Antimicrobial activity of autoclaved and non autoclaved copaiba oil on *Listeria monocytogenes*

Atividade antimicrobiana de óleo de copaíba autoclavado e não autoclavado sobre *Listeria monocytogenes*

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ABSTRACT

The aim of this study was to evaluate the antimicrobial effect of different copaiba oil concentrations against the growth of *Listeria monocytogenes*, and analyze differences in inhibition of microorganisms with autoclaved and non autoclaved oil. This study provided an agar diffusion test with six isolates of bacteria and six different concentrations of autoclaved or non autoclaved copaiba oil and a negative control. The results showed sensitivity of five *L. monocytogenes* isolates related to the 10% autoclaved solution of copaiba oil. Four strains also showed sensitivity to the 5% autoclaved solution and one to 2.5% autoclaved solution. The 10% non autoclaved oil solution showed growth inhibition only for two strains. These results had pointed the 10% autoclaved solution of copaiba oil with higher inhibition as all other solutions and concentrations tested (*P*<0.05). For the other concentrations of both solutions, the 5 and 2.5% autoclaved and 10% non autoclaved solutions had presented statistically equal. All other concentrations of both copaiba solutions and the negative control did not presented any bacteria inhibition. In conclusion, the results of this study suggest that the autoclaved copaiba oil may be a potential new agent source for infection control or for food preservation, inhibiting the growth of food-borne bacteria such as *L. monocytogenes*.

Key words: phytotherapics, Copaifera, antimicrobial drugs, *Listeria monocytogenes*.

INTRODUCTION

*Listeria monocytogenes*, a food-borne pathogen, is the causative agent of listeriosis, a severe zoonotic disease that can lead to abortion, neurological disorders, sepsis and gastrointestinal disorders. It has been associated with foods such as raw milk, supposedly pasteurized milk, cheese, ice cream, raw vegetables, sausage meat, cooked and raw chicken.

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raw meat and raw or smoked fish. His ability to develop at temperatures as low as 3°C, allows its multiplication in refrigerated foods (NORRUNG et al., 2009). Through FoodNet data (CDC, 1989), is estimated that 2493 cases of food-borne listeriosis occur each year, with 499 deaths, which makes *L. monocytogenes* one of the five pathogens that cause more death in the U.S.A., accounting for 28% of deaths caused by diseases of dietary origin in that country (MEAD et al., 1999).

With the increase of bacterial resistance to antibiotics, there is considerable interest to investigate the antimicrobial effects of essential oils and different extracts against the bacteria range, to develop other classes of natural antimicrobials useful for the infection control or for food preservation (ALREZA et al., 2009).

The folk medicine has served as a source of compounds for use in the treatment of many diseases and combating several pathogens (PIERI et al., 2010; PIERI, et al., 2009). It has been highlighted, among the natural products in Brazil, the Copaiba (*Copaifera* sp.), that is a large tree that can reach up to 40 meters in height (ARAÚJO-JÚNIOR et al., 2005). Out of 72 species described worldwide by Index Kewensis, 16 are exclusive of Brazilian flora (HOOKER et al., 2009).

From the Copaiba tree is extracted an oil, called Copaiba oil, which has been used by more than 500 years in folk medicine with many potential properties for its use in medicine, as its healing properties like anti-inflammatory, antimicrobial and antisepsic action (PIERI et al., 2009).

This information obtained from folk medicine, indicating its use for many purposes, from most different kinds, for many years has been subject of several studies, to prove or adapt them to new therapies. In 1972, the Food and Drug Administration approved the Copaiba oil (FOOD AND DRUG ADMINISTRATION, 1972), after being subjected to tests for sensitization and irritation, using 25 volunteers, obtaining negative results for both (KLIGMAN, 1966).

Several studies have been conducted to identify microorganism’s sensitivity to copaiba oils as antimicrobials, to their further use in drug therapies for diseases or in preventing them (PACHECO et al., 2006; PIERI et al., 2010; VALDEVITE et al., 2007). Gram-positive bacteria have been constantly described as sensitive to this phytotherapic by researchers in this last years. The copaiba oil has many different substances that could act in different cell targets, acting synergistically in various structures and mechanisms of bacterial cell, resulting in a way to prevent or hinder the emergence of resistant bacteria (MENDONÇA & ONOFRE, 2009; PACHECO et al., 2006; PACKER & LUZ, 2007; PIERI et al., 2009; SANTOS et al., 2008).

The aim of this study was to evaluate the antimicrobial effect of different copaiba oil concentrations on the growth of *L. monocytogenes*, and analyze differences in microorganism’s inhibition with autoclaved and non autoclaved oil.

**MATERIAL AND METHODS**

The Copaiba oil (*Copaifera langsdorffii*) oil used here was exuded directly from the trunks of trees, according to PIERI et al. (2009). It was collected at Alfenas City, Minas Gerais State, Brazil, in September 2008 and the tree has the following geographic coordinates: 21°26’33”S and 46°0’55”W. After collecting, this material was maintained in amber bottle and stored at a temperature of ±4°C for later analysis.

Three solutions were used. The solution used as negative control was prepared with 10mL of tween 80 and 90mL of distilled water and than autoclaved. The first test solution consisted of a solution with 80mL destilled water and 10mL of tween 80 autoclaved and added with 10mL of copaiba oil non autoclaved. The second test solution was prepared with 10mL of copaiba oil, 10mL of tween 80 and 80mL of destilled water, and autoclaved after.

A *L. monocytogenes* (ATCC 7644) strain and five other isolates of same bacterium specie (LM1, LM2, LM3, LM4 and LM5) were used for the tests, obtained from carcasses in a slaughterhouse.

Adapting the technique of diffusion in agar cylinder plating (ESMERINO et al., 2004) the bacteria was grown in Petri dishes containing a thin layer of bacteriological agar no1 (HIMEDIA, Mumbai, India), and another layer above of Tryptic soy agar (TSA) (OXOID, Hampshire, United Kingdom) supplemented with 0.6% yeast extract (OXOID, Hampshire, United Kingdom). Orifices were made only in TSA distributed on each plate, performing a center orifice and other six orifices marginally and harmoniously distributed. Each of these orifices, with 5mm of diameter, was filled with a one different concentration of test solutions serially diluted between 10% and 0.3125% of copaiba oil. The negative control solution was used into the center orifice of all plates.

The bacteria were inoculated on the TSA surface using a sterile swab, and each orifice was filled with its solution. Once prepared, the plates were incubated at 37°C for 24h. The antimicrobial activity was identified by the presence of any inhibition halo around the orifice. The inhibition halo was measured by the diameter length of inhibition halos to compare the activity concentrations. The tests were performed in triplicate.

RESULTS AND DISCUSSION

Many natural oils had been tested and presented good results in *L. monocytogenes* inhibition as *Cestrum nocturnum* L. (ALREZA et al., 2009), *Origanum vulgare* (GUTIERREZ et al., 2009; MUÑOZ et al., 2009), *Lonicer japonica* Thunb (RAHMAN & KANG 2009), *Thymus vulgaris*, *Melissa officinalis* (GUTIERREZ et al., 2009) and *Rosmarinus officinalis* (MUÑOZ et al., 2009). These natural drugs can present many active principles acting in different structures in bacterial cells, hindering the emergence of resistant strains, then presenting a great advantage over conventional antimicrobial drugs.

The results of this research showed sensitivity of *L. monocytogenes* strains related to the 10% autoclaved solution of copaiba oil, except the isolate LM3 which was resistant to all concentration of both copaiba oil solutions. The isolates LM1, LM2, LM4 and LM5 also showed sensitivity to the 5% autoclaved solution. The LM4 also showed sensitivity to the 2.5% autoclaved solution. The 10% non autoclaved oil solution showed growth inhibition only for LM4 and LM5 isolates. This growth inhibition of *L. monocytogenes* by a phytotherapeutic points some advantages over the emergence of resistant bacteria due to possible different action mechanisms performed by many active substances present in the oil, which could inhibit the bacterial resistance against the antimicrobial.

These results had pointed the 10% autoclaved solution of copaiba oil with higher inhibition as all other solutions and concentrations tested (P<0.05). For the other concentrations of both solutions, the five and 2.5% autoclaved and 10% non autoclaved solutions had presented statistically equal. No other concentration of both solutions and the negative control presented inhibition halo. Table 1 presents the mean of inhibition halos length for the solutions that present some activity against each *L. monocytogenes* strains. The other concentrations did not show microbial inhibition, showing no inhibition halo, as well as the negative control. No article had described yet a comparison between autoclaved or non autoclaved copaiba oil different actions against any microorganism. It was supposed that the heat treatment could cause a change in the molecular structures that has given or potentiated the antimicrobial activity of the oil. This could explain the different results between the solutions.

Although there are no reports of antimicrobial testing of this oil against *L. monocytogenes*, many studies have shown antimicrobial effect against several other bacterial species such as *Staphylococcus epidermidis* (SANTOS et al., 2008), *S. aureus*, *Bacillus subtilis* (PACHECO et al., 2006; SANTOS et al., 2008), *Streptococcus mutans*, *S. pyogenes*, *S. salivarius*, *Enterococcus faecalis* among others (PIERI et al., 2010; VALDEVITE et al., 2007).

Study has showed a synergistic effect, against *Listeria* species, with the combination of terpenic fractions of citrus oil and organic acids as malic acid and ascorbic acid (FRIEDLY et al., 2009). The copaiba oil, which is composed by a sesquiterpenic and terpenic fractions, may has a similar action in combination with organic acids. Thus some authors present a possible modification in the *Listeria* sensitivity to antimicrobial related to pH and other physicochemical alterations (NAÏTALI et al., 2009).

CONCLUSION

In conclusion, the results of this study suggest that the autoclaved copaiba oil may be a potential new agent source to the infection control or for food preservation, inhibiting the growth of food-borne bacteria such as *L. monocytogenes*.

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<thead>
<tr>
<th>Listeria monocytogenes</th>
<th>Autoclaved solution 10%</th>
<th>Autoclaved solution 5%</th>
<th>Autoclaved solution 2.5%</th>
<th>Non autoclave solution 10%</th>
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<tr>
<td>ATCC 7644</td>
<td>9mm</td>
<td>R</td>
<td>R</td>
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<tr>
<td>LM1 isolate</td>
<td>12mm</td>
<td>10mm</td>
<td>R</td>
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<td>LM2 isolate</td>
<td>13mm</td>
<td>11mm</td>
<td>R</td>
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<td>LM3 isolate</td>
<td>R</td>
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<td>LM4 isolate</td>
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<td>LM5 isolate</td>
<td>11mm</td>
<td>11mm</td>
<td>10mm</td>
<td>8mm</td>
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</table>

Table 1 - Means of inhibition halos (in mm) produced by different solutions of copaiba oil against six *L. monocytogenes* strains (one ATCC an five slaughterhouse isolates). Any other concentrations of these solutions present microbial inhibition. R is presented resistance of microorganism to the solution.
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