New species of Temnocephala (Platyhelminthes, Temnocephalida) ectosymbiont on vulnerable species of aeglids (Crustacea, Anomura) from the Neotropical Region

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Abstract: A new species of the genus Temnocephala Blanchard, 1849 from southern Brazil was found on two species of anomuran crustaceans, Aegla spinipalma Bond-Buckup & Buckup, 1994 and Aegla grisella Bond-Buckup & Buckup, 1994, the latter classified as a vulnerable species by the “Lista de Referência da Fauna Ameaçada de Extinção no Rio Grande do Sul. Decreto no 41.672, de 11 junho de 2002”. The crustaceans were collected from a tributary creek of the Forqueta river, Perau de Janeiro, Arvorezinha and a tributary creek of the Fão river, Pouso Novo, Rio Grande do Sul, Brazil; both localities belong to the Sub-Basin of Forqueta River. The new species differs from seven other temnocephalans epibionts on Aegla Leach, 1820, by having the following characters: 1. a long and slightly curved cirrus, 2. two vaginal sphincters, one proximal, big and asymmetric, and one distal, smaller and symmetric, and; 3. longer than wide, elongated epidermal ‘excretory’ syncytial plates (EPs), with a almost horizontally central excretory pore, displaced to the anterior portion of the plate. The new species' EP is the largest in total length among epibionts temnocephalans in crustaceans already registered. Regarding the similarities with the male reproductive system of Temnocephala axenos Monticelli, 1898, the new species has important differences in the female reproductive system. It has a larger proximal vaginal sphincter, located in the middle of the vagina, while the smaller distal one is at the extreme end of the organ. Besides that, the vaginal portion between the proximal and distal sphincters is conspicuous, with a strong muscular wall. This is the first record of a species of Temnocephala in the Taquari Valley, as well in the ‘Perau de Janeiro’, which is an area with a rich endemic fauna.

Keywords: crustacean, ectosymbiont, South America, symbiosis, Taxonomy.

Nova espécie de Temnocephala (Platyhelminthes, Temnocephalida) ectosimbionte sobre espécies vulneráveis de eglídeos (Crustacea, Anomura) da Região Neotropical


Palavras-chave: América do Sul, crustáceos, ectosimbionte, simbiose, Taxonomia.
**Introduction**

The first host taxon of temnocephalans was recorded on Crustacea Brünnich, 1772. This group also has the largest number of epibiont species of the genus *Temnocephala* Blanchard, 1849. From the 21 species of the genus *Brünnich*, 1772. This group also has the largest number of epibiont species of the genus *Aegla* Leach, 1820: *Temnocephala chiensis* (Moquin-Tandon, 1846), *Temnocephala axenos* Monticelli, 1898, *Temnocephala mexicana* Vayssière, 1898, *Temnocephala talicei* Dioni, 1967, *Temnocephala cyanoglandula* Amato, Amato & Daudt, 2003, *Temnocephala mertoni* Volonterio, 2007, and *Temnocephala dionii* Ponce de León, Berón Vera & Volonterio, 2015.

*Temnocephala chilensis* was the first species of the genus to be described and was recorded consistently after that (Dioni 1967a, Damborenea 1992). However, the authors have not updated the species description using more recent techniques.

Dioni (1972) recorded *T. mexicana* in *Aegla* sp. and *Parastacus* sp. from Argentina. The species was described by Vayssière (1898) and re-described by Lamothe-Argumedo (1968). Both publications were based on specimens collected from *Procambarus dieri* (Bouvier, 1897) and *Pseudotherelphusa jouyi* Rathbun, 1893 (added by Lamothe-Argumedo in 1968) from Mexico.

*Temnocephala talicei* has also been recorded a few times (Dioni 1968, Damborenea 1992, et al. 1997), being subsequently re-described and having a neotype designated by Volonterio (2009).

*Temnocephala axenos* is the most well-studied species (Baer 1931, Dioni 1967b, 1968, Damborenea 1992, et al. 1997), but it has substantial taxonomic problems, such as a misidentification of the host type (Amato et al. 2003) and the loss of the holotype at the Berlin Natural History Museum because of war damage. The superficial description of *T. axenos* (Monticelli 1898) lead to the consideration of this species as a senior synonym of *Temnocephala brasiliensis* Merton, 1922 by Baer (1931) and *Temnocephala bresslaui* Pérez-González, 1949 by Dioni (1967c). Volonterio (2007) stated that *T. bresslaui* was erroneously synonymized by Dioni (1967c) and it is, probably, still a valid species. The author re-described *T. axenos*, solving some of these issues. The incomplete description of *T. cyanoglandula*, with only data of the male reproductive system, has led Volonterio (2007) to suggest a synonymy for this species with *T. axenos* or *T. bresslaui*. However, a recent study of the female reproductive system (Seixas et al. 2015a) has confirmed *T. cyanoglandula* as a valid species.

While describing *T. mertoni*, an epibiont species on anomuran crabs, Volonterio (2007) pointed out the difficulties of distinguishing temnocephalans species on crustaceans given the similarities in the males’ reproductive system. Among other features, the author highlighted the importance of describing in detail the female reproductive system of ectsosymbionts hosted by crustaceans.

*Temnocephala dionii* was the last species described as an ectsosymbiont on *Aegla neauquensis* Schmitt, 1942 from Argentina (Ponce de León et al. 2015).

There are no records of temnocephalans at the Forqueta River Sub-Basin (Fig. 1), where the crustaceans fauna is less well investigated. The Forqueta and Fão Rivers, localized at the municipalities of Arvorezinha and Pouso Novo, respectively, represent the two main rivers of the sub-basin. The present study aims to describe a new species of *Temnocephala* ectsosymbiont on *Aegla spinipalma* Bond-Buckup & Buckup, 1994 and *Aegla grisella* Bond-Buckup & Buckup, 1994 (Fig. 2a), which is classified as a vulnerable species by Rio Grande do Sul State Law (Marques et al. 2002). Both are being registered as new host species for neotropical temnocephalans.

**Material and Methods**

One hundred and six specimens of *A. grisella* were collected from a tributary creek of the Forqueta river (28°51’9.85"S; 52°17’55.02"W), Peraú de Janeiro, Arvorezinha, Rio Grande do Sul, Brazil; and eighty-two specimens of *A. spinipalma* were collected from a tributary creek of the Fão river (29°12’2.81"S; 52°11’31.80"W), Pouso Novo, Rio Grande do Sul, Brazil. Both localities belong to Forqueta River Sub-Basin.

The collections occurred monthly between August 2014 and April 2015 as part of a larger project for taxonomic and ecological studies. All crustaceans were collected with dip nets, sexed, measured, and returned to their natural habitat. Only a few specimens (≥ 10) were taken alive to the “Setor de Evolução e Ecologia, Univates” to be examined for temnocephalans.

The temnocephalans were studied through a series of techniques focusing especially on the morphology of the vagina and other female reproductive organs, as well as the morphology of the cirrus structure and the epidermal ‘excretory’ syncytial plates (EPs).

For general measurements, the helminths were fixed with AFA, under slight cover slip pressure, following the protocols established by Amato et al. (2007) and Seixas et al. (2010). The specimens were stained with Delafield's hematoxylin or aceto-carmine/fast green, cleared in cedar oil, and mounted as permanent slides on Canada balsam.

For the EPs studies, the specimens were dehydrated according to a protocol adapted from Kashi et al. (2014) for Scanning Electron Microscopy (SEM). The SEM preparations and photomicrographs were made at the ‘Laboratório MEV (Microscopia Eletrônica de Varredura)’ at Tecnovates, Univates. The images of the EPs were measured, according to Seixas et al. (2015b), using the AxioVision Zeiss LE 4.7.2 software.

Cirrus measurements were taken from extracted cirri mounted on Faure’s mounting medium (F). The terminology used to describe the male reproductive structures followed Seixas et al. (2010).

Photomicrographs of the temnocephalans were taken with the microscope Zeiss Axiolab. The line drawings and photographic images were prepared using Adobe’s Fireworks® CS6 and Adobe’s Photoshop® CC 2017. Measurements are in micrometers (µm) unless otherwise indicated, ranges are followed (between parentheses) by the mean, the standard deviation values, and the number of specimens measured for a given character (when different than 25). The ecological concepts applied to the symbiotic organisms follow Bush et al. (1997).

The whole mounts of adult and juvenile specimens, as well as slides containing individual cirri mounted on F were deposited in the following scientific collections: 1. ‘Coleção Helmintológica do Instituto Oswaldo Cruz (CHIOC)’, Rio de Janeiro, RJ, Brazil; 2. ‘Colección de Invertebrados, División Zoología Invertebrados, Museo de La Plata (MLP)’, La Plata, Argentina; and 3. ‘Colección de Invertebrados do Instituto Nacional de Pesquisas da Amazônia (INPA)’, Manaus, AM, Brazil. Some host specimens were deposited in the ‘Coleção de Crustáceos, Departamento de Zoologia, UFRGS’, Porto Alegre, RS, Brazil, and ‘Coleção Zoológica, Museu de Ciências Naturais da Univates (MCN/UNIVATES)’, Lajeado, RS, Brazil. The remaining
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Figure 1. Map of Rio Grande do Sul showing the Forqueta River Sub-Basin and collection points in the municipalities of Arvorezinha and Pouso Novo.
specimens are kept in the laboratory for completion of the other studies. All material will be deposited at the ‘Coleção Zoológica, Museu de Ciências Naturais da Univates (MCN/UNIVATES)’ upon conclusion of these studies.

Results

Description. Based on 63 temnocephalans specimens collected from *A. grisella* and 34 specimens from *A. spinipalma*: 13 whole mounted adults from *A. grisella*, 12 whole mounted adults from *A. spinipalma*, 3 dissected cirri from *A. grisella*, and 2 dissected cirri from *A. spinipalma* measured.

External characteristics. Body (without tentacles) (Figs 3a and 8d) 1.42–4.05 mm (2.65 mm ± 700) long, 1.05–2.39 mm (1.69 mm ± 360) wide; adhesive disk ventral, subterminal, partially covered by the body (Fig. 3a) 276–790 (502 ± 116) long, 434–889 (597 ± 127) wide; disc peduncle 217–632 (424 ± 111) wide. Eyespots with red pigmentation (observations made on live specimens). Two EPs longer than wide (Figs 5 and 6) 347.5–447.5 (397.5 ± 58; 4) total length, 110–132.5 (121 ± 13; 4) total width; length of the anterior portion of the EP from the excretory pore 117–158 (138 ± 24; 4), length of the posterior portion of the EP from the excretory pore 230–289 (260 ± 34; 4); width of the external limit of the EP from the excretory pore 62–69 (65 ± 4; 4); width of the internal limit of the EP from the excretory pore 49–64 (56 ± 4; 4). The excretory pore is almost central horizontally, but displaced to the anterior portion of the plate. Seventeen percent of the total length of the EP is beyond the limit of the tentacles with the body. Ratio of total length of the EPs/total body length (without tentacles): 6.7: 1.
Figure 3. *Temnocephala grisella* sp. nov. (3a) Diagram of an adult specimen showing adhesive disk (ad), disc glands (asterisks), anterior testes (at), cyanophilous glands (cg), excretory vesicle (ev), paranephrocytes (head arrows), Haswell glands (hg), intestine (i), mouth (m), pharynx (ph), posterior testis (pt), rhabditogenic glands (rg), tentacles (t), and vitelline glands (vg). Scale bar = 500 μm. (3b) Cirrus, showing the proximal limit of the introvert (arrow). Scale bar = 20 μm.

Glands. Rhabditogenic glands (Figs 3a and 8d) forming bunches (average 92 cells) extending from the level of the Haswell glands to the end of the posterior testes, in lateral fields of the body, 30-90 (67 ± 15) in diameter, ducts inconspicuous. Grape-like bunches of cyanophilous glands (Figs 3a and 8d) (average 20 cells), located at the level of the excretory vesicles. Two groups of two Haswell glands (Fig. 3a), showing little affinity with hematoxylin/aceto-carmine/fast green, in front of the cerebral transverse band; diameter of largest cell 65–160 (107 ± 30). Disc glands between adhesive disc and genital complex, 30–80 (52 ± 16; 24) in diameter, including two pairs of large, round, more central paranephrocytes, 52.5–160 (107 ± 32; 24) long (Fig. 3a).

Reproductive system. Female. Vitellarium arborescent and thin (Fig. 3a); vagina elongated 65–137 (99 ± 23; 10) total length (Figs 4a, 8b - 8c); divided into two portions, before (BPS) and after (APS) the proximal sphincter. BPS portion 40–87 (60 ± 15; 9) long, 37–75 (48 ± 11; 9) wide, with thin wall (Figs 4a and 8b); APS portion 25–52 (38 ± 9; 13) long, 40–75 (57 ± 10; 13) wide, with strong muscular wall (Figs 4a, 8a - 8b). Ovary 97–242 (145 ± 29; 23) long, 82–172 (113 ± 25; 23) wide, located in the middle of the BPS portion of the vagina (Figs 4a, 8a - 8c). Proximal vaginal sphincter asymmetrical 62–105 (83 ± 13; 16) total diameter (Figs 4a, 8a and 8c), diameter of anterior portion 15–27 (24 ± 4; 14) (Figs 4a and 8b - apvs), diameter of posterior
Figure 4. *Temnocephala grisella* sp. nov. (4a) Female reproductive system, showing: Anterior portion of the distal vaginal sphincter (advs), after proximal sphincter of the vagina (aps), anterior portion of the proximal vaginal sphincter (apvs), before proximal sphincter of the vagina (bps), distal vaginal sphincter (dvs), genital atrium (ga), ovary (o), posterior portion of the distal vaginal sphincter (pdvs), proximal vaginal sphincter (pvs), vagina (va), vitelline duct (vd), vesicula intermedia (vi), and vesicula resorbens (vr). Scale bars = 100 μm. (4b) Male reproductive system, showing: Cirrus (c), deferent vessels (dd), ejaculatory vesicle (ejv), prostatic bulb (pb), prostatic cells (pc), prostatic secretions (ps), and seminal vesicle (sv). Scale bars = 100 μm.

Portion 27–65 (45 ± 14; 14) (Figs 4a and 8b - ppvs); distal vaginal sphincter symmetrical 40–85 (56 ± 13; 16) total diameter (Figs 4a, 8a - 8b), diameter of anterior portion 15–37 (24 ± 6; 16) (Figs 4a and 8c - advs), diameter of posterior portion 15–37 (25 ± 7; 16) (Figs 4a and 8c - pdvs). Vesicula intermedia 35–100 (66 ± 21; 11) long (Figs 4a and 8a); vesicula resorbens usually full of sperm, 60–287 (151 ± 60; 15) long, 100–257 (188 ± 44; 15) wide, wall thickness 2.5–22 (10 ± 8; 8) (Figs 4a and 8a).

Male. Four testes rounded to oblique (Figs 3a and 8d); deferent vessels unite in large, pyriform seminal vesicle 70–245 (146 ± 42) long, 52–125 (85 ± 23) wide, wall thickness 2.5–7.5 (4 ± 2; 20) (Figs 4b and 8a); prostatic bulb short, 145–400 (255 ± 64) long, 57–192 (120 ± 33) wide, wall thickness 2.5–20 (11 ± 5; 19) (Figs 4b and 8a); vesicula intermedia usually full of sperm, 60–287 (151 ± 60; 15) long, 100–257 (188 ± 44; 15) wide, wall thickness 2.5–22 (10 ± 8; 8) (Figs 4a and 8a).

Type locality. Tributary creek of the Forqueta river, Peraú de Janeiro, Arvorezinha, Rio Grande do Sul, Brazil (28°51′9.85″S; 52°17′55.02″W).

Other locality. Tributary creek of the Fão river (29°12′2.81″S; 52°11′31.80″W), Pouso Novo, Rio Grande do Sul, Brazil (29°12′2.81″S; 52°11′31.80″W).

Site. Branchial chambers and body surface; eggs cemented on external surfaces of exoskeleton (Figs 2b-2d).

Prevalence. 92.4%.

Average intensity of infestation. 14.8

Helminth specimens deposited. ‘Coleção Helmintológica do Instituto Oswaldo Cruz’: CHIOC 38212 (HOLOTYPE); CHIOC 38213 (cirrus).

‘Colecção de Invertebrados do Instituto Nacional de Pesquisas da Amazônia’: INPA 663 (paratype); INPA 664 (cirrus). ‘Colecção de Invertebrados, Divisão Zoologia Invertebrados, Museo de La Plata’: MLP-He 7100 (paratype); MLP-He 7101 (cirrus).

Host specimens deposited: ‘Coleção de Crustáceos, Departamento de Zoologia, UFRGS’: 6119 - 6142 (*A. grisella*); ‘Coleção Zoológica, Museu de Ciências Naturais da Univates (MCN/UNIVATES)’: ZAUMCN 1072-1076 (*A. grisella*).

Etymology. The specific epithet *grisella* refers to the type host and act as a reminder of the importance of its preservation.

Remarks. The EPs of the species hosted by crustaceans usually present a great variation in shape, however *T. grisella* sp. nov. presents an EPs’ shape similar to *Temnocephala pignalberiae* Dioni, 1967 (Seixas et al. 2015b). In the male reproductive system, the seminal vesicle has a thin muscular wall (4 μm thick on average), in contrast with a strong
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Figure 5. Scanning Electron Microscopy of Temnocephala grisella sp. nov. showing the highlighted epidermal ‘excretory’ syncytial plate; excretory pore (ep) and limits of the plate (head arrows). Scale bar = 100μm.

muscular wall of the prostatic bulb (11 μm thick on average). The cirrus showed intraspecific variation of the curvature from straight to slightly curve. The introvert portion of the cirrus has a small variation in length (25-30 μm) but the same measure in width at base (15 μm) in all specimens measured. The total length of the cirrus is three times bigger than the maximum width of the shaft at its base. The posterior pair of the testes is two times bigger than the anterior pair.

Discussion

Temnocephala chilensis, T. axenos, T. talicei and T. mertoni present cirrus measuring, on average, between 123-149 μm long (Damborenea & Cannon 2001, Volonterio 2007), while T. cyanoglandula has the largest cirrus among anomuran crabs temnocephalans, having an average length of 256 μm (Amato et al. 2003). Among these species, T. grisella sp. nov. has a cirrus of intermediate size, measuring 179 μm on average (Table 1). Dioni (1967c) studied specimens of T. axenos ectosymbiont on species of Aegla and Parastacus Huxley, 1879 from Uruguay and Brazil, finding a great cirrus’ size variability (125-150 μm), although, due to the lack of data on female reproductive system, is impossible to compare with the new species described in the present work. Nonetheless, both cirri measurements presented by Dioni (1967c) and Volonterio (2007), on their description of Uruguayan specimens of T. axenos, differ from that of T. grisella sp. nov. (Table 1).

Lamothe-Argumedo (1968) also found a great cirrus’ size variability (144-206 μm) while re-describing T. mexicana. Although the range was similar to T. grisella sp. nov., the shape of the cirrus differed. In fact, the cirrus’ shape of T. mexicana presented by the author differs greatly from the original description of the species. Vayssière (1898) describe the cirrus as a little curved, with an “exsertile” portion at its outer end, meaning that it have a portion projected beyond the organ, which is visible in the diagram provided by the author. The cirrus described by Lamothe-Argumedo (1968) doesn’t have this characteristic introvert and was similar to T. mertoni by having a slightly sinuous portion in the shaft. The single vaginal sphincter of T. mexicana, evidenced by the diagram presented, is also similar to T. mertoni. These cirrus and vaginal sphincter characteristics differs T. mexicana and T. mertoni from T. grisella sp. nov.
Damborenea & Cannon (2001), on a neotropical temnocephalans revision, pointed out the absence of any muscular structure (sphincter) in *T. chilensis* vagina. They also recorded a conic cirrus with a swollen introvert measuring an average of 149 µm in length. All these characteristics differs from *T. grisella sp. nov.* The authors also assert the lack of sphincter on *T. talicei.* Volonterio (2009), while re-describing *T. talicei,* has shown the existence of a single conspicuous and asymmetric distal sphincter, pointing out similarities of this species with *T. mertoni.* Likewise *T. talicei* and *T. mertoni,* *T. cyanoglandula* also have only one vaginal sphincter, which is also distal and asymmetric (Seixas et al. 2015a).

*Temnocephala dionii* have a unique cirrus with a “groove between introvert and the shaft” and a single vaginal sphincter (Ponce de León et al. 2015), which differentiates *T. dionii* from the new species. This cirrus’ shape characteristics have some similarities with the original description of *T. mexicana*’ cirrus, pointing out the importance of a future revision of *T. mexicana.*

*Temnocephala axenos,* alike *T. grisella sp. nov.*, has two vaginal sphincters, one proximal asymmetric and a symmetric distal, but they greatly differ on size. In the re-description made by Volonterio (2007), *T. axenos* presented both vaginal sphincters with similar average sizes (43 µm proximal e 45.5 µm distal), whereas *T. grisella sp. nov.* presents one proximal large vaginal sphincter (83 µm on average) and a smaller distal one (56.5 µm on average). The author also pointed out that both sphincters of *T. axenos* are located at the final portion (distal) of the vagina, very close to one another. In contrast, in *T. grisella sp. nov.*, the vaginal portion between the proximal and distal sphincters is quite long with a strong muscular wall (Fig. 4a - aps), measuring 38 µm in length on average. Therefore, in *T. grisella sp. nov.*, the proximal sphincter is located in the middle of the vagina, while the distal one is at the tip end of the organ. The total vaginal length of *T. grisella sp. nov.* is larger than *T. axenos,* and both species have a vesicula intermedia, rather than seminal receptacles, that it is also slightly larger in specimens of *T. grisella sp. nov.* (Table 1).

Volonterio (2007) described *T. axenos*’ EPs like “elliptical excretory syncitia, small, extends from base of external tentacles to level of anterior portion of intestine”, however she did not provide character measurements. *Temnocephala grisella sp. nov.* has elongate EPs, wider in the area surrounding the excretor pore. The excretory pore is central and in the anterior portion of the plates. *Temnocephala grisella sp. nov.*’ EP is the larger in total length (397.5 µm on average) among epibionts temnocephalans on crustaceans already registered, *T. cyanoglandula*’s being the second largest, with a total length of 284.4 µm on average (Seixas et al. 2015b). The larger than wide EPs of *T. grisella sp. nov.* are evidenced with the ratio of total length of the EPs/total body length. Six EPs, approximately, could occupy the total length of the body while in the wider than long EPs’ species, such as *Temnocephala trapeziformis* Amato, Amato & Seixas, 2006 (Seixas et al. 2015b), 17 EPs are necessary to occupy the total length of the body.

**Figure 7.** Cirrus of *Temnocephala grisella sp. nov.*, showing the limit of introvert-shaft (arrow). Scale bar = 25µm.
Figure 8. *Temnocephala grisella* sp. nov. (8a) Reproductive system, showing: after proximal sphincter - vagina (aps), cirrus (c), distal vaginal sphincter (dvs), genital atrium (ga), ovary (o), prostatic bulb (pb), proximal vaginal sphincter (pvs), seminal vesicle (sv), vesicula intermedia (vi), and vesicula resorbens (vr). Scale bar = 100 μm. (8b-8c) Partial female reproductive system. (8b) after proximal sphincter of the vagina (aps), anterior portion of the proximal vaginal sphincter (apvs), before proximal sphincter of the vagina (bps), distal vaginal sphincter (dvs), ovary (o), and posterior portion of the proximal vaginal sphincter (ppvs). Scale bars = 50 μm. (8c) anterior portion of the distal vaginal sphincter (advs), genital atrium (ga), ovary (o), posterior portion of the distal vaginal sphincter (pdvs), and proximal vaginal sphincter (pvs). Scale bars = 50 μm. (8d) Adult specimen, showing the cyanophilous glands (arrows). Scale bar = 500 μm.
The accelerated process of degradation becomes even more worrying in a region where a great part of the fauna is still unknown, thus, it is essential to carry out taxonomic, ecological and environmental studies whose results make the elaboration of conservation actions possible.

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Author contributions

Samantha Alves Seixas - Substantial contribution in the concept and design of the study; Contribution to data collection; Contribution to data analysis and interpretation; Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

Norton Dametto - Contribution to data collection; Contribution to data analysis and interpretation; Contribution to manuscript preparation;

Eduardo Périco - Contribution to manuscript preparation; Contribution to critical revision, adding intellectual content.

Table 1. Morphometric data from *Temnocephala grisella* (present work) comparing with *Temnocephala mertoni* and *Temnocephala axenos* (Volonterio 2007). Measurements are in micrometers (μm), unless otherwise indicated.

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<td>Range (mean ± SD; N)</td>
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<td>Total body length (without tentacles)</td>
<td>1.04 - 1.48 mm (1.26 ± 0.14; 11)</td>
<td>1.16 - 2.66 mm (1.75 ± 0.44; 13)</td>
<td>1.42 - 4.05 mm (2.65 ± 0.70; 25)</td>
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<td>Total body width</td>
<td>0.50 - 0.73 mm (0.64 ± 0.08; 11)</td>
<td>0.82 - 1.83 mm (1.44 ± 0.35; 13)</td>
<td>1.05 - 2.39 mm (1.69 ± 0.36; 25)</td>
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<td>Vagina (total length)</td>
<td>58 - 92 (75 ± 12; 15)</td>
<td>38 - 107 (70 ± 22; 15)</td>
<td>65 - 137 (99 ± 23; 10)</td>
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<td>Proximal vaginal sphincter (total diameter)</td>
<td>_</td>
<td>31 - 60 (43 ± 9; 15) A</td>
<td>62 - 105 (83 ± 13; 16) A</td>
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<td>Distal vaginal sphincter (total diameter)</td>
<td>36 - 56 (44 ± 6; 15) A</td>
<td>27 - 62 (45.5 ± 12; 15) S</td>
<td>40 - 85 (56 ± 13; 16) S</td>
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<td>Vesicula intermedia (length)</td>
<td>29 - 127 (56.5 ± 24; 15)</td>
<td>33 - 91 (50 ± 17; 14)</td>
<td>35 - 100 (66 ± 21; 11)</td>
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<td>Seminal vesicle (length)</td>
<td>83 - 176 (130 ± 26; 15)</td>
<td>81 - 212 (146 ± 43; 15)</td>
<td>70 - 245 (146 ± 42; 25)</td>
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<td>Seminal vesicle (width)</td>
<td>43.5 - 107 (79 ± 21; 15)</td>
<td>58 - 154 (90 ± 27. 14)</td>
<td>52 - 125 (85 ± 23; 25)</td>
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<td>Prostatic bulb (length)</td>
<td>83 - 136 (108 ± 15; 14)</td>
<td>69 - 161 (108 ± 33; 15)</td>
<td>145 - 400 (255 ± 64; 25)</td>
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<tr>
<td>Prostatic bulb (width)</td>
<td>54 -100 (82 ± 13; 15)</td>
<td>47 -107 (68 ± 19; 14)</td>
<td>57 - 192 (120 ± 33; 25)</td>
</tr>
<tr>
<td>Cirrus (length)</td>
<td>123 -158 (138 ± 10; 15)</td>
<td>129 -163 (141 ± 11; 15)</td>
<td>195 - 212 (202 ± 9; 3)</td>
</tr>
<tr>
<td>Shaft (width at base)</td>
<td>38 -56 (46 ± 6; 15)</td>
<td>31 -54 (42 ± 6; 15)</td>
<td>65 - 72 (68 ± 4; 3)</td>
</tr>
<tr>
<td>Introvert (length)</td>
<td>24-31 (27 ± 3; 15)</td>
<td>18 -29 (24 ± 3; 14)</td>
<td>25 -30 (27 ± 2; 3)</td>
</tr>
<tr>
<td>Introvert (maximum width)</td>
<td>13-16 (14 ± 1; 15)</td>
<td>11 -13 (12 ± 0.9; 14)</td>
<td>15 - 17 (17 ± 1;3)</td>
</tr>
</tbody>
</table>

SD = Standard deviation; N = number of specimens measured for a given character; A = asymmetric; S = symmetric.
Conflicts of interest

The authors declare that they have no conflict of interest related to the publication of this manuscript.

References


