Genetic parameters and predicted gains with selection of interspecific hybrids of *Paspalum* for seed production

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Abstract: The aim of this study was to estimate the genetic parameters and predicted gains with selection of interspecific hybrids of *Paspalum* in relation to seed production traits. Data were analyzed in randomized block design, with 23 genotypes arranged into ten blocks, according to the methodology of mixed models by the REML/BLUP procedure. The traits considered in the study were: number of total tillers/plant, number of reproductive tillers/plant, number of racemes/inflorescence, number of seeds/inflorescence, weight of a thousand seeds and seed production. Variability among genotypes, high heritability (>0.50), except total tillers (0.44) and number of racemes (0.36), and high accuracy (>0.90) were identified for all traits. The hybrids 10E5052, 10E4026, 10E507, 10E4025 and 10E40104 are among the top ten because they have high genetic values in three or more traits, indicating that these genotypes should be recommended for direct use in planting or potential parents to be used in new crosses.

**Keys words**: *Paspalum plicatulum, Paspalum lepton, REML/BLUP, heritability, selective accuracy.**

**INTRODUCTION**

The *Paspalum* genus has a large number of species considered as excellent forages, not only because of its ecological, productive, and bromatological quality, but also because of its great heterogeneity: apomixis, polyploidy, and interspecific hybridization (Sartor et al. 2011).

The improvement of apomictic species, totally or highly sexual plants are necessary, so that it is possible to perform crosses, thus releasing genetic variability (Savidan et al. 1989). According to Sartor et al. (2009), the discovery of sex diploid plants in natural populations of *P. plicatulum* made it possible to obtain a sexual tetraploid plant, called 4c-4x, by inducing polyploidy with colchicine. From crosses with compatible species, new interspecific hybrids were obtained, producing new genetic combinations, permanently fixing a heterozygous progeny for immediate evaluation such as F1 varieties potencies (Aguilera et al. 2011, Pereira et al. 2017).

However, the genetic improvement in the animal production species prioritizes effective gains in forage traits, such as total dry matter and leaves (Pereira et al. 2017). Currently, strategies that incorporate a selection vision that considers
Agronomic characteristics and the potential capacity to produce viable seeds are crucial for the wide dissemination of new cultivars (Lopes and Franke 2011). In this context, genetic improvement depends on the correct selection of the best individuals as parents of the next generations, in order to reduce efforts and shorten the time needed to obtain superior genetic material through apomixia (Sartor et al. 2009).

According to Resende (2016), the success of breeding programs depends on precise estimates of genetic parameters, including reliable predictions of breeding values. Restricted Maximum Likelihood (REML) followed by the Best Linear Unbiased Prediction (BLUP) is the standard procedure in quantitative genetics and selection in perennial plants, allowed enables to achieve maximum genetic gain in selected breeding populations (Resende and Duarte 2007). The principle of REML/BLUP procedure lies in interative maximization of a likelihood function to estimate genetic variances through REML that are then employed by BLUP procedures in order to predict individual breeding values (Lynch and Walsh 1998). BLUP is considered a preferred procedure because it offers more precision for various experimental conditions, and it maximizes the correlation between the true and predicted genotypic values and the predicted genotypic value relative to other methodologies, which is essential for the breeder (Resende 2002). The objective of this study was to estimate the genetic parameters and predicted gains with selection, based on the production seed traits of interspecific hybrids P. plicatum × P. lepton.

**MATERIAL AND METHODS**

The experiment was conducted in the 2013/2014 and 2014/2015 growing seasons in Eldorado do Sul (Rio Grande do Sul state, RS), located in the Central Depression region (lat 30° 05’ 52” S, long 51° 39’ 08” W, and alt 32 m asl). The climate is type Cfa, according to the Köppen classification system, with annual rainfall of 1440 mm (monthly average of 120 mm). The soil is dystrophic Argisol (Rhodustults, Pvd) (Santos et al. 2013), with the following chemical characteristics: pH (H₂O) = 5.6; SMP index = 6.1; P = 4.5 mg dm⁻³; K = 80 mg dm⁻³; organic matter (OM) = 1.2%; Al³⁺ = 0.0 cmol dm⁻³; Ca²⁺ = 2.9 cmolc dm⁻³; Mg²⁺ = 1.4 cmolc dm⁻³; and cation exchange capacity (CEC) = 8.4 cmolc dm⁻³. The experimental area (11.0 m × 28.0 m = 308.0 m²) was corrected with 2215 kg ha⁻¹ of lime on July 7, 2013. In October 2013, 250 kg ha⁻¹ of 5-20-20 fertilizer completed the demand for potassium. This recommended amount was supplemented with 100 kg ha⁻¹ of N (urea) and 125.5 kg ha⁻¹ of P₂O₅ (triple superphosphate), meeting the needs of perennial warm-season grasses (CQFS-RS/SC 2016). In the second year, 25 kg ha⁻¹ of N (urea) and 65 kg ha⁻¹ of P₂O₅ (replenishment dose) were applied.

Twenty-three apomictic genotypes of *Paspalum* were used (10E6086, 10E5052, 10E4071, 10E5023, 10E4026, 10E507, 10E4025, 10E40104, 10E6084, 10E6069, 10E4076, 10E4077, 10E5017, 10E6024, 10E6088, 10E6022, 10E4099, 10E4087, 10E40177, 10E50150, 10E4039, 10E6058 and 10E6047), belonging to Universidade Federal do Rio Grande do Sul, originated from artificial crosses between *P. lepton* (male genitor; native genotype from Rio Grande do Sul state), and *P. plicatum* 4c-4x (female genitor), from Botanical Institute of the Northeast (IBONE, Argentina), with the objective of get promising combinations related to forage production. The male genitor (*P. lepton*) stood out in preliminary studies for greater forage production (Pereira et al. 2011, Pereira et al. 2012).

Seedlings were kept in a greenhouse during the winter until the time for planting in the field (Oct. 24, 2013). In the second year, rejuvenation pruning was performed on all the genotypes (Sept. 26, 2014), leaving 0.15 m of residue. Seeds were manually harvested, when more than 50% of inflorescences exhibited brown coloring and signs of dehiscence in caryopses. All inflorescences per plant were harvested (Mar. 25, 2014 - first year, and Feb. 18, 2015 - second year). Prior to harvesting, the following traits were assessed: a) TT, number of total tillers - direct count of all the tillers/plant; b) RT, reproductive tillers/plant - direct count of the tillers that formed inflorescences; and c) NRI, number of racemes/inflorescence - average number of racemes of six inflorescences from each plant, randomly chosen for each genotype.

The following analyses were conducted after seed harvesting: a) SP, seed production/plant - the inflorescences collected were dried in a forced air oven at 30 °C for 72 h and manually threshed. Sieves were used to remove impurities and a “South Dakota” blower to separate heavy and empty seeds. After cleaning, the pure seeds of each genotype were weighed, expressed in g; b) WTS, weight of a thousand seeds - average weight of eight subsamples of 100 seeds, multiplied by 10 (Brasil 2009); and c) NSI, number of seeds/inflorescence - rule of three between WTS and the average weight of seeds per inflorescence. The average weight of seeds/inflorescence was determined at harvest, when six intact inflorescences were separated from each genotype, manually threshed, processed and individually weighed on
a precision scale (expressed in g).

The experimental design was a randomized block design, with the genotypes arranged individually in ten blocks. The experiment consisted of 230 experimental units (spacing on the row and interrow of 1.0 m), corresponding to the twenty-three genotypes in ten replications. The data were analyzed using the mixed model methodology, using the SELEGEN software (Resende 2016), obtaining the estimate for genetic parameter and the predicted genotypic values, using the REML/BLUP procedure, for the genotype ordering. The genetic-statistical model used considered the randomized block design in one location and two harvest, according to the model below (Resende 2007):

\[ y = Xm + Zg + Wp + e \]

In which, \( y \) is the data vector; \( m \) is the vector for the effects of the measurement-repeat combinations (assumed as fixed) added to the general mean; \( g \) is the vector of the genotypic effects (assumed to be random); \( p \) is the vector of the permanent environment effects (experimental units) (random); \( e \) is the vector of errors or residuals (random). The capital letters (\( X, Z \) and \( W \)) represent the incidence matrices for the said effects (\( m, g \) and \( p \), respectively).

The structures of means and variances associated with the model are described by the following:

\[
\begin{align*}
    y & \sim N(Xm, V) \\
    g & \sim N(0, A\sigma^2_g) \\
    p & \sim N(0, I\sigma^2_p) \\
    e & \sim N(0, I\sigma^2_res)
\end{align*}
\]

In which, \( V \) is the phenotypic covariance matrix; \( I \) is an identity matrix; \( \sigma^2_g, \sigma^2_p \) and \( \sigma^2_res \) are the genotypic, environment and residual variances, respectively.

The covariance between all of the random effect models was assumed by the following:

\[
    \text{Cov}(g, p') = 0; \quad \text{Cov}(g, e') = 0 \quad \text{and} \quad \text{Cov}(p, e') = 0
\]

The assumed distributions and mean structures (S) and variance (Var) were:

\[
\begin{bmatrix}
    y \\
    g \\
    p \\
    e
\end{bmatrix}
= \begin{bmatrix}
    Xm \\
    0 \\
    0 \\
    0
\end{bmatrix}
; \quad
\begin{bmatrix}
    y \\
    g \\
    p \\
    e
\end{bmatrix}
= \begin{bmatrix}
    V \\
    ZG \\
    WC \\
    R
\end{bmatrix}

\[
G = A\sigma^2_g R = I\sigma^2_p C = I\sigma^2_e \quad \text{and:} \quad V = Z A\sigma^2_g Z' + W I\sigma^2_p W' + I\sigma^2_e = ZGZ' + WCW' + R
\]

The system of linear equations [mixed model equations (MMEs)] that were used to obtain the solutions of the model were:

\[
\begin{bmatrix}
    XX \\
    ZX \\
    WZ \\
    WX \\
    WW + b \lambda_1 \\
    WW + b \lambda_1
\end{bmatrix}
\begin{bmatrix}
    \hat{m} \\
    \hat{g} \\
    \hat{p} \\
    \hat{e}
\end{bmatrix}
= \begin{bmatrix}
    Xy \\
    Zy \\
    W'y
\end{bmatrix}

\[
\lambda_1 = \frac{\delta^2_{res}}{\delta^2_g} = \frac{1 - h^2_g - c^2}{h^2_g}; \quad \lambda_2 = \frac{\delta^2_{res}}{\delta^2_e} = \frac{1 - h^2_e - c^2}{c^2}
\]

The estimates of variances and genetic parameters are given as follows: genotypic variance (\( \delta^2_g \)); environment variance (\( \delta^2_p \)); residual variance (\( \delta^2_{res} \)); phenotypic variance (\( \delta^2 = \delta^2_g + \delta^2_p + \delta^2_{res} \)); individual heritability in the broad sense (\( h^2 = \delta^2_g / \delta^2 \)); genetic variation coefficient \( CV_g = 100 / \hat{\delta}^2_g \); residual variation coefficient \( CV_{res} = \sqrt{\delta^2_{res}} / \hat{\delta}^2_g \); relative variation coefficient \( CV_r = CV_g / CV_{res} \); overall mean (\( \bar{x} \)) and genetic accuracy in genotype selection (Acgen) (Resende and Duarte 2007):

\[
\text{Acgen} = \left[ \frac{b h^2_g}{1 + (b - 1) h^2_g} \right]^{1/2}
\]

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In which, $b$ is number of blocks considering randomized complete block design.

The significance of the random effects was obtained through Deviance Analysis, using the REML method (Restricted Maximum Likelihood), via LRT (Likelihood Ratio Test). The deviances were obtained as described by Resende (2016), using the model with and without the respective effects, subtracting the deviance obtained in the complete model, of the model without the effect, and compared to the value of the Chi-square ($\chi^2$) at 1% probability, with one degree of freedom. The factor block, considered as fixed effect, was tested by the F test of Snedecor.

**RESULTS AND DISCUSSION**

The highest estimates of the genetic variance ($\hat{\sigma}^2_g$) were for WTS, NSI and RT traits, indicating that, in this germplasm, there are favorable conditions for selection and improvement of these traits (Table 1). When values are positive and non-zero, there is greater genetic variation and the greater the chances of genetic gains with selection. In *Paspalum* species, a large portion of the components linked to seed yield are related to inflorescence (NSI and WTS) and its reproductive tillers (RT) (Lopes and Franke 2011, Lopes et al. 2016) serving as an indicator for breeding programs aimed at the production of forage species seed (Biligetu et al. 2012).

The success with the selection also depends on the environmental variation ($\hat{\sigma}^2$), since the present variation in the phenotype is due to the environmental conditions. The estimates of the environmental variance were low when compared to the others, inferring that there was high experimental precision and low environmental interference on the genotypes, confirming the adequate planning for the plot size and number of experiment repetition (Table 1).

It is of great importance that the phenotypic variation ($\hat{\sigma}^2$) be composed, for the most part, of variations from the genotype of the selection candidates, since it contributes to higher heritability of the trait in question. We highlight the high phenotypic variation of the WTS and NSI traits, basically composed by the variation of the genotype (Table 1). According to Lopes et al. (2017), WTS and NSI were the most efficient (largest relative contribution) in explaining genetic variability among *P. plicatulum × P. lepton* hybrids, showing the importance of these variables as a component in the production of seeds and, possibly in the selection of superior genotypes.

Estimates of individual heritability in the broad sense ($h^2_g$) for the evaluated traits revealed values considered medium ($0.15 < h^2_g < 0.50$) and high ($h^2_g > 0.50$) (Resende 2007) (Table 1). Şeker et al. (2014), when evaluating the seed production from *Dactylis glomerata* L. populations, found high values of $h^2_g$ (0.64) showing strong genetic control of

<table>
<thead>
<tr>
<th>Parameters</th>
<th>TT</th>
<th>RT</th>
<th>NRI</th>
<th>NSI</th>
<th>WTS</th>
<th>SP</th>
</tr>
</thead>
<tbody>
<tr>
<td>$LRT_{genotype}$ ($x^2$)</td>
<td>195.46**</td>
<td>215.67**</td>
<td>140.44**</td>
<td>418.41**</td>
<td>542.03**</td>
<td>239.32**</td>
</tr>
<tr>
<td>$LRT_{res}$ ($x^2$)</td>
<td>0.00</td>
<td>0.00</td>
<td>0.01</td>
<td>0.00</td>
<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td>$F_{(block)}$</td>
<td>44.53**</td>
<td>17.53**</td>
<td>2.45</td>
<td>3.41**</td>
<td>0.57</td>
<td>39.41**</td>
</tr>
<tr>
<td>$\hat{\sigma}^2_g$</td>
<td>338.33</td>
<td>364.02</td>
<td>0.30</td>
<td>1207.29</td>
<td>0.21</td>
<td>271.93</td>
</tr>
<tr>
<td>$\hat{\sigma}^2_e$</td>
<td>1.60</td>
<td>1.30</td>
<td>2.10^3</td>
<td>2.40</td>
<td>2.10^4</td>
<td>0.91</td>
</tr>
<tr>
<td>$\hat{\sigma}^2_p$</td>
<td>767.99</td>
<td>688.06</td>
<td>0.84</td>
<td>1651.54</td>
<td>0.25</td>
<td>532.97</td>
</tr>
<tr>
<td>$\hat{\sigma}^2_{res}$</td>
<td>428.06</td>
<td>322.74</td>
<td>0.54</td>
<td>441.86</td>
<td>0.04</td>
<td>260.13</td>
</tr>
<tr>
<td>$h^2_g$</td>
<td>0.44±0.09</td>
<td>0.53±0.09</td>
<td>0.36±0.08</td>
<td>0.73±0.11</td>
<td>0.84±0.12</td>
<td>0.51±0.09</td>
</tr>
<tr>
<td>$CV_g$ (%)</td>
<td>11.61</td>
<td>15.91</td>
<td>9.17</td>
<td>18.25</td>
<td>12.59</td>
<td>25.78</td>
</tr>
<tr>
<td>$CV_{res}$ (%)</td>
<td>13.06</td>
<td>14.98</td>
<td>12.31</td>
<td>11.04</td>
<td>5.49</td>
<td>25.22</td>
</tr>
<tr>
<td>$CV_e$ (%)</td>
<td>0.89</td>
<td>1.06</td>
<td>0.75</td>
<td>1.65</td>
<td>2.29</td>
<td>1.02</td>
</tr>
<tr>
<td>Acgen</td>
<td>94.20</td>
<td>95.84</td>
<td>92.03</td>
<td>98.21</td>
<td>99.05</td>
<td>95.52</td>
</tr>
<tr>
<td>Overall mean ($\bar{X}$)</td>
<td>158.43</td>
<td>119.94</td>
<td>5.97</td>
<td>190.39</td>
<td>3.64</td>
<td>63.96</td>
</tr>
</tbody>
</table>

1. Description of genetic parameters: genotypic variance ($\hat{\sigma}^2_g$), permanent environment variance ($\hat{\sigma}^2_p$), phenotypic variance ($\hat{\sigma}^2$), residual variance ($\hat{\sigma}^2_{res}$), individual heritability in the broad sense ($h^2_g$), genetic variation coefficient ($CV_g$), residual variation coefficient ($CV_{res}$), relative variation coefficient ($CV_e$), genetic accuracy in genotype selection (Acgen), overall mean ($\bar{X}$). **Significant at the likelihood ratio test (LRT, 1%= 6.63), considering one degree freedom by the $\chi^2$ test. †† Significant at 1% of probability, by the Snedecor F test.
Table 2. Predicted genotypic effect (g), genotypic value (u+g), genetic gains (gain) and new average ($\bar{X}_{new}$) for number of total tillers (TT), reproductive tillers (RT), number of racemes per inflorescence (NRI), number of seeds per inflorescence (NSI), weight of a thousand seeds (WTS) and seed production (SP) for interspecific hybrids of *Paspalum*

<table>
<thead>
<tr>
<th>Order</th>
<th>Genotype</th>
<th>TT (number plant$^{-1}$)</th>
<th>NRI (number inflorescence$^{-1}$)</th>
<th>WTS (g)</th>
<th>SP (g plant$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Order</td>
<td>Genotype</td>
<td>g</td>
<td>u+g</td>
<td>gain</td>
<td>$\bar{X}_{new}$</td>
</tr>
<tr>
<td>1</td>
<td>10E5052</td>
<td>33.5</td>
<td>192.0</td>
<td>33.5</td>
<td>192</td>
</tr>
<tr>
<td>2</td>
<td>10E50150</td>
<td>25.8</td>
<td>184.2</td>
<td>29.7</td>
<td>188</td>
</tr>
<tr>
<td>3</td>
<td>10E40104</td>
<td>21.6</td>
<td>180.0</td>
<td>27.0</td>
<td>185</td>
</tr>
<tr>
<td>4</td>
<td>10E6047</td>
<td>18.3</td>
<td>176.8</td>
<td>24.8</td>
<td>183</td>
</tr>
<tr>
<td>5</td>
<td>10E608</td>
<td>16.9</td>
<td>175.4</td>
<td>23.2</td>
<td>182</td>
</tr>
<tr>
<td>6</td>
<td>10E5017</td>
<td>15.7</td>
<td>174.0</td>
<td>22.0</td>
<td>180</td>
</tr>
<tr>
<td>7</td>
<td>10E4026</td>
<td>15.6</td>
<td>174.0</td>
<td>21.1</td>
<td>180</td>
</tr>
<tr>
<td>8</td>
<td>10E6068</td>
<td>8.8</td>
<td>167.2</td>
<td>19.5</td>
<td>178</td>
</tr>
<tr>
<td>9</td>
<td>10E507</td>
<td>8.6</td>
<td>166.1</td>
<td>18.3</td>
<td>177</td>
</tr>
<tr>
<td>10</td>
<td>10E4025</td>
<td>1.6</td>
<td>160.1</td>
<td>16.6</td>
<td>175</td>
</tr>
</tbody>
</table>

In general, the residual coefficients of variation (CV$_{res}$) showed medium to low magnitude for traits, indicating an acceptable environmental control and satisfactory accuracy of the genetic parameter estimates (Table 1). Since it is a larger the value of the CV$_{g}$ estimate, the greater the probability of releasing genetic variability (Falconer 1987).

The coefficient of genetic variation (CV$_{g}$) diverged between the evaluated genotypes, with the highest value regarding the SP trait, followed by NSI and RT (Table 1). According to Oliveira et al. (2015), values above 10.00 allow the breeder a better understanding of the genetic variability and, consequently, of the advances that can be obtained through the selection of a certain traits. Because it is directly proportional to the genetic variance, it allows an information of the relative magnitude of the changes that can be acquired through selection during an improvement program, since the larger the value of the CV$_{g}$ estimate, the greater the probability of releasing genetic variability (Falconer 1987).
question of productivity, a trait strongly influenced by the environment, the SP showed higher value of $CV_{\text{res}}$, because it is results from the performance of all other traits (seed production components) (Nakagawa 2014), accumulating the measurement errors. However, the result of the statistics in this study, are within acceptable limits for agricultural experimentation, providing safety and consistency for this type of data (Pimentel-Gomes 2009).

Another important parameter in defining the best breeding strategy for each trait is the relative variation coefficient ($CV_{r}$), or variation rate, since this represents the ratio between the $CV_{r}$ and $CV_{\text{res}}$. Therefore, it is not influenced by the trait mean. According to Resende and Duarte (2007), when this rate is close to or greater than 1.0, there is a very favorable situation to obtain gains in the selection of a certain characteristic. Based on this parameter, the relative variation coefficient values, with the exception of the TT and NRI traits, presented a genetic component ($CV_{g}$) approximate or greater than the residual component ($CV_{\text{res}}$). The values obtained in this study are in agreement with what was expected in situations of reduced experimental errors, which maximize accuracy.

In the genotypic evaluation context, the parameter known as accuracy (Ac$_{\text{gen}}$) refers to the correlation between the true genotypic value of the genetic material and the estimated or predicted value, based on the field experiment information, and also on the efficacy of the inference regarding the genotypic value of the hybrid or its value for cultivation and use (VCU) (Resende and Duarte 2007). In this study, the accuracy values varied between 92.03% and 99.05%, classified as very high, providing evidence that the selection based on the traits under consideration will be safe. According to Resende and Duarte (2007), accuracies above 0.70 are sufficient for a breeding program, but when the objective is to evaluate the VCU, the precision must be greater than 0.90.

When analyzing the effect of treatments, taking them as random, tests of multiple comparisons between averages should not be used, since these tests are derived from an assumption of fixed effects of treatments, and also because they produce inferences about phenotypic averages and not genotypic averages (Resende 2007). The use of more refined analytical procedures, such as the use of mixed linear models, allows a decreasing ordering of genotypes according to their genetic values, corrected and penalized for the occurrence of environmental effects (Duarte and Vencovsky 2001).

Out of the twenty-three genotypes assessed, the best ten for the traits TT, RT, NRI, NSI, WTS and SP were selected through the BLUP methodology and account for 43.5% of the evaluated germplasm (Table 2). Genetic gains were predicted, and the new estimated averages were higher than the overall average for all traits.

The selection based on the genetic value for the TT trait revealed that genotype 10E5052 had the best performance, with genetic gains, which varied between 16.6 and 33.5 tillers per plant (Table 2), raising the new average by 21.2%. It has been observed that many of the same previously selected genotypes were found to also appear in the ordering of the RT trait, especially genotypes 10E5052 and 10E40104. The genetic gain in this trait varied from 15.3 to 28.5 reproductive tillers per plant, raising the new average by 23.7%. Monteiro et al. (2016), selecting interspecific hybrids of B. decumbens, obtained gains with the selection of 73, 114 and 174% (30, 20 and 10% intensity, respectively) with high heritability amounts (0.88) in the RT trait, which may be a parameter to predict the seed production potential in forage species. In another context, Pereira et al. (2017), evaluating the genetic gain for forage traits in apomictic species of the genus Paspalum, low genetic variance and heritability (0.24) was observed for the number of tillers (TT), indicating difficulty in obtaining a genetic gain in the evaluated populations. A strategy to increase the expression of these traits is the recombination of a sexual parent in the formation of new individuals, increasing the number of heterozygous alleles, maximizing the benefits of hybrid vigor, generating greater stability to phenotypic expression.

For NRI, gains above 7.7% can be obtained with the selection of the ten best genotypes, of which only one (10E4026) is coincident in the other evaluated traits (Table 2). Genetic gains ranged from 0.5 to 1.3 racemes per inflorescence (21.9%) for this trait. According to Lopes et al. (2017), the expression of the variability of the NRI trait was equal in two years of evaluations, emphasizing the same genotype of this study (10E4026). It is worth noting that the P. plicatulum species (female genitor) presents 3-7 racemes per inflorescence and the P. lepton species (male genitor) presents 2-5 racemes per inflorescence (Wunderlin et al. 2017), probably contributing to the variability of this trait after hybridization.

High genetic gains can be obtained by selecting individuals based on their genotypic values for the NSI trait, with emphasis on genotype 10E5052. The genetic gain varied from 28.6 to 96.0 seeds per inflorescence (15.0 to 54.4%, respectively) for this trait (Table 2). The selection in the WTS trait presents three genotypes in common with the NSI
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(10E507, 10E40104 e 10E4025). With better performance, genotype 10E4077 was promising in relation to the genetic gains, which varied from 0.4 to 1.0 g 1000 seeds⁻¹ (10.4 to 28.3%, respectively) for the trait. Both traits presented the highest estimates of individual heritability in the broad sense (0.73 e 0.84, respectively) (Table 1), and it should be noted that heritability directly participates in the prediction of genetic gain, regardless of the selection method, and consequently, in decision making during the breeding process (Ramalho et al. 2012).

Genotype 10E5052 stood out in the TT, RT and NSI traits; however, it did not show the same performance in the NRI and WTS traits (Table 2). This fact is explained by the proportional behavior between the traits for seed production, in other words, the increment of one trait leads to the decrease of the other, in view of the plasticity or compensatory capacity of the plant (Nakagawa 2014).

Of course, this order was carried out taking into account the purpose of the program in increasing the expression of the SP trait. The genetic gain, in this trait, varied from 14.8 to 28.3 g seeds⁻¹ (23.2 and 44.3%, respectively) (Table 2). The genotype 10E4071 presented the best performance (highlighted only in WTS and SP) followed by genotype 10E5052 (higher expression in TT, RT and NSI, second in SP), where these two were the only ones to raise the new average above 40.0% for seed production.

According to the literature, the main components of the production of Paspalum seeds are the TT, RT, NSI and WTS traits (Lopes and Franke 2011, Lopes et al. 2016, Lopes et al. 2017). The use of these traits favors the choice of superior performance genotypes, through indirect selection in traits of difficult selection and measurement, such as seed production. The structural characteristics of the plant, theoretically the number of tillers that pass to the reproductive stage, can be used to predict the production of seeds before the beginning of flowering.

Thus, the elaboration of a comprehensive record of these traits is not only scientifically interesting, but also an important contribution in guiding the breeding program in forage plants, acting as a facilitator of the selection process.

CONCLUSIONS

The evaluated genotypes presented potential for improvement in view of the high genetic variability, high heritability, and high accuracy for the seed production traits. The hybrids 10E5052, 10E4026, 10E507, 10E4025 and 10E40104 are among the top ten because they have high genetic values in three or more traits, indicating that these genotypes should be recommended for direct use in planting or potential parents to be used in new crosses.

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REFERENCES


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