Abstract

The functional effect of the A>G transition at position 2756 on the MTR gene (5-methyltetrahydrofolate-homocysteine methyltransferase), involved in folate metabolism, may be a risk factor for head and neck squamous cell carcinoma (HNSCC). The frequency of MTR A2756G (rs1805087) polymorphism was compared between HNSCC patients and individuals without history of neoplasias. The association of this polymorphism with clinical histopathological parameters was evaluated. A total of 705 individuals were included in the study. The polymerase chain reaction-restriction fragment length polymorphism technique was used to genotype the polymorphism. For statistical analysis, the chi-square test (univariate analysis) was used for comparisons between groups and multiple logistic regression (multivariate analysis) was used for interactions between the polymorphism and risk factors and clinical histopathological parameters. Using univariate analysis, the results did not show significant differences in allelic or genotypic distributions. Multivariable analysis showed that tobacco and alcohol consumption (P < 0.05), AG genotype (P = 0.019) and G allele (P = 0.028) may be predictors of the disease and a higher frequency of the G polymorphic allele was detected in men with HNSCC compared to male controls (P = 0.008). The analysis of polymorphism regarding clinical histopathological parameters did not show any association with the primary site, aggressiveness, lymph node involvement or extension of the tumor. In conclusion, our data provide evidence that supports an association between the polymorphism and the risk of HNSCC.

Key words: Head and neck cancer; Polymorphism; Folate metabolism; MTR gene

Introduction

Head and neck squamous cell carcinomas (HNSCC) include malignant tumors of any site in the upper aerodigestive tract, occurring in 95% of cases and being the fifth most common cancer worldwide and the most common histological type (1). Smoking and alcohol consumption, viral infections, especially the papilloma virus subtypes 16 and 18, and vitamin and micronutrient deficiencies, including deficits of folate, vitamins A, C, and E, selenium and zinc, are risk factors for this disease (1-6).

Folate metabolism plays an important role in carcinogenesis because of its involvement in both DNA methylation and nucleotide synthesis. DNA methylation is the transfer of methyl groups (CH3) to the C5 position of cytosine residues located in cytosine-guanine dinucleotides, by reactions catalyzed by proteins called DNA methyltransferases (7). This epigenetic modification of the DNA has several functional roles including control of gene expression (8), maintenance of genomic stability (9) and stability of the chromatin structure (10).

The relationship between polymorphisms of genes involved in folate metabolism and the risk for cancer is related to their effect on DNA methylation and synthesis and, thus, on the maintenance of chromatin structure and chromosome stability (6).

The 5-methyltetrahydrofolate-homocysteine methyltransferase (MTR) gene, located in chromosome 1q43 (10), encodes the methionine synthase enzyme, which has a role in folate metabolism, catalyzing the remethylation of

Correspondence: E.M. Goloni-Bertollo, Departamento de Biologia Molecular, FAMERP, Av. Brigadeiro Faria Lima, 5416, 15090-000 São José do Rio Preto, SP, Brasil. E-mail: eny.goloni@famerp.br

homocysteine (Hcy) to methionine, a reaction essential to adequately maintain normal methionine and intracellular Hcy concentrations (11). An adenine to guanine transition at position 2756 (rs1805087) of the MTR gene results in the substitution of the amino acid, aspartic acid, with glycine in codon 919 of the protein and is related to alterations in the folate metabolic pathway, thus possibly influencing the risk of cancer (12,13).

Research results on the influence of this polymorphism on the development of cancer are contradictory. Some studies have shown associations with colon (14), lung (15) and breast cancer (16), and relationships with tumor aggressiveness and variable responses to esophageal cancer treatment (17). Other studies did not confirm associations between the polymorphism and different types of cancer, including colorectal (18), lung (19), bladder (20), and stomach cancer (21).

Three studies investigated the MTR A2756G polymorphism with respect to head and neck cancer. Zhang et al. (22), when analyzing the MTR A2756G polymorphism in 721 cancer patients and 1234 control individuals, observed differences in the genotype frequencies between the two groups. They also showed that the 2756AG genotype was associated with a significantly increased risk of HNSCC among younger subjects, women and former drinkers, in particular tumors of the oral cavity. In another study, Suzuki et al. (23) analyzed 237 HNSCC patients and 711 controls in Japan but did not identify differences between the two groups. Kruszyna et al. (24) analyzed 131 men diagnosed with squamous cell carcinoma of the larynx and 250 randomly selected, unrelated healthy male blood donors and other healthy volunteers in the Polish population and suggested that the MTR 2756G allele may contribute to the risk of laryngeal cancer.

Based on the above evidence, the aim of this study was to compare the frequencies of the MTR A2756G polymorphism between head and neck cancer patients and individuals with no history of cancer, and to determine if there were an association of this polymorphism with clinical histopathological parameters.

Patients and Methods

A total of 705 individuals (236 patients and 469 controls) with a mean age of 52.5 ± 13.7 years were included in the study.

The study protocol was approved by the National Ethics Committee (SISNEP 0976.0.140.000-05). The case group consisted of 236 patients who were diagnosed with head and neck cancer at Hospital de Base, São José do Rio Preto, SP, Brazil. Informed consent was obtained from all individuals enrolled in the study. The diagnosis was made from pathological specimens after total excision or a biopsy. The inclusion criterion was squamous cell carcinoma tumor cell types and the exclusion criterion was patients previously treated for tumors. The primary anatomic sites were oral cavity (N = 92, 40%), pharynx (N = 59, 25%), and larynx (N = 76, 32.2%) and 9 (2.8%) patients with unknown primary site of the tumor.

All required information about clinical histopathological parameters was obtained from the patients’ medical records. The average survival was 31.7 ± 27.1 months. The treatment options for the patients were surgery, radiotherapy and chemotherapy.

The tumors were classified according to the parameters of the International Union of Cancer Control (UICC), 2002, and the American Joint Committee for Cancer (AJCC), 2002, based on three criteria: extension of the tumor (T), presence of regional lymph node involvement (N), and presence of metastasis at a distance (M) (25). The clinical stage (TNM) was used to analyze aggressiveness, with tumors being grouped as non-aggressive (stages I and II) and aggressive (stages III and IV).

The control group consisted of 469 Brazilian blood donors without a diagnosis of cancer according to government guidelines for donated blood that is tested for 20 related diseases (http://www.hemonline.com.br/portarias/rdc153/indexframe.htm). The inclusion criterion was age of more than 40 years and the exclusion criterion was a family history of cancer. Each eligible subject was interviewed to obtain data on age, gender, smoking habit, use of alcohol, and family history of cancer. Patients and controls were followed up for 60 months.

The variables analyzed were gender, exposure to risk factors (tobacco and alcohol consumption), primary site of occurrence, aggressiveness, extension of the tumor, and lymph node involvement. Individuals who had smoked more than 100 cigarettes in their lifetime and at the time of the interview continued to smoke either every day or at least on some days were considered to be smokers. Individuals who drank 4 doses of alcohol per week were considered to be alcohol consumers (26,27).

Genomic DNA was obtained from peripheral blood according to technique of Miller et al. (28). The polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) technique was used to determine genotypes of the MTR A2756G polymorphism (rs1805087). The primers used were: sense 5’-CCA GGG TGC GAC GTA TAC TG-3’ and anti-sense 5’-GCC TTT TAC ACT CCT CAA AAC C-3’. Amplification was obtained with initial denaturation at 94°C for 4 min, followed by 30 cycles of 1 min of DNA denaturation at 94°C, 1-min primer annealing at 56°C, and 1-min extension at 72°C. A final extension of 10 min at 72°C was carried out. The product of 498 bp was submitted to digestion with the HaeIII restriction enzyme for 2 h at 37°C according to manufacturer guidelines. Fragments of 390, 123, and 85 bp were generated when the G polymorphic allele was present and 413- and 85-bp fragments were observed with the A allele.
Statistical analysis

Groups were compared by the chi-square test (univariate analysis). Utilizing the BioEstat computer program, this test was also used to analyze Hardy-Weinberg equilibrium. Multiple logistic regression models were used to determine the interaction effect between the genetic polymorphisms and variables related to HNSCC. One model included gender (reference: female), smoking (reference: non-smokers) and drinking habits (reference: non-drinkers) using the Minitab for Windows computer program (Version 12.22). P < 0.05 was considered to be statistically significant. Results are shown as odds ratio (OR) and 95% confidence intervals (95%CI).

The clinical histopathological parameters were analyzed by multiple logistic regression. Tumors were classified as low T (T1, T2) and high T (T3, T4). The N classification was dichotomized into no lymph node involvement (N0) and involvement (N1, N2, N3). Tumors were divided into early stage (stages I and II) and advanced stage (stages III and IV) categories.

The Kaplan-Meier method was used to evaluate survival rates and time of disease recurrence. The log-rank test was used to assess differences related to the different genotypes.

Results

There were statistically significant differences between patients and controls regarding gender, alcohol consumption and smoking (P < 0.05). Five hundred and twenty-nine (75%) participants were men (203 patients and 326 controls) and 176 (25%) were women (33 patients and 143 controls). Of the cases, 77.97% consumed alcohol compared to 46.70% of the controls. Smoking also differed greatly between cases and controls, with 77.97% of cases smoking compared to 46.70% of the controls. However, when compared to controls, the difference was not statistically significant (P = 0.335) was observed for the women.

Table 1. Distribution of demographic data, risk factors, genotypes, MTR 2756 alleles, and odds ratio (OR) for head and neck cancer.

<table>
<thead>
<tr>
<th>Variables</th>
<th>N (%)</th>
<th>OR (95%CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tobacco consumption</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-smokers</td>
<td>301 (37.3)</td>
<td>Reference</td>
</tr>
<tr>
<td>Smokers</td>
<td>504 (62.7)</td>
<td>4.49 (2.68-7.54)*</td>
</tr>
<tr>
<td>Alcohol consumption</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alcohol non-consumers</td>
<td>338 (42)</td>
<td>Reference</td>
</tr>
<tr>
<td>Alcohol consumers</td>
<td>467 (58)</td>
<td>2.30 (1.46-3.63)*</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>263 (32.6)</td>
<td>Reference</td>
</tr>
<tr>
<td>Male</td>
<td>595 (70.4)</td>
<td>1.18 (0.69-2.01)</td>
</tr>
<tr>
<td>Age</td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;42 years</td>
<td>217 (27)</td>
<td>Reference</td>
</tr>
<tr>
<td>42-51 years</td>
<td>185 (23)</td>
<td>4.72 (2.29-9.72)*</td>
</tr>
<tr>
<td>52-63 years</td>
<td>191 (23.7)</td>
<td>19.45 (9.55-39.59)*</td>
</tr>
<tr>
<td>&gt;63 years</td>
<td>212 (26.3)</td>
<td>11.01 (5.33-22.72)*</td>
</tr>
<tr>
<td>MTR 2756 genotypes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AA</td>
<td>523 (65)</td>
<td>Reference</td>
</tr>
<tr>
<td>AG</td>
<td>247 (30.6)</td>
<td>1.69 (1.09-2.62)*</td>
</tr>
<tr>
<td>GG</td>
<td>35 (4.4)</td>
<td>1.08 (0.37-3.13)</td>
</tr>
<tr>
<td>MTR 2756 alleles</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>682 (76.4)</td>
<td>Reference</td>
</tr>
<tr>
<td>G</td>
<td>210 (23.6)</td>
<td>1.60 (1.05-2.44)*</td>
</tr>
</tbody>
</table>

MTR = 5-methyltetrahydrofolate-homocysteine methyltransferase. *P < 0.05 (multiple logistic regression).

Statistics

For the MTR 2756 polymorphisms, AA, AG, and GG genotype frequencies were 65.7, 31.0, and 3.3%, respectively, for the cases, and 72.5, 24.0, and 3.5%, respectively, for the controls. Smoking also differed greatly between cases and controls, with 77.97% of cases smoking compared to 46.70% of the controls. However, when compared to controls, the difference was not statistically significant (P = 0.335) was observed for the women.

As matching demographic data and risk factors between patients with cancer and control individuals was not possible, multivariable analysis was performed to adjust these variables. The following variables were included: gender, tobacco and alcohol consumption, and MTR A2756G polymorphism. Smoking (P < 0.05), alcohol consumption (P < 0.05), AG heterozygous genotype (P = 0.019), and the G polymorphic allele (P = 0.028) were predictors of the disease (Table 1).

Analysis of polymorphisms and clinical parameters did not detect any association with the primary tumor site (P = 0.12), tumor extension (P = 0.38), lymph node involvement (P = 0.35), or tumor stage (P = 0.18).

The Kaplan-Meier survival curves by genotype are presented in Figure 1A, and did not demonstrate any association between polymorphisms and overall survival (P = 0.52). Moreover, no association was observed for the time of recurrence of the disease (P = 0.25; Figure 1B).

Discussion

A review of published data showed that the most important factors predisposing factors to the development of HNSCC, present in 90% of cases, are alcohol and tobacco consumption (1) and our study confirms this association. Additionally, a significant association was confirmed between
gender and this disease, as previously reported by Argiris et al. (29), who showed that males are the most affected by this disease.

Using univariate analysis, the results of the current study did not demonstrate significant differences between groups with respect to allele and genotype distributions, similar to the data reported by Suzuki et al. (23). The results also show that men with head and neck cancer had a higher frequency of the G polymorphic allele than men with no history of cancer, thus supporting the results of Kruszyna et al. (24), who showed that the frequency of the G allele was 1.55 times higher in male patients with laryngeal cancer compared to male controls. Our findings did not confirm those of Zhang and et al. (22), who reported that the MTR 2756AG genotype is associated with an increased risk for this type of cancer, especially in younger individuals, women and former smokers.

Multivariable analysis performed after adjusting for age, gender and tobacco and alcohol consumption showed that the G polymorphic allele and the AG heterozygous genotype are associated with increased risk for the disease with OR of 1.60 and 1.69, respectively. Associations between MTR A2756G polymorphism and the development of some types of cancer have been shown in previous studies (30,31).

Only three studies have investigated this polymorphism regarding head and neck cancer (22-24). Zhang et al. (22) and Kruszyna et al. (24) reported differences in the MTR A2756G genotype frequencies between head and neck cancer patients and controls and between larynx cancer patients and controls, respectively, while Suzuki et al. (23) did not observe any association between the presence of the polymorphism and the risk for the disease. The latter investigators demonstrated an interaction of alcohol consumption with the 2756GG genotype in the risk for head and neck cancer.

Folate is an essential nutrient, which has important roles in the synthesis (genetics), repair, and methylation (epigenetics) of DNA. It is used to transport a methyl group, which is essential for the de novo synthesis of deoxynucleoside triphosphate and methionine.

The functional variant, MTRA2756G, encoding the MTR enzyme (OMIM 156570) uses methyl-tetrahydrofolate as a methyl donor for the remethylation of Hcy to form methionine. Methionine adenosyltransferase, using methionine and ATP, induces the formation of S-adenosylmethionine (32).

Folate deficiency associated with the MTR GG or MTR AG genotypes may increase cancerogenesis by reducing the formation of S-adenosylmethionine, leading to DNA hypomethylation and cancer and may also induce uracil misincorporation into DNA in place of thymine during DNA replication, which, along with DNA hypomethylation, results in impaired DNA repair, DNA strand breakage, and chromosome damage (33). The study of Paz and collaborators (34) showed that hypomethylation of genomic DNA may be associated with the MTR A2756G polymorphism.

Studies on other types of cancer also found a link between the MTR GG genotype and extensive genomic hypomethylation as well as decreased promoter hypermethylation of tumor suppressor genes (35,36).

In the present study, Hcy concentration was not measured but several studies have shown that the MTR A2756G polymorphism has also been associated with variations in Hcy concentrations. However, the quantitative impact of this polymorphism on cellular Hcy concentrations has not been determined. Studies have shown an association between the MTR GG genotype and lower plasma Hcy levels compared to the MTR AA genotype, showing that the MTR A2756G polymorphism contributes to enhanced methionine synthesis and an increase in intracellular S-adenosylmethionine concentration (13,37). However, Laraqui et al. (38) observed a correlation between the polymorphic allele (G) and moderately high Hcy levels.
Our study did not detect any associations of the MTR A2756G polymorphism with the site of occurrence, aggressiveness, tumor extension, or lymph node involvement. Zhang et al. (22) showed an increased frequency of the polymorphism related to tumors of the oral cavity, while Kruzsyna et al. (24) did not find any association of this polymorphism with laryngeal cancer. After an exhaustive literature review, no previous studies were found regarding the possible association between this polymorphism and clinical histopathological parameters or with the risk of cancer development.

Marchal et al. (39) reported that the presence of the MTR 2756 G allele is a factor of tumor aggressiveness in prostate cancer. No data were found in the literature assessing survival according to MTR A2756G polymorphism in patients with head and neck cancer.

In conclusion, although there is an association between MTR A2756G polymorphism and the risk for head and neck cancer and a higher frequency of carriers of the G allele variant in men with head and neck cancer compared to men without any history of cancer, studies of other enzymes involved in folate metabolism, their plasma concentrations and other derivatives could contribute to a better understanding of the factors involved in the etiology of head and neck cancer.

Acknowledgments

The authors wish to thank all those participating in this study, Prof. Adília M. Pires Sciarra for her help in writing the text, Prof. Dr. José A. Cordeiro and Prof. Dr. Moacir F. Godoy for their help in statistical analysis and FAMERP/FUNFARME. Research supported by FAPESP (#04/14573-3, CAPES and CNPq (#477665/2004-7, #305462/2005-9).

References


33. Ames BN. DNA damage from micronutrient deficiencies is likely to be a major cause of cancer. *Mutat Res* 2001; 475: 7-20.


