ABSTRACT

Objective: To evaluate the expression of matrix metalloproteinases and TGFβ in patients with spinal stenosis and in younger patients who have herniated disc. Methods: 19 samples of LA were analyzed, nine of them with lumbar canal stenosis and 10 with disc herniation. Of the total, five patients were aged between 15 and 40 years, 10 were between 40 and 65 years and four had more than 65 years. Representative areas of LF were chosen based on the staining of tissues with hematoxylin-eosin. The 3μm-thick sections embedded in paraffin and fixed in formalin were deparaffinized and rehydrated. All ligaments were incubated overnight at 4 °C with primary antibodies. Results: An increase of TGFβ was verified in older individuals, although without statistical significance. Conclusion: Metalloproteinases showed no significant difference between both groups with respect to age and type of abnormality of the spine.

Keywords: Ligamentum flavum; Metalloproteinases; Spinal stenosis.

INTRODUCTION

With the aging of the population, the incidence of diseases of the spine has increased, narrowing of the spinal canal being one of the main causes of pain and functional limitation in elderly patients. The cause of this narrowing may be due to facet arthrosis, disc bulging and in particular, ligamentum flavum hypertrophy (LFH).1 The factors related to hypertrophy of this ligament have been partially elucidated, and no effective prophylaxis or treatment option, except for decompression surgery, has been well established.

Histologically, the ligamentum flavum is composed of 70% elastic fibers and 30% collagen fibers, which are arranged parallel to each other in layers.2,3 During hypertrophy of the ligamentum flavum, there is reduction in the content of elastic fibers and an increase in collagen fibers, calcification, ossification and chondrometaplasia.4,5,6

Sairyo et al.7,8 have shown a correlation between the hardening of the ligamentum flavum and the degree of fibrosis, resulting in repetitive inflammatory processes due to the mechanical stresses that the ligament suffers during the movements of the spine. Park and collaborators demonstrated that an increase in TGFβ (transforming growth factor beta) expression, the protein that controls cell proliferation and acts in the early stages of oncogenesis, is related to hypertrophy of the ligamentum flavum.9

In the attempt to develop future therapeutic options, it will be necessary to understand whether the hypertrophy originates in the biomechanical stress caused by movement of the spine, or whether this process is due to inflammatory changes in the disc and adjacent tissues that increase the risk of inflammation of the ligamentum flavum, and its hypertrophy. According to this theory, patients with...
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a history of intervertebral disc disease would have a higher predisposition to develop changes in the ligamentum flavum in the future.

The objective of our study is to evaluate the expression of matrix metalloproteinases and TGF-B in the ligamentum flavum in patients with stenosis of the spinal canal, and in younger patients with other diseases of the spine, such as herniated disc.

METHOD

We studied 19 samples of ligamentum flavum (deep region) of individuals with stenosis of the lumbar canal and disc herniation, collected by the surgeon in the period May 2013 to January 2014, using nuclear magnetic resonance images of the segment of the lumbar spine to complement the diagnosis. These ligaments were obtained during surgery for the treatment of a herniated disc and lumbar canal stenosis. The study was approved by the Research Ethics Committee of the Faculdade de Medicina do ABC.

The subjects were divided into three groups according to age: under 40 years, between 40 and 65 years, and over 65 years. In relation to age: group I, 15 to 40 years, had five samples (average age: 29.2), group II, 40 to 65 years, had ten samples (average age: 48.4) and group III, over 65 years, had four samples (average age: 74). In relation to pathology: group I, lumbar canal stenosis, had nine patients and group II, disc herniation, had ten patients.

The inclusion criteria were:

- In the disc herniation group: extruded fragments located in the central-lateral region of the vertebral canal, more than three months of symptoms, without previous surgery of the lumbar segment and location of the disc herniation in segment L4-L5 or L5-S1.
- In the stenosis of the lumbar canal group: main complaint of neurogenic claudication, main factor of compression of the lumbar canal to hypertrophy of the ligamentum flavum, location of the stenosis in segment L4-L5 or L5-S1.

Immunohistochemistry

Representative areas of the ligamentum flavum were selected based on the staining of tissue sections with hematoxylin-eosin (HE). Sections 3 μm thick, embedded in paraffin and fixed in formalin, were deparaffinized and rehydrated.

Recovery of the antigen was performed by heating the slides to 100 °C for 30 minutes in a citrate buffer 10 mmol/L, pH 6.0. The endogenous peroxidase activity was blocked with an aqueous solution of 3% hydrogen peroxide, for 35 minutes.

The sections were then incubated overnight at 4°C with the primary antibodies: anti-decorin (N-15), anti-biglycan (L-15), TGFβ1 (sc-146), anti-MMP-9 (H-129), and anti-MMP-2 (H-76) (Santa Cruz Biotechnology, CA, USA). Finally, the slides were incubated with a complex of peroxidase-labeled streptavidin (LSAB®, DakoCytomation, Glostrup, Denmark) for 30 minutes. The sections were revealed using 3,3’-diaminobenzidine (DAB) for 1 minute and were contrasted with hematoxylin. Some samples were incubated with phosphate buffer 1 M in the absence of primary antibody, as negative controls. The presence of brown staining was considered as evidence of positive expression of the respective molecules in the cell.

Digital quantification

The slides were analyzed under a TS100 Nikon Eclipse® light microscope to identify areas that best represented the immunostaining of the molecules analyzed (hot spots). In each case, the quantification of immunostaining was performed by a method of digital analysis by computer. The 640 x 480 pixel photomicrographs were obtained from non-coincident consecutive fields at 400X magnification with a 4300 Nikon Coolpix® digital camera adjusted for the same parameters. The images were analyzed by the system for image processing and analysis ImageLab® (Softim Informática®, São Paulo, Brazil), adjusted to the micrometric scale (μm).

Index of positivity (IP)

In each case, at least 1,000 cells were counted by ImageLab®, and the observer classified as cells that are positive or negative. For this reason, the percentage of labeled cells was determined according to the following equation:

\[ IP = \frac{\text{number of labeled cells} \times 100}{\text{total cells counted}} \]

Intensity of expression (IE)

The ImageLab® was used to quantify the intensity of the brown color that resulted from the immunostaining. For each case, the same photomicrographs that were used to determine the IP were considered. Twelve cytoplasmic regions of different cells, randomly labeled, were accessed with the same-sized square (tool from the ImageLab® system). The mean optical density (OD) of these areas was automatically calculated and represents the average compositions of colors red, green and blue (RGB) per area of cytoplasm analyzed; the OD was expressed in optical units by square micrometer (ou/μm²). The same procedure was applied to obtain the background optical density (BOD) of an area without tissue or vascular space for each photomicrograph. A single area was sufficient for this purpose, because the background is homogeneous in each image. The absolute white color, which corresponds to the maximum optical density (320.7 ou/μm²) is composed of a complete mixture of red, green, and blue, while black represents the absence of these colors. Therefore, the optical density values calculated by the program comprised a decreasing scale, with the highest values corresponding to the colors that were clearly visible. The equation shown below was used to calculate the digital intensity of expression (IE) in each case. Their values comprised an increasing scale that is subtracted from the BOD proportional to the optical density of the absolute white.

\[ IE = 320.7 - \frac{320.7 \times \sum \text{OD}}{\sum \text{BOD}} \] [ou/μm²]

Expression index (EI)

The digital expression index (DEI) was obtained by multiplying the percentage of labeled cells (PLC) by the digital immunolabeling index (DII) for each case, using the following equation:

\[ EI = \frac{\text{IP} \times \text{IE}}{100} \] [ou/μm²]

RESULTS

In relation to age, there was an increase in TGF-B expression in patients aged over 65 years. Of metalloproteinases 2 and 9, only 9 showed an increase by age, but without statistically significant variation. (Figures 1-3)

The stenosis of the lumbar canal group presented higher TGF-B expression compared with the disc herniation group. (Figures 4-6) Metalloproteinases 2 and 9 showed greater expression in the disc herniation group compared with the lumbar canal stenosis group, but without statistically significant variation.

DISCUSSION

In neurogenic claudication, due to the osteoligamentary stenosis, there is a degenerative process that occurs in elderly patients. These changes cause pain, significant functional limitation, and neurological changes. The precise reason for the hypertrophy of the ligamentum flavum is little known, but the mechanism of mechanical stress is believed to be the main etiological factor; the precise reason for this progressive hypertrophy is still unknown. Neither do we know whether there is some factor that triggers these changes in the ligamentum flavum, besides mechanical stress; for example, inflammation of the surrounding tissues, such as the intervertebral disc.
Lohr et al. demonstrated a significant presence of inflammatory infiltrate in the peripheral region of the hypertrophied ligamentum flavum composed mainly of macrophages, endothelial cells and T lymphocytes, corresponding to a chronic immune reaction. These cells presented significant TGFβ expression, known as an important factor of arrangement of extracellular collagen. This inflammation may, therefore, be an important factor in the progression of hypertrophy of the ligamentum flavum, due to the replacement of elastic fibers by collagen fibers. Our study showed an increase in TGFβ expression in patients with stenosis of the lumbar canal with hypertrophy of the ligamentum flavum, but there was also an increase in TGFβ expression in individuals with lumbocytalgia due to lumbar disc herniation. Our hypothesis is that significant changes in the intervertebral disc can influence changes in the ligamentum flavum, but to clarify this, we would need to study different intervertebral disc changes and their relationship to the ligamentum flavum.

The research was carried out with a low number of samples, and does not show significant differences in the evaluation of the
material analyzed. The results may contribute to more advanced studies that elucidate the molecular mechanisms for the development of target molecules for new therapies, or other forms of diagnosis/prognosis of the process of degeneration of the ligamentum flavum.

The metalloproteinases are responsible for the degradation and modification of the extracellular matrix, and include more than 20 types. Metalloproteinases 2 and 3 are closely associated with degradation of the matrix of the joint cartilage and intervertebral disc. Soo and Kee-Young showed, in their study, that in degenerative spondylolisthesis, the mechanical stress to which the ligamentum flavum is submitted leads to an increase in metalloproteinase expression. This biomechanical stress occurs due to vertebral laxity and tensional instability of the ligament structures of the spine. In our study, there was an increase in expression of the metalloproteinases in both groups, with patients with disc herniation presenting a greater increase, compared with the stiffness of the lumbar canal group, adding the inflammatory factor to the biomechanical hypothesis of Soo and Kee-Young. Saiyoro et al detected the presence of inflammatory cytokines, such as COX-2 and interleukin 1, 6, 8 and 15 in ligaments with and without increased thickness. They also demonstrated, in their article, that RNAm expression of these inflammatory cytokines occurs before the actual ligament hypertrophy. We show, in our study, that in patients without hypertrophy of the ligamentum flavum, there was also an increase in metalloproteinases and TGFβ expression.

CONCLUSION

There was an increase in TGFβ in older individuals, although without statistical significance. The metalloproteinases did not present any significant differences between the groups, either in relation to age, or to the type of alteration in the spine.

All authors declare no potential conflict of interest concerning this article.