 h. The treatment of all cell lines with the ethyl acetate extract clearly reduced the cell numbers (Figure 1A,B), whereas the petroleum ether and n-butanol extracts did not have an evident inhibitory effect on cell proliferation.

The effect of extracts on cell viability was evaluated by determining the percentage of MTT reduction upon incubation of HepG2 and H1299 cells with increasing extract concentrations in the range 15-200 µg/mL. As shown in Figure 1A,B, the ethyl acetate extract produced a dose-dependent reduction in cell viability. Figure 1A shows that a high concentration of ethyl acetate extract (100 µg/mL) rapidly reduced the number of H1299

<table>
<thead>
<tr>
<th>Extract</th>
<th>H1299 IC₅₀ (µg/mL)</th>
<th>HepG2 IC₅₀ (µg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Petroleum ether</td>
<td>&gt;500</td>
<td>&gt;500</td>
</tr>
<tr>
<td>Ethyl acetate</td>
<td>100</td>
<td>30</td>
</tr>
<tr>
<td>n-butanol</td>
<td>&gt;200</td>
<td>&gt;200</td>
</tr>
</tbody>
</table>

Cells (5 x 10⁵ mL⁻¹) were cultured with various amounts of each extract at 37°C and 5% CO₂ for 72 h. Cytotoxicity was measured with the MTT assay. IC₅₀ = extract concentration causing a 50% decrease in the survival curve.
cells. At lower concentrations of the extract (15, 30, and 40 µg/mL), the numbers of HepG2 cells started to decrease markedly (Figure 1B), whereas no H1299 cell inhibition was observed at the concentration of 15 µg/mL. These results suggest that the ethyl acetate extract has an obvious toxic effect on HepG2 cells at low concentrations but inhibits H1299 cells only at higher concentration. The inhibition of proliferation and induction of cell death observed occurred in a concentration- and dose-dependent manner.

The petroleum ether and n-butanol extracts of *M. convoluta* had no effect on the human non-small cell lung carcinoma cell line H1299 (IC50 >500 and 200 µg/mL, respectively) and human liver carcinoma cell line HepG2 (IC50 >500 and 200 µg/mL, respectively) at any concentration.

**Gas chromatography-mass spectrometry**

Extracts of different compositions can be obtained by different extraction methods applied to natural products (20-24). The GC-MS profile of the ethyl acetate extract is shown in Figure 2. The separation obtained by GC was excellent. The GC-MS analytical results for the ethyl acetate extract are shown in Table 2.

As shown in Figure 2, a total of eleven compounds accounting for 70.04% of the

![Gas chromatography-mass spectrometry profile of the ethyl acetate extract.](image)

Table 2. Composition of the ethyl acetate extract obtained by GC-MS.

<table>
<thead>
<tr>
<th>Compound</th>
<th>RT/min</th>
<th>Reliability to MS standard</th>
<th>Relative content</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caryophyllene</td>
<td>13.49</td>
<td>99%</td>
<td>1.82%</td>
</tr>
<tr>
<td>9-Cedranone</td>
<td>15.25</td>
<td>90%</td>
<td>12.75%</td>
</tr>
<tr>
<td>Diepi-alpha-cedrene epoxide</td>
<td>15.42</td>
<td>91%</td>
<td>1.37%</td>
</tr>
<tr>
<td>Caryophyllene oxide</td>
<td>15.56</td>
<td>93%</td>
<td>1.15%</td>
</tr>
<tr>
<td>Eudesma-4(14),11-diene</td>
<td>16.38</td>
<td>80%</td>
<td>2.27%</td>
</tr>
<tr>
<td>Ledene oxide</td>
<td>16.44</td>
<td>90%</td>
<td>7.22%</td>
</tr>
<tr>
<td>Bi-1-cycloocten-1-yl</td>
<td>17.21</td>
<td>88%</td>
<td>1.46%</td>
</tr>
<tr>
<td>14-Methylpentadecanoic acid</td>
<td>19.66</td>
<td>98%</td>
<td>3.07%</td>
</tr>
<tr>
<td>Tetradecanoic acid, methyl ester</td>
<td>20.84</td>
<td>96%</td>
<td>2.42%</td>
</tr>
<tr>
<td>1,2,4-Tripropylbenzene</td>
<td>22.27</td>
<td>84%</td>
<td>13.09%</td>
</tr>
<tr>
<td>Phytol</td>
<td>22.64</td>
<td>87%</td>
<td>23.42%</td>
</tr>
</tbody>
</table>

RT = Retention time; Reliability to MS standard = match degree (%) of mass spectrum to the standard compounds; Relative content = % of total absorbance of all peaks.
total area were identified in the ethyl acetate extract, which largely consisted of terpenes and their oxo-derivatives. Phytol (23.42%), 1,2,4-tripropylbenzene (13.09%), 9-cedranone (12.75%), and ledene oxide (7.22%) were the major compounds identified in the ethyl acetate extract of *M. convoluta*, with smaller amounts of caryophyllene (1.82%), and caryophyllene oxide (1.15%).

**HPLC analysis**

HPLC analysis result showed that there were no flavonoids in ethyl acetate extract and petroleum ether extract. As reported in a previous study by our group (17), there were many flavonoids in the n-butanol extract. Some chemical constituents of these extracts could not be identified by GC-MS or HPLC.

**Discussion**

Hepatocellular carcinoma is one of the most common malignancies worldwide. The high incidence of liver cancer has been attributed to factors such as persistent infection with hepatitis virus and contact with hepatocarcinogens such as nitrosamines and aflatoxins (25). Because of the multifocal nature of liver carcinoma, most cancer patients are considered non-resectable at presentation. In these patients, chemotherapy is the only choice of treatment. Unfortunately, development of drug resistance in the tumor after treatment is always a major obstacle to the successful management of liver cancer (26).

Non-small cell lung cancers commonly develop resistance to radiation and chemotherapy, and they often cannot be treated by surgical resection. Since current treatment modalities are inadequate, novel therapies are necessary to reduce the effects of the increasing incidence of pulmonary neoplasms (27). Thus, developing new therapeutic agents that can overcome drug resistance is an urgent need for cancer patients.

The leading cause of death in China is cancer, followed by stroke. Conventional western cancer therapies, such as chemotherapy, radiation, and surgery, have been increasingly used since the 1960’s in Chinese hospitals. However, the side effects of these treatments have been often highly debilitating both in western countries and in China. The management of cancer with traditional Chinese medicine not only can effectively kill cancer cells, but also has no toxic or side effects on the normal cells of the body. It has no side effect in terms of bone marrow immunosuppression, and has no serious influence on the digestive system. Thus, it could be called “green therapy”.

Caryophyllene has shown anti-inflammatory activity in several animal models, including carrageenan- and PGE-induced hind paw edema, which does not require the integrity of adrenal glands (28). ß-caryophyllene seems to have gastric cytoprotective effects in rats (29). Ghelardini et al. (30) found that ß-caryophyllene has a strong local anesthetic action that appears to be strictly dependent on its chemical structure since the oxidized derivative ß-caryophyllene oxide is devoid of this effect. Caryophyllene oxide, an oxygenated terpenoid, well known as a preservative of food, drugs and cosmetics, has been tested in vitro as an antifungal agent against dermatophytes (31). Caryophyllene oxide can reduce rested-state contractions in the presence of isoprenaline which are thought to be due to the Ca2+ current [I(Ca)] and can inhibit the I(Ca) in single cells from the guinea-pig ventricle in a concentration-dependent and reversible manner. ß-caryophyllene oxide strongly inhibited the potassium current (32) and α-caryophyllene oxide showed strong antimalarial (33) as well as anti-inflammatory activity (34). Caryophyllene oxide has also shown in vitro anti-platelet aggregation activity (35). Sibanda et al. (36) have found caryophyllene oxide to exhibit a modest cytotoxic activity (IC50 values ranging from 32.48 to 77.51 µg/mL) on a panel of human
tumor cell lines, consistent with an earlier report by Kubo et al. (37). But ethyl acetate extract inhibits liver carcinoma cells with IC\textsubscript{50} value of 30 µg/mL. So ethyl acetate extract is more potent than caryophyllene oxide. Caryophyllene oxide, therefore, may be responsible for the cytotoxicity of the ethyl acetate extract against human non-small cell lung carcinoma cell line H1299 and human liver carcinoma cell line HepG2.

The biological activities reported in the literature for the major components of the ethyl acetate extract of \textit{Marchantia convoluta}, i.e., phytol, caryophyllene and caryophyllene oxide, are consistent with the traditional uses of this plant to treat skin tumefaction, to protect the liver and to treat hepatitis B, and as an anticancer drug. The present study indicates that the ethyl acetate extract possesses greater cytotoxicity against HepG2 cells than H1299 cells. Further studies are needed regarding the identification of active compounds and their mechanism of action.

References
