TWO INTRATHecal TRANSPLANTS OF BONE MARROW MONONUCLEAR CELLS PRODUCE MOTOR IMPROVEMENT IN AN ACUTE AND SEVERE MODEL OF SPINAL CORD INJURY

ABSTRACT

Objective: We studied transplants of bone marrow mononuclear cells (BMMC) by lumbar puncture (LP) in a severe model of spinal cord injury (SCI) using clip compression. Methods: BMMCs or saline solution were transplanted by LP 48 hours and 9 days post injury. Motor function was evaluated by BBB scale, histological analysis by Nissl technique and the verification of cell migration by PCR analysis. Results: The BBB had significantly improved in rats treated with BMMCs by LP compared with controls (p < 0.001). The histological analysis did not showed difference in the lesional area between the groups. The PCR analysis was able to found BMMCs in the injury site. Conclusions: Two BMMC transplants by LP improved motor function in a severe model of SCI and BMMC was found in the injury site.

Keywords: Spinal cord injuries; Lumbar puncture; Bone marrow cells; Stem cells.

INTRODUCTION

Spinal cord injuries (SCI) are devastating, and often affect young and healthy individuals.1 SCIs have a high incidence, between 15-40 cases per million per year globally, and impose a heavy burden on government and society.2 New therapeutic strategies have been investigated to try to reduce the severe morbidity, such as the use of bone marrow stem cell (BMSC) transplantation. The use of BMSC presents advantages when compared with other cells types, because these cells are easily expanded in vitro, there are no ethical issues regarding their use and no risk of rejection, since they are collected from the patient himself.3,4 Previous studies have presented benefits in their application in experimental SCI models showing an improvement in gait, migration to the lesion and differentiation into neurons and oligodendrocytes.5,6

One of the main factors in stem cell (SC) therapy is the route of cell delivery and the number of times SC should be administered during...
the window of opportunity. In animal models of SCI, the most common mode of delivery is direct injection (DI) into the injury site, which allows a defined number of SC to be delivered. The clinical translation with the DI transplantation is not straightforward, because of the need for a new surgical procedure and the risk of further injuring the spinal cord.7,9 Less invasive SC delivery methods need a more profound study. The intrathecal delivery by lumbar puncture (LP) is a minimally invasive approach that decreases the risk to the patient and allows delivery of multiple cell doses over a preplanned length of time.10-12 The efficacy of a single BMSC transplantation by LP was previously studied in mild SCI but showed marginal efficacy.13 We aimed to study the efficacy of two infusions of bone marrow mononuclear cells (BMMCs) by LP in a model of severe SCI.

**METHODS**

**Study design**

Eighteen female Wistar rats (60 days old) were used as recipients and 15 male Wistar rats (60 days old) were used as BMMCs donors. Animals were divided into 3 groups of 6 animals each: sham (controls); receiving BMMCs by LP; and receiving saline solution by LP.

On day 1, animals were subjected to SCI. A motor function evaluation using the Basso, Beattie, and Bresnahan (BBB) scale was performed 24 hours later in order to have the BBB baseline or exclude the animals that presented a BBB score higher than 3 points.14 BMMCs or saline were administered by LP on day 3 and day 9 after SCI. All animals were given BBB scores on days 7, 14, and 21 after SCI. After the last evaluation, they were euthanized and histologic analysis was performed. The experimental protocols used in this study followed international standards for animal testing, and were approved by the Animal Care and Ethics Committee of the Pontificial Catholic University of Rio Grande do Sul (CEUA 09/00109).

**Surgical Procedures**

Animals were weighed and anesthetized by intraperitoneal injection with 2% xylazine (0.1 ml/g) and 10% ketamine (0.22 ml/g). The rats were positioned in ventral decubitus and the skin over the thoracic spine was shaved and cleaned using antiseptic solution. The incision site was determined by palpation of the last rib and counting to define the eighth rib. A 3-cm incision at the midline over the dorsal region was performed with removal of paravertebral muscles and locating the vertebral lamina of the eighth thoracic vertebrae (T8). A T8 laminectomy was performed carefully under magnification and the dural sac identified. Spinal cord compression was performed using a 6.7 mm clip (Vicra®) with a closing pressure of 50 g/strength during 60 seconds.15 (Figure 1) After the injury, the incision was sutured with mononylon 3-0 and the antiseptic solution was applied to the incision site. Postoperatively, ceftriaxone (100 mg/kg, i.p.) and tramadol (1.5 mg/kg, s.c.) were administered and subcutaneous hydration was given with 10 ml of 0.9%. The Credé maneuver was performed at the end of the procedure to empty the bladder. Postoperative care included: Credé maneuver twice daily, inspection of the skin; pressure ulcer prevention; and fluid restriction during the acute period and at night, to decrease bladder strain. Rats were weighed once a week, and nutrition was limited to 20 grams to avoid weight gain. In case of anorexia, 10 ml of warm normal saline were administered intraperitoneally. Urinary tract infection, manifested by hematuria with pus or cloudy urine, was treated with amoxicillin with clavulanic acid (2 mg/100 g i.p.).16,17 The rats with urinary infection that were not getting better with antibiotic treatment and becoming progressively sicker were excluded from the study.

**Preparation of Bone Marrow Mononuclear Cells**

BMMCs were obtained from male Wistar rats with the Y chromosome used as a reporter of transplanted cells. Bone marrow was extracted from the humerus, femur, and tibia by flushing with 10,000 U of heparin in 50ml of Dulbecco's phosphate buffered saline (DPBS). Extracts were centrifuged at 400Xg for 10 minutes. Cell pellets were resuspended with RPMI-1640 and fractionated on a density gradient generated by centrifugation at 400Xg over a Ficoll-Paque solution (Histopaque 11191, Sigma Aldrich, St. Louis, MO, USA). The mononuclear fraction over the Ficoll-Paque layer was collected and washed twice with DPBS. Cell concentrations were determined with a Neubauer-counting chamber with the number of viable cells determined by Trypan Blue exclusion. For the detection of surface antigen, BMMCs were incubated with conjugated antibodies against CD34, BMMCs were incubated with conjugated antibodies against CD34, CD11b, CD117, CD45, and Sca1. Labeled cells were collected and analyzed using a FACSCalibur cyrometer.

**Lumbar Puncture**

BMMCs or saline solution were infused into the subarachnoid space via LP. A volume of 40 μL containing 106 cells was used.10-13 After anesthesia and skin preparation, the lumbar spinous processes were palpated to identify the L2-3 interspace. After the device was placed, a 27G needle was introduced into the subarachnoid space with a slight flexion of the animal’s body, to facilitate access to the spinal canal. The correct needle positioning in the subarachnoid space was determined by the contraction of the tail at the time of needle insertion or cerebrospinal fluid leakage. Cell infusion was performed for 1 minute to avoid reflux. LP transplants were performed twice at 48 hours and 9 days after SCI.

**Evaluation of Motor Function**

Hind limb motor function was evaluated using the BBB score.14 The animals were assessed 24 hours postoperatively; only those that scored less than 3 were included in the study. After the procedure, rats were evaluated once a week for 21 days. The rats were placed in an open field and videotaped for five minutes. The films were subsequently analyzed and the animals were scored by two evaluators. If there was disagreement between evaluators, a third reviewer was consulted.

**Histology**

On day 21, after evaluation of motor function, animals were euthanized with high doses of 2% xylazine (0.1ml/g) and 10% ketamine (0.22ml/g) intraperitonealy. Nine rats were perfused with saline followed by 4% paraformaldehyde immersion for 24 hours. The spine was carefully dissected and the spinal cord was removed. A piece of spinal cord, measuring 1cm, was dissected to perform...
the histological analysis, the main lesion produced in our study was located in the center of the piece. The next day the spinal cords were placed in 30% sucrose for dehydration for an additional 24 hours. Finally, the spinal cords were frozen in isopentane previously cooled in liquid nitrogen, acclimatized in a cryostat (Cryotome® SME, Shandon, England) sectioned at −20°C at 30µm. All sections were processed using the Nissl technique.18 The images of these sections were digitized using a stereomicroscope (Stemi SV6, Zeiss, Germany), coupled with a CCD camera (Pixera 1.2, Pixera, Pixera Corporation, CA, USA) and analyzed by software Image Pro Plus 6.1 (Media Cybernetics, San Diego, CA, USA). The sections were classified as lesioned or intact. The sections were considered lesioned when they did not have a clear anatomical definition between grey and white matter and consequently presented a “reticular” aspect. The number of injured sections was used to estimate the length of lesion in an adaptation of previously published protocols.19,20

Y-Chromosome detection using Polymerase Chain Reaction

The polymerase chain reaction (PCR) analysis was performed in order to identify the presence of transplanted BMMCs in the spinal cords. Three rats were subjected to the injury model. 3 received BMMC transplantation by LP 48 hours and 9 days after injury. Twenty-four hours after the last transplantation, rats were euthanized and samples of the spinal cord collected. We used forward primer 5'-direct atggagaattctgaggaggaga-3' and reverse primer 5'-attcacaaagctgacctccagttg-3', generating a PCR product of 324 bp, using a nested PCR technique. A positive control sample (Y-chromosome + DNA extracted from a male rat) and a negative control (no Y-chromosome + DNA extracted from a female rat) were assayed with experimental samples in every reaction. Amplified products were detected by gel electrophoresis (agarose 2% containing ethidium bromide) and viewed under ultraviolet light.

Statistical analysis

Data are presented as the mean ± standard deviation (SD) or the mean ± standard error of the mean (SEM) as indicated in the Figure Legends. Two-way analysis of variance (ANOVA) followed by the Bonferroni post-hoc test was used to analyze data obtained by the BBB test. Data obtained on histology were analyzed using one-way ANOVA followed by the Tukey test and Student’s t test. Statistical analysis was performed with PrismGraph 5.0 software (Graph-Pad Software, San Diego, CA). An α level of 0.05 was applied and a p-value of less than 0.05 was considered statistically significant.

RESULTS

Recovery of motor function

The BBB scores in the sham group were 21 while animals that underwent compression scored a maximum of 7 ± 0.5, which showed that the SCI model performed by clip compression was efficient.

Post-hoc analysis indicated that function improved significantly beginning week 2 after transplantation. The BBB had significantly improved in rats treated with BMMCs by LP compared with controls in the last two evaluations, on days 14 and 21 (p<0.001). (Figure 2)

Lesional Area Measurement

The animals treated with BMMCs by LP have similar lesional areas when compared with the control group.

Migration of donor-derived BMMCs to the site of injury

The PCR analysis was performed on 3 spinal cord samples from injured rats transplanted with BMMCs and there was a positive finding for the 342 bp band, corresponding to the specific sequence of the Y-chromosome after the second amplification at the A3 column. (Figure 3)

DISCUSSION

Studies have shown neurological benefits by using BMSC in experimental SCI models and also could present the cellular migration to the lesion and the differentiation in neurons and oligodendrocytes.5,6

In order for SC therapy to be available in clinical practice, the most effective and reasonable route of administration must be established. In this way, despite some controversies about which is the best delivery in animal studies, there seems to be a tendency for administration by DI approach to result in more neurological improvement and grafting at the lesion site.5,6,13,21,22

Vaquero et al.22 reported a more efficient implantation of the stem cells via DI than intravenous injection at the lesion site in a chronic model of SCI. Likewise, Paul et al.13 reported that a single dose of stem cells administered by DI resulted in better neurological improvement and a higher number of implantation cells when compared with the injection through the subarachnoid space or the intravenous route. The drawback of this study is the lack of a control group. However, Kang et al.23 showed that the fate of transplanted allogenic mesenchymal stem cells derived from bone marrow and the expression of neuronal growth factors are different between intravenously or DI routes in animal SCI. The mean score of the BBB motor scales was better in the intravenous group, when compared to the control and DI groups.23 We can conclude that the best route for the delivery of stem cells in the SCI model is still unknown.

Fujioka et al.24 reported a significant functional recovery in SCI after CD 133 cells were transplanted by LP when compared with the control group. In our study, the BMMCs transplanted group with two LP administration showed better BBB results starting in the second postoperative week, while saline controls had low scores throughout the 21-day follow-up period.

In an animal SCI model, Neuhuber et al.21 showed that although motor
recovery was greater after a single transplantation of neural precursor cells (NPCs) using a DI than in administration by LP, the neurological improvement in the DI and LP group was significantly better than the control groups and a similar number of SC was found in the injury site regardless of the route used. In general, this represents the opinions found in the literature, which are similar to the grafting of SC in the acute stage to be transplanted at the injury site independently of administration technique. In other words, these cells had been found at the injury site after being transplanted by intravenous, intraparenchymal, intrathecal, and intraventricular routes. 11,13,22,25

Some studies used the LP approach to inject the stem cells into the subarachnoid space and for the subsequent evaluation of their implantation. 12,13,26 Paul et al. 13 reported an SC implantation rate of 4.1% and 3.4%, respectively, 4 and 21 days after LP infusion. Motthe et al. 26 reported that the NPCs and BMSC transplanted by LP, one week after the surgery, were found at the lesion SCI site.

Baksi et al. 15 found that 1x10^6 BMSCs administered by a single LP in a mild SCI model had an effective migration and implantation in the injured site, was capable of reducing the cavity lesion when administered up to two weeks post-injury, and that larger volumes of cells infusions did not correlate with increased implantation. Nevertheless, Lepore et al. 12 reported, in a hemisection ablation model using a PL delivery, that the migration and implantation rates of NPCs was greater in the group receiving multiple transplantation than with a single transplantation. In the present study 10 BMMCs were injected at two different times of administration in the subarachnoid space by LP techniques and it was observed that the migration and the implantation of these cells could be verified, as well as the motor recovery.

Although these studies show a variety of neurological improvements and SC migration and implantation using the direct or subarachnoid approach, it is difficult to perform a direct comparison and come to a clear conclusion. This is due to the fact that some of these studies did not have control groups for the vehicle and the type of cell, 12 and the animal breed, the severity of injury 20,21 and the time of observation were different from those used in the present work. In our study, the histological data showed a similar lesional area between the BMMC and control group. Although there was no statistical difference between the groups, the group treated with BMMCs showed better motor recovery.

The difficulty in finding a relation between histology and neurological outcome could be one of the limitations of the present study. The histology techniques used had a low specificity to identify the myelin reorganization at the injury site when compared with a immunohistochemical study. This was consistent with the evidence that the reorganization of the spinal cord circuitry was independent of modifications in injury size and can explain the motor improvement. 27

Assuming that the DI route is the best route to infuse the SC, the translation to clinical practice could be of some concern. The reason is that using this technique spinal surgery was required, there was a risk of neurological deterioration due to damage to the spinal cord by needle puncture and the intramedullary injection of SC volume, higher costs, possible spinal deformity secondary to a laminectomy, difficulties in distinguishing the normal anatomy because of bone or muscle injuries or previous surgeries, greater chances of cerebrospinal fluid fistula, inherent clinical risks, such as deep vein thrombosis and pulmonary embolism and difficulty in closing muscle and skin due to previous scar tissue. 26 These problems may increase with subsequent injections, as further healing occurs. Thus, a study to verify the effectiveness of a minimally invasive route for SC delivery, like LP into the subarachnoid space is warranted.

The subarachnoid route allows multiple applications by minimally invasive technics which can supplant the theoretical advantage of the primer. In patients receiving transplants by the LP route, the complication that can be expected is a headache, although this is less likely to occur, since removal of spinal fluid is minimal. The present study showed that multiple applications of BMMCs via LP, in the acute stage of injury, may be an alternative for SCI treatment. 12

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