Abstract

The in vitro ability of Pothomorphe umbellata ethanolic crude extract to inhibit matrix metalloproteinase (MMP) in normal cornea and in cornea after alkali injury was demonstrated. Corneas of albino rabbits were injured with 1 N NaOH for 20 s. After 48 h the corneas were excised, homogenized and analyzed for MMP-9 (92 kDa), pro-MMP-2 (72 kDa) and MMP-2 (67 kDa) activity by gelatin zymography. The activity was also measured in untreated corneas. After electrophoresis of 20 µg protein, gels were incubated with 50, 100, or 250 µg/mL lyophilized hydroethanolic (1:1) root crude extract of P. umbellata standardized for 4-nerolidylcatechol (7.09%). The activity of the enzymes was compared with that of untreated gel. At 48 h after injury, the activity of all MMPs was increased compared with untreated eyes. When the gels were incubated with P. umbellata extract the activity of MMP-2, pro-MMP-2 and MMP-9 decreased in a dose-dependent manner. MMP-9 activity decreased by approximately 50% after incubation with 50 µg/mL and was completely abolished at 100 and 250 µg/mL of the extract. After incubation with 50 µg/mL the activity of pro-MMP-2 and MMP-2 also decreased by 50%. The activity of pro-MMP-2 was almost completely abolished after incubation with 250 µg/mL of the extract. For MMP-2 the incubation with 100 or 250 µg/mL of the extract of P. umbellata promoted a 10-fold decrease in activity. In conclusion, P. umbellata root crude extract can be useful as an alternative therapy to control MMP activity after corneal injury.
to an increase in MMP activity (3). Treatment of alkali-burned corneas with MMP inhibitors improves basement membrane integrity (4) and promotes corneal wound healing with decreased inflammation (5). Thus, therapeutic strategies that modulate corneal MMP expression may prevent and/or arrest the stromal ulceration that often follows inflammation-mediated injury (6).

_Pothomorphe umbellata_ L. Miq. (“pariparoba”), a plant of the pepper family Piperaceae, is widely used in Brazilian folk medicine for the treatment of liver diseases and healing of skin wounds. The crude ethanolic root extract of this plant is able to inhibit skin MMP-2 and MMP-9 in vitro and to modulate in vivo the extracellular matrix alterations that follow UVB exposure by inhibiting constitutive MMP-9 activity (7). The objective of the present study was to evaluate the in vitro capacity of _P. umbellata_ crude extract to inhibit cornea MMPs after alkali injury.

The study was conducted on 4 New Zealand rabbits (one untreated animal and three submitted to corneal injury). Corneal injury was induced in both eyes under general anesthesia by intramuscular injection of 10 mg/kg ketamine and 2 mg/kg xylazine with a 5-mm diameter Whatman 42 filter paper soaked in 1 N NaOH for 20 s. Normal corneas were included to evaluate the basal activity of MMPs. After 48 h the animals were euthanized with an overdose of sodium thiopental and both corneas excised under the microscope. Each individual cornea was homogenized on ice at 20,000 rpm for 20 s in 50 mM Tris-HCl buffer, pH 7.4, containing 0.2 M NaCl, 5 mM CaCl₂, and 0.1% Triton X-100, using an Ultra-Turax blender (Marconi, Campinas, SP, Brazil). The homogenate was centrifuged at 8000 g at 4°C for 20 min. Zymography was performed with the supernatant in a gel containing 10% acrylamide and 0.03% bisacrylamide copolymerized with 0.5% gelatin. Electrophoresis was performed at 100 V for 1.5 h. After electrophoresis (20 µg protein) gels were incubated with 50, 100, or 250 µg/mL freeze-dried ethanol:water (1:1) root crude extract of _P. umbellata_. The gel was stained at room temperature for 30 min in a 0.5% solution of Coomassie brilliant blue R-250 containing 25% methanol and 10% acetic acid and destained with 10% acetic acid in water. Gelatinase activity in the cornea extract was detected as unstained zones of degraded gelatin. Each MMP band was quantified densitometrically by computer imaging analysis with a GS-700 densitometer and molecular analysis software, version 1.4 (Bio-Rad, Hercules, CA, USA) (7). _P. umbellata_ crude extract was prepared as previously described (8). Briefly, plant material of _P. umbellata_ was obtained from the medicinal herb garden of the Faculdade de Ciências Farmacêuticas, Universidade de São Paulo, and a sample was deposited in the Herbarium of the Instituto de Biociências, Universidade de São Paulo (Silva/Ropke 01) and identified by Prof. José Rubens Pirani (Institute of Biology, University of São Paulo). The roots were dried and ground to a powder. A 100-mL/g amount of the powder was extracted by percolation with ethanol:water (1:1). The recovered filtrate was freeze-dried. The final concentration of 4-NC, a catechol derivative, in the crude extract was 7.09% (w/w), as evaluated by HPLC with electrochemical detection (8).

Experiments were conducted according to the ARVO statement for the use of laboratory animals in Ophthalmic and Vision Research (9).

The data in Figures 1 and 2 indicate a dose-related inhibition of MMPs. MMP-9 activity decreased by approximately 50% after incubation with 50 µg/mL and was completely abolished with 100 and 250 µg/mL of the extract. After incubation with 50 µg/mL the activity of pro-MMP-2 and MMP-2 also decreased by approximately 50%. The activity of pro-MMP-2 was almost completely abolished after incubation with 250
µg/mL of the extract. For MMP-2 the incubation with 100 or 250 µg/mL of ethanolic root crude extract of *P. umbellata* promoted a 10-fold decrease in activity.

Many substances have been shown to inhibit gelatinase activity, such as ethylenediaminetetraacetic acid (EDTA), doxycycline, N-acetylcysteine, and autoserum. *In vivo* only EDTA was able to appreciably inhibit gelatinase activity present on normal canine corneal surface (10). This activity is attributed to chelation of Zn, a metal required for MMP activity, by EDTA. A major component of *P. umbellata* root extract, 4-NC, exhibits potent antioxidant activity (11) and is also able to inhibit skin MMP-2 and MMP-9 *in vitro* (7). Grimm et al. (12) determined the effects of maritime pine bark extract and its two major metabolites on the activity of MMPs. These metabolites (catechols) with a structure closely similar to that of the active compound in *P. umbellata*, 4-NC, interact directly with the active site of MMP-9, irreversibly inhibiting the enzyme. Green tea (*Camellia sinensis*) extract and epigallocatechin-3-gallate, a polyphenol constituent of green tea, also directly inhibit the expression of MMP-9 and MMP-2 (13,14). Synthetic MMP inhibitors such as Ilomastat are able to suppress the degradation of the epithelial basement membrane of cornea in an experimental alkali-burned model (15). The present results show that, in addition to skin MMP, corneal MMP can also be inhibited by *P. umbellata* extract, suggesting that this plant may be a candidate for the therapy of alkali-burned cornea.

**Figure 1.** *In vitro* effect of a crude extract of *Pothomorphe umbellata* on rabbit corneal metalloproteinase (MMP) activity in alkali-treated eyes. Absorbance of zymograms of cornea homogenates of alkali-treated eyes incubated in the presence or absence of 50, 100, or 250 µg/mL *P. umbellata* crude ethanolic root extract. The height of the bars indicate the intensity of the bands obtained from gelatin zymography by densitometry. Data are reported as the mean ± SD for six independent experiments.

**Figure 2.** Zymogram of corneal homogenate incubated with increasing concentrations (µg/mL) of *Pothomorphe umbellata* extract. C = untreated cornea; L = alkali-treated cornea; MMP = metalloproteinase.

### References


