Effectiveness of a pre-procedural mouthwash in reducing bacteria in dental aerosols: randomized clinical trial

Abstract: The aim of this randomized, single blinded clinical trial was to evaluate the effect of a pre-procedural mouthwash containing cetylpyridinium chloride (CPC), zinc lactate (Zn) and sodium fluoride (F) in the reduction of viable bacteria in oral aerosol after a dental prophylaxis with ultrasonic scaler. Sixty systemically healthy volunteers receiving dental prophylaxis were randomly assigned to one of the following experimental groups (15 per group): (i) rinsing with 0.075% CPC, 0.28% Zn and 0.05% F (CPC+Zn+F), (ii) water or (iii) 0.12% chlorhexidine digluconate (CHX), and (iv) no rinsing. Viable bacteria were collected from different locations in the dental office on enriched TSA plates and anaerobically incubated for 72 hours. The colonies were counted and species were then identified by Checkerboard DNA–DNA Hybridization. The total number of colony-forming units (CFUs) detected in the aerosols from volunteers who rinsed with CPC+Zn+F or CHX was statistically significantly (p<0.05) lower than of those subjects who did not rinse or who rinsed with water. When all locations were considered together, the aerosols from the CPC+Zn+F and CHX groups showed, respectively, 70% and 77% fewer CFUs than those from the No Rinsing group and 61% and 70% than those from the Water group. The mean proportions of bacterial species from the orange complex were statistically significantly (p<0.05) lower in aerosols from the CPC+Zn+F and CHX groups compared with the others two groups. In conclusion, the mouthwash containing CPC+Zn+F, is effective in reducing viable bacteria in oral aerosol after a dental prophylaxis with ultrasonic scaler.

Keywords: Cetylpyridinium; Chlorhexidine; Aerosols; Mouthwashes; Microbiology.

Introduction

The propagation of oral microorganisms in the dental office during different oral procedures has been a concern. The use of certain equipment such as ultrasonic devices, highspeed dental handpieces or three-way syringes may spread aerosols and splatters containing microorganisms in the environment. These microorganisms may cause cross-infections in the dental office, jeopardizing the health of patients and dental professionals.

Different procedures, materials and antimicrobial agents have been proposed to minimize microbial cross-contamination in the dental office,
such as immunization of dental staff, decontamination of surfaces, sterilization of instruments, use of personal protective barriers and pre-procedural mouthwashes\textsuperscript{9,10,11,12}. Chlorhexidine (CHX) is considered the gold standard substance in controlling oral biofilm growth in the oral cavity or microbial spread by oral aerosols\textsuperscript{3,4,13,14,15} due to its broad antibacterial spectrum\textsuperscript{14,16} and substantivity of 8 to 12 hours.\textsuperscript{14} However, other antiseptics have also been used as pre-procedural mouthwashes, such as essential oils\textsuperscript{17} and cetylpyridinium chloride (CPC).\textsuperscript{3,15} CPC has an important antimicrobial activity\textsuperscript{18,19} and is considered a safe product for marketing.\textsuperscript{20}

The authors of this study have previously demonstrated the effectiveness of a pre-procedural mouthwash containing CPC in reducing viable bacteria in dental aerosol after a prophylaxis with an ultrasonic scaler. This study introduced CPC as a good alternative to CHX, due to its effective antibacterial action, fewer adverse effects and lower cost.\textsuperscript{3} In addition, a recent clinical study has indicated that a higher concentration of CPC – 0.07%, is very effective in controlling plaque accumulation and gingivitis;\textsuperscript{20} however, to date no studies have evaluated the effectiveness of CPC at this higher concentration as a pre-procedural mouthwash.

The present trend has been to combine more than one active substance in mouthwash and toothpaste formulations with the aim of increasing the efficacy of the products, or treating more than one clinical problem, such as plaque accumulation and halitosis;\textsuperscript{21,22} caries\textsuperscript{23} or gingival inflammation.\textsuperscript{24} Following this line of thought, zinc has been added to some oral hygiene products due to its good antimicrobial and anti-inflammatory properties\textsuperscript{25} and its capacity of neutralizing volatile compounds that play an important role in halitosis.\textsuperscript{21,26} Similarly, fluoride has been added to mouthwashes formulations due to its effect in caries prevention\textsuperscript{23} and in dental remineralization.\textsuperscript{27} Based on this body of evidence, the aim of this study was to evaluate the effectiveness of a mouthwash formulation containing 0.075\% CPC, 0.28\% zinc lactate and 0.05\% sodium fluoride (CPC+Zn+F), in reducing viable bacteria present in oral aerosol/splatter.

### Methodology

#### Sample size calculation

The ideal sample size to assure adequate power for this study was calculated based on the data of Feres et al.\textsuperscript{3} Considering a difference of at least 682 CFUs between groups CPC+Zn+F and Water and assuming a standard deviation of 435, it was defined that 15 subjects per group is adequate to provide a 95\% power with an \( \alpha \) of 0.05.

#### Subject population

Systemically healthy volunteers were selected from the population referred to the Center for Clinical Trials at Guarulhos University (Guarulhos, SP, Brazil). The medical and dental histories were obtained and a full-mouth periodontal examination was performed. The study protocol was explained to each volunteer and those who met the inclusion/exclusion criteria, signed the term of Free and Informed Consent, filled out a health questionnaire, and were enrolled in the study. The study protocol was previously approved by the Guarulhos University Ethics Committee on Clinical Research (CAAE: 41244315.4.0000.5506). The ClinicalTrials.gov identifier of the present study is NCT02875769.

#### Inclusion criteria

The inclusion criteria were as follows: male or female aged between 18–70 years, minimum of 20 natural teeth, at least 80\% of the sites with visible supragingival plaque, fewer than 10\% of sites with visible supragingival calculus, fewer than 30\% of sites with probing depth (PD) \( \geq 5\) mm.

#### Exclusion criteria

The exclusion criteria were as follows: presence of orthodontic bands; partial removable dentures; lesions of the soft or hard tissues of the oral cavity; carious lesions requiring immediate restorative treatment; history of allergy to CHX, CPC, zinc lactate or sodium fluoride; participation in any other clinical study within the one-month period prior to entering into the study; professional tooth cleaning procedure (oral prophylaxis) during the period of one month prior to entering the study; pregnant or breast-feeding
women; antibiotic therapy in the previous 6 months; continuous use of oral mouthwashes and any systemic condition that may require prophylactic medication for dental treatment (e.g., mitral valve prolapse).

Clinical assessment

In order to select the volunteers for the study, one trained periodontist (G.M.S.) performed a clinical examination that included an oral tissue evaluation and periodontal assessment. The oral examination included an evaluation of the soft and hard palate; gingival mucosa; buccal mucosa; mucogingival fold areas; tongue, sublingual and submandibular areas; salivary glands, and the tonsilar and pharyngeal areas. Visible plaque (0/1) and PD (mm) were measured at six sites per tooth (mesiobuccal, buccal, distobuccal, mesiolingual, lingual and distolingual) in all teeth, excluding third molars. The PD was recorded to the nearest millimeter using a North Carolina periodontal probe (Hu-Friedy, Chicago, IL, USA). The volunteers have refrained from any oral hygiene procedure for twelve hours prior to the clinical examination, and from eating, drinking or smoking for four hours prior to the examination.

Experimental design, randomization, allocation concealment and treatment protocol

In this single blinded randomized clinical trial (RCT), using simple randomization and a 1:1 allocation ratio, the study coordinator (L.C.F.) used a computer-generated table to randomly assign the volunteers to the following groups (n = 15 per group): 1- CPC+Zn+F (test): mouthwash containing 0.075% CPC, 0.28% zinc lactate and 0.05% sodium fluoride in an Alcohol-free base; 2- Water (negative control): water from a three-way syringe; 3- CHX (positive control): 0.12% CHX with 10% alcohol; 4- No Rinsing (negative control): standard of care - no rinsing.

In an unoccupied dental office, previously cleaned with detergent, alcohol and sodium hypochlorite, in which no regular patients had been treated on that day, the study coordinator placed five plates containing Tryptic Soy Agar with Yeast Extract enriched with 5% menadione, 5% sheep blood, and 1% N-Acetylmuramic acid (HNK plates) to sample microorganisms contained in the unit atmosphere. Three plates were placed on a support board attached to the reflector, which had been previously designed for this type of experimental design, one on the bracket tray, and another on the office bench (Figure 1). The room was locked and after 30 minutes, the plates were closed and labeled properly and sent to the laboratory for incubation. Subsequently, the volunteers were invited to sit on the dental chair to proceed with the first phase of the study. A researcher, who did not participate in any of the other phases of the study (B.R.V.), checked the randomization list containing the names of the Water and No Rinsing groups, and codes for CPC+Zn+F and CHX groups, and selected the solutions for the volunteers from groups CPC+Zn+F, CHX and Water. CPC+Zn+F and CHX mouthwashes were placed in identical opaque coded flasks, which assured the allocation concealment for these two groups, but not for the Water and No Rinsing groups. The researcher entered the dental office and handled one of three solutions to the volunteer, who rinsed for one minute with 20 ml of the solution and then expectorated the remaining liquid. The clinician (G.M.S.) did not participate on this phase of the study, and only entered the office after the rinsing phase had been completed. The volunteers were instructed to not mention to the clinician whether or not they had rinsed with a given solution. These procedures assured the masking of the clinician (G.M.S.) and the allocation concealment of the study.

After this phase, one volunteer at a time was reclined to a supine position and three new HNK agar plates were placed on the support board, one on the volunteer’s chest (immediately in front of the volunteer’s mouth) and one on the clinician’s forehead. Subsequently, the study coordinator opened the plates and the clinician proceeded with the full mouth dental prophylaxis using an ultrasonic scaler at a frequency of 25 kHz (Cavitron Select, Dentsply, York, Pa) at less than 50% power for 10 minutes. After the oral prophylaxis was completed, the five HNK agar plates were covered, labeled and sent to the laboratory for incubation. Volunteers were dismissed from the dental room. The dental office was then cleaned and disinfected with detergent, alcohol and sodium hypochlorite and the next subject was attended at least 5 hours later.
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Microbiological procedures

a. Microbial quantification: the HNK plates were incubated under anaerobic condition in jars containing 10% CO₂ at 37°C for 72 hours under anaerobic conditions, using the BD GasPak EZ system (Becton Dickinson, Sparks, MD). Using a Lab Line Colony Counter (Phoenix model CP 600 N 302), a laboratory technician counted the colony-forming units (CFUs). For this purpose, the technician was calibrated using the kappa test and a value of 90% was obtained and accepted. The technician was not aware of which plates were exposed to each treatment.

b. Microbial identification: after counting, the colonies were washed off the agar plates. The technician added 1 ml of TE buffer (pH = 7.6) to the plates and the bacterial colonies were scraped off the surfaces using L-shaped Pasteur pipettes. The suspensions were placed in individual Eppendorf tubes and sonicated for 10 seconds (Analyzer model SP 2000 UV) and the optical density (OD) at 700 nm of each suspension was adjusted to a final value of 1.0, corresponding to approximately 10⁶ bacterial cells/ml. After this, 10 ul of each suspension was removed and placed in another Eppendorf tube and 140 ul of TE buffer and 100 ul of 0.5 M NaOH were added. The samples were analyzed for 40 oral bacterial species using the Checkerboard DNA-DNA Hybridization technique.⁸⁻²⁹

c. Checkerboard DNA-DNA hybridization: the samples were boiled for 10 min. and neutralized using 0.8 ml of 5 M ammonium acetate. The released DNA was then placed into the extended slots of a Minislot 30 apparatus (Immuneletics, Cambridge, MA, USA), concentrated on a 15 x 15 cm positively charged nylon membrane (Boehringer Mannheim, Indianapolis, IN, USA) and fixed to the membrane by baking it at 120°C for 20 min. The membrane was placed in a Miniblotter 45 (Immuneletics, Cambridge, MA, USA) with the lanes of DNA at 90° to the lanes of the device. Digoxigenin-labelled whole genomic DNA probes for 40 bacterial species were hybridized in individual lanes of the Miniblotter. After hybridization, the membranes were washed at high stringency and the DNA probes were detected using the antibody to digoxigenin conjugated with alkaline phosphatase and chemiluminescence detection using the scanner G:Box iChemi XL (Syngene/ USA). The last two lanes in each run contained standards at concentrations of 10⁵ and 10⁶ cells of each species and the signals were converted to absolute counts by comparing them with the controls in each membrane. The sensitivity of the assay was adjusted to allow detection of 10⁴ cells of each bacterial species.

Primary and secondary outcome variables

The primary outcome variable was the difference between groups for the mean number of total CFUs in all plates (all locations). Secondary outcome variables were differences among groups for the following parameters: mean number of total CFUs in plates positioned on the clinician forehead, volunteer’s chest and support board; mean proportion of the microbial complexes, as well as the mean percent reduction in CFUs in the CPC+Zn+F and CHX groups in comparison with the Water and No Rinsing groups.

Statistical analysis

The total number of CFUs in the plates placed on the support board, clinician and volunteer, as well as all locations grouped were computed for each volunteer and then averaged in each group.
For the support board, the mean number of CFUs of the three plates per volunteer was computed. For the Checkerboard DNA-DNA hybridization analysis, the mean levels of each bacterial species was averaged within each participant and then across participants in each of the four groups. After this, the percentage of the total DNA probe counts (proportions) was determined per volunteer, and averaged across volunteers within each group. In addition, the proportions of the species belonging to the six complexes and the “other” group defined by Socransky et al. were added to generate the proportion of each complex per group. The significance of differences between groups for the microbial data and for age were determined by Kruskall-Wallis and Dunn tests. Chi-square test was used to compare the differences in the frequency of gender and smoking status. The level of significance was set at 5%.

Results

This RCT was conducted between February and April of 2015. Figure 2 presents the flow diagram of the study. Sixty volunteers participated in the study. Table 1 presents the demographic characteristics of the volunteers according to the different experimental groups. No statistically significant differences were observed among the four groups at the time of data collection. No volunteer reported any adverse events associated with the use of the mouthwashes evaluated.

No bacterial growth was detected in the agar plates located in the dental office before the experiment started (environmental contamination). The mean CFUs in different locations according to the experimental groups are presented in Figure 3. The aerosol/splatter from volunteers that rinsed with CPC+Zn+F or CHX harbored statistically significantly (p < 0.05) lower CFUs counts than those who rinsed with water or who did not rinse.

CPC: cetylpyridinium chloride; Zn: zinc lactate; F: sodium fluoride; CHX: chlorhexidine digluconate.

Figure 2. Flow chart of the study design.
not rinse. This occurred in the agar plates placed on the clinician’s forehead, volunteer’s chest and when all plates were evaluated together. However, when the plates placed on the support board were evaluated, this difference was not statistically significant (p > 0.05) between the Water group and the other two mouthwash (CPC+Zn+F or CHX) groups.

Table 2 shows the mean decrease in CFUs in the CPC+Zn+F and CHX groups compared with No Rinsing and Water groups. When all locations were considered together, the aerosols/splatters from the CPC+Zn+F and CHX groups showed, respectively, 70% and 77% fewer CFUs than the group that did not rinse, and 61% and 70% than those from the group that rinsed with water. Figure 4 presents the mean proportions of DNA probe counts of the 40 subgingival species evaluated by Checkerboard DNA-DNA Hybridization technique. No statistically significant (p > 0.05) differences between groups were observed for any individual proportion of the species tested. When the bacterial species were grouped into “Microbial Complexes” (Figure 5), the data showed that aerosols/splatters from CPC+Zn+F and CHX groups had a lower proportion of bacterial species from the orange complex (41.4% and 39.1%, respectively) when compared with aerosols/splatter from No rinsing and Water groups (51.1% and 47.1%, respectively).

Discussion

The results of the present study showed that oral aerosol/splatter from subjects who rinsed with a mouthwash containing 0.075% CPC, 0.28% Zn and 0.05% F, before receiving an ultrasonic prophylaxis, harbored a significantly lower bacterial content that from subjects who did not rinse or rinsed with water. This pre-procedural mouthwash was as effective as the CHX positive control mouthwash in reducing viable bacteria in dental aerosols/splatter. These results are in agreement with those of a previous study that evaluated a mouthwash containing CPC.3

According to our results, the clinician was the person who most benefited from the pre-procedural rinsing. The blood agar plates positioned on the clinician’s forehead, from the volunteers who rinsed with CPC+Zn+F or CHX, harbored, 89% and 94% fewer CFUs respectively, than the plates from the volunteers.
who did not rinse. This represents important protection for the dentist and dental hygienists, who are the main targets of the microorganisms generated during oral procedures.\(^6\) It is important to emphasize that the dental/surface barriers, the methods most commonly used to minimize cross-infection in the dental office, do not reduce the levels of microorganisms in the environment. Pre-procedural rinsing provides a viable

![Figure 4](image-url)

**Figure 4.** Mean percentage of DNA probes counts of 40 subgingival species in the four experimental groups. The species were ordered according to the microbial complexes described by Socransky et al.\(^{30}\) The proportion that each species was determined for each participant and then averaged in each experimental group. The significance of differences among the four groups was assessed using the Kruskal-Wallis test (p < 0.05, adjusted for multiple comparisons).

![Figure 5](image-url)

**Figure 5.** Pie charts of the mean proportion of each microbial complex in the four experimental groups. The colors represent different microbial complexes described by Socransky et al.\(^{30}\) The significance of differences among experimental groups was assessed using the Kruskal-Wallis and Dunn tests (different capital letters indicate significant differences between groups, p < 0.05).
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Mean of protection because these barriers, such as gloves, masks and glasses may have openings, smaller pores or defects, through which bacteria can pass. Furthermore, the aerosol particles may remain in the environment for up to 4 hours after a procedure and normally, the clinician and patients remove the protective barriers shortly after completion of the procedure. Therefore, the risk of airway contamination by these microorganisms even after the completion of the appointment is high. Thus, minimizing the quantity of microorganisms in the oral cavity before the aerosol/splatter is generated is essential to reduce the risk of cross-infection in the dental environment.

In addition, the pre-procedural rinsing also represented a major benefit for the patients. In the present study, high bacterial counts, which reached an average of ≈1,000 CFUs in the Water and No Rinsing groups, were observed on the blood agar plates positioned on the volunteer’s chest. The plates positioned on the chest of the volunteers who pre-rinsed with CPC+Zn+F showed 55% fewer bacterial colonies than those who did not rinse or who rinsed with water, suggesting a major effect of this solution in reducing the levels of bacteria in the patient’s oral cavity. A similar effect was observed for the CHX group, which showed 60% fewer bacterial colonies than the Water and No Rinsing groups. These are relevant items of information for the dental practitioner, as they may benefit the patients in several ways during dental procedures, such as for example, by reducing the odds of bacterial spread to other areas of the body, such as the eyes through aerosol or even into the lungs through inhalation.

The previous studies that have evaluated the effects of different antiseptics in reducing aerosol contamination, have recommended CHX as the gold standard. CHX as a pre-procedural rinse has been compared with other antiseptics, such as essential oils, povidone iodine and ozone, tea tree oil, a solution containing different types of herb extracts and CPC. CHX was shown to be more effective than most of these other antimicrobial agents. Only CPC has shown to be as effective as CHX in reducing the content of bacteria in dental aerosols, results that corroborate those reported in the present study for 0.075% CPC+Zn+F. An additional advantage for CPC compared with CHX is its lower cost and the reduced chance of patient intolerance to the product. Therefore, the main strength of this study is that it is the first RCT to determine the efficacy of a new formulation containing CPC in reducing oral bacterial load in the patient’s mouth and consequently, in the aerosols, as well as determining the microbial composition of these aerosols.

The analysis of the microbial composition showed statistically significantly higher proportions of putative pathogens from the orange complex in the aerosols/splatters from the Water and No Rinsing groups in comparison with those from the CPC+Zn+F and CHX groups, in agreement with a previous study. This represents an additional beneficial effect of the pre-rinsing, since certain species from the orange complex, such as the Fusobacterium species, may be associated with the etiology of ophthalmic and respiratory infections.

One limitation of the present study is that it presents results for a single dental procedure, prophylaxis with an ultrasonic scaler that has been shown to have a great potential for aerosol generation in the dental office. However, other dental procedures can also generate a large amount of aerosol with infectious components launched into the dental environment, such as the air turbine handpiece, air-water from a three-way syringe and sodium bicarbonate jet. Therefore, one

Table 2. Mean percentage of CFUs reduction (average values) in the CHX and CPC+Zn+F groups compared with the No Rinsing and Water groups considering all evaluated locations.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Clinician</th>
<th>Volunteers</th>
<th>Board</th>
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<td>CHX versus No rinsing</td>
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<td>60</td>
<td>81</td>
<td>77</td>
</tr>
<tr>
<td>CPC+Zn+F versus No rinsing</td>
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<td>55</td>
<td>70</td>
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<tr>
<td>CHX versus Water</td>
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<td>70</td>
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<tr>
<td>CPC+Zn+F versus Water</td>
<td>78</td>
<td>55</td>
<td>59</td>
<td>61</td>
</tr>
</tbody>
</table>

CFUs: colony-forming units; CPC: cetylpyridinium chloride; Zn: zinc lactate; F: sodium fluoride; CHX: chlorhexidine digluconate; %: percentage.
could assume that rinsing with CPC would yield similar benefits if used before the majority of dental procedures.

Conclusion

In conclusion, the results of this study showed that a mouthwash containing 0.075% CPC, 0.28% zinc lactate and 0.05% sodium fluoride as a pre-procedural mouthwash was effective in reducing bacterial species present in viable oral aerosols during prophylaxis with ultrasonic instruments.

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References

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