ABSTRACT

To evaluate the effect of supplementation with mannan oligosaccharides, avilamycin and halquinol, alone or in combination, on the performance, carcass characteristics and antibody production in broilers (1-49 days old), male broiler chicks (n=1440; Cobb 500; one day old) were housed and distributed into a completely randomized design into six treatments (eight replicates; 30 animals per pen). To produce the experimental diets, three types of performance enhancer additives were used. Halquinol (HAL), avilamycin (AVI) and mannan oligosaccharides (MOS) were included (alone or in combination) in the basal diet (instead of corn starch). Effects of diet were observed on results of animal performance in the period 1-21 and 1-42 days old. Broilers fed with a diet without growth promoter showed lower weight gain in relation to those fed with diets with antimicrobials, MOS or a combination of them. In the period 1-49 days old, feed conversion increased in broilers fed with rations without promoter. At the end of the experimental period no influence of diets was observed on the carcass yield and cuts, and titles of specific antibodies to avian infectious bronchitis. The use of MOS and/or antimicrobials (AVI or HAL), alone or in combination, improves feed conversion of broilers reared until 49 days of age.

Key words: avilamycin, additive, halquinol, mannan oligosaccharides.

INTRODUCTION

Currently, there is a major concern in the society with the use of antimicrobial agents in livestock feed, and such chemicals can be added to the feed as either therapeutic agents or performance enhancers. As enhancers, lower antimicrobial doses are used for longer periods, which can potentiate selection of resistant bacteria (BAURHOO et
Mannan oligosaccharides (MOS) are yeast wall-derived prebiotics that can bind to the fimbriae of pathogenic Gram-negative bacteria such as *E. coli* and *Salmonella* spp. Such binding causes pathogenic bacteria to be eliminated from the intestinal lumen, as they cannot adhere to the epithelial surface of enterocytes. Benefits of including MOS in diets for broiler chickens come from the change in their intestinal microbiota and decrease in the turnover rate of their intestinal tract. Besides, addition of MOS results in an increased production of immunoglobulins (Ig) in broilers and laying hens (SHASHIDHARA & DEVEGOWDA, 2003). Despite these benefits, the use of MOS instead of antimicrobials as performance enhancer has shown inconsistent results (ALBINO et al., 2006; BAURHOO et al., 2009; KIM et al., 2011). Thus, the objective of this study was to evaluate the effect of supplementation with MOS, avilamycin, and halquinol, alone or in combination, on performance, carcass characteristics and antibody production in broilers in the period 1-49 days old.

**MATERIALS AND METHODS**

The experiment was conducted in an experimental farm (Centro de Tecnologia Animal Ltda./Center for Animal Technology Ltd.) located in the city of Domingos Martins, state of Espírito Santo (ES), Brazil. Male broiler chicks (n=1440; Cobb 500 strain; one-day old) were used. Birds were vaccinated against the Marek’s and avian Bouba diseases.

The experiment was conducted in a facility with concrete floor; covered with fiber cement tiles (3.0-m height ceiling); lateral short walls (0.5-m height), closed laterally with wire mesh (3.0cm), adjustable external curtains (braided polyethylene), at experimental unit (1.5 x 1.5m) to approach the challenges to the maximum, which usually occur during poultry growth.

In order to promote a conventional management in a commercial farm, the birds were housed in boxes with wood shaving beds (10-cm thick), which were reused for two consecutive batches of broiler raising. A continuous (24-h; natural and artificial) light program was used.

In each phase (Table 1), the basal diet without performance enhancers was formulated based on corn and soybean meal, and supplemented with industrial amino acids to meet the aminoacidic proportions recommended by ROSTAGNO et al. (2011) based on the ideal protein. Feed and water were provided ad libitum.

Broilers were distributed in a completely randomized design into six treatments, eight replicates and 30 birds per pen. Three types of performance enhancing additives, halquinol (HAL), avilamycin (AVI) and mannan oligosaccharides (MOS) were added to the diets, alone or in combination, replacing corn starch to compose the experimental diets in the growth (1-21 days), termination (22-42 days) and pre-slaughter (43-49 days) phases. The experimental diets were composed according to table 2. The performance data were obtained in the cumulative periods 1-21, 1-42, and 1-49 days old. They are as follows: weight gain, feed intake (obtained by the difference between total feed provided and leftovers collected at the end of each period, based on the average number of birds), feed conversion (calculated by dividing the total feed consumed by the weight gain, corrected for the weight of dead birds), viability (calculated by dividing the total chickens withdrawn by the chicks received, multiplied by 100), and productive efficiency index (PEI; calculated by multiplying the daily weight gain by viability, divided by the feed conversion, and multiplied by 100).

At the 49th day, after six hours of feed fasting, two birds were slaughtered by cervical dislocation and selected according to the average weight of each pen (±10% of the average weight) to evaluate the carcass yield and prime cuts (breast, drumstick, and thigh). The carcasses were weighed without head, feet, viscera and abdominal fat. The
Table 1 - Percentage and calculated composition of basal diet.

<table>
<thead>
<tr>
<th>Ingredients (%)</th>
<th>1-21 days</th>
<th>22-42 days</th>
<th>43-49 days</th>
</tr>
</thead>
<tbody>
<tr>
<td>Corn bran</td>
<td>52.40</td>
<td>58.37</td>
<td>72.942</td>
</tr>
<tr>
<td>Soy bean meal</td>
<td>40.15</td>
<td>34.451</td>
<td>22.047</td>
</tr>
<tr>
<td>Soybean oil</td>
<td>3.630</td>
<td>4.177</td>
<td>2.720</td>
</tr>
<tr>
<td>Phosphate, dicalcium</td>
<td>1.640</td>
<td>1.204</td>
<td>0.788</td>
</tr>
<tr>
<td>Limestone, calcite</td>
<td>0.930</td>
<td>0.757</td>
<td>0.626</td>
</tr>
<tr>
<td>Sodium chloride (common salt)</td>
<td>0.490</td>
<td>0.395</td>
<td>0.361</td>
</tr>
<tr>
<td>L-Lysine.HCl (78%)</td>
<td>0.150</td>
<td>0.141</td>
<td>0.202</td>
</tr>
<tr>
<td>DL-Methionine (98%)</td>
<td>0.300</td>
<td>0.232</td>
<td>0.148</td>
</tr>
<tr>
<td>L-Threonine (98%)</td>
<td>0.040</td>
<td>0.015</td>
<td>0.016</td>
</tr>
<tr>
<td>1^Premix, Vitamin</td>
<td>0.120</td>
<td>0.100</td>
<td>0.100</td>
</tr>
<tr>
<td>2^Premix, Mineral</td>
<td>0.050</td>
<td>0.050</td>
<td>0.050</td>
</tr>
<tr>
<td>Cornstarch</td>
<td>0.100</td>
<td>0.100</td>
<td>0.100</td>
</tr>
<tr>
<td>Avilamycin (10%; AVI)</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Halacinol (60%; HAL)</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>3^Mannan oligosaccharide (MOS)</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

*Calculation composition

Energy, metabolizable, kcal/kg | 3,020 | 3,174 | 3,250 |
Protein, crude, % | 22.60 | 21.00 | 16.60 |
Lysine, digestible, % | 1.250 | 1.100 | 0.862 |
Methionine + Cystine, digestible, % | 0.900 | 0.804 | 0.629 |
Threonine, digestible, % | 0.810 | 0.715 | 0.560 |
Valine, digestible, % | 0.960 | 0.858 | 0.672 |
Tryptophan, digestible, % | 0.259 | 0.229 | 0.164 |
Isoleucine, digestible, % | 0.897 | 0.802 | 0.598 |
Leucine, digestible, % | 1.752 | 1.624 | 1.359 |
Arginine, digestible, % | 1.451 | 1.291 | 0.947 |
Glycine + Serine, digestible, % | 1.908 | 1.368 | 1.313 |
Calcium, % | 0.870 | 0.717 | 0.528 |
Available Phosphorus (%) | 0.420 | 0.335 | 0.246 |
Sodium (%) | 0.210 | 0.198 | 0.180 |

1^Vitamin supplements (per kg of product): Vitamin A: 9,000,000 I. u., Vitamin D: 2,500,000 I. u., Vitamin E: 20,000mg, Vitamin K: 2500mg, Vitamin B1: 1500mg, Vitamin B2: 6000mg, Vitamin B6: 3000mg, Vitamin B12: 12mg, Folic acid: 800mg, Nicin: 25,000mg, Pantothenic acid: 12,000mg, Selenium: 250mg, Biotin: 60mg. 2^Mineral supplements (per kg of product): Copper: 20,000mg, Iron: 100,000mg, Iodine: 2000mg, Manganese: 160,000mg, Zinc: 100,000mg, Cobalt: 2000mg. 3^Actigen®, 280kg kg^-1 crude protein (100% yeast cell wall).

RESULTS AND DISCUSSION

An effect (P<0.05) of feed was observed on the results of animal performance in the period 1-21 days old (Table 3). Birds fed with diet without performance enhancer presented lower weight gain and worse feed conversion compared to those fed with diets with antimicrobials, MOS or a combination of them. Similarly, ALBINO et al. (2006) reported a decrease (P<0.05) in weight gain of birds fed without performance enhancer compared to those fed with diet containing AVI (7ppm). However, the same authors observed no increase in weight gain with addition of MOS (0.2%). BAURHOO et al. (2009) reported no improvement in weight gain, feed intake and feed conversion with addition of virginiamycin (16.5mg kg^-1) and bacitracin (55.0mg kg^-1) or MOS (0.2 and 0.5%) compared to the control diet in the period 1-21 days old.

Regarding feed conversion, SMITH (2011) reported that use of diet without antibiotics decreases uptake of nutrients and reduces weight gain. Results of the present study are in agreement with those of SMITH (2011), since lower weight gain and increased feed conversion (P<0.05) were observed in animals that consumed diet without performance enhancer. The use of the same chicken litter in two consecutive batches may be linked to such lower performance. Because of worsening in weight gain and feed...
conversion a decrease (P<0.05) was observed in the productive efficiency index.

However, the results of various authors (ALBINO et al., 2006; BAURHOO et al., 2009; KIM et al., 2011) are inconsistent with each other relative to the use of performance enhancers. Such differences between results may be associated with the degree of sanitation on the farm, specificity of the performance enhancer, dose, age, type of bacterial challenge (quantity, species, and virulence) and management.

Despite the lower weight gain and worsening in feed conversion in animals fed control diet, difference (P>0.05) was not observed in feed intake between treatments, except between the association of AVI + MOS, which differed (P<0.05) from the control diet. This lower feed intake could be related to increased absorption of nutrients (energy) compared to the control diet, since feed conversion was also lower (1.34 vs 1.73). According to LEESON et al. (1996) the broiler possesses a good ability to control its feed intake based on energy intake.

An effect of (P<0.05) diets was observed on weight gain and viability in the period 1-42 days old (Table 3). Birds fed control diet presented weight gain and viability lower than those of (P<0.05) birds fed with antimicrobial diets, MOS or their combination. Similar results for weight gain were obtained by ALBINO et al. (2006) with addition of AVI in the periods 1-21 (7ppm) and 22-42 (5ppm) days old and use of MOS (0.2%), alone or in combination with AVI. Conversely, OLIVEIRA et al. (2009) reported that difference was not found between feed added with MOS or antimicrobials in the performance of broilers at 42 days of age. MUNYAKA et al. (2012) also reported no effect on the performance of broiler chickens with the use of MOS or bacitracin compared to the control diet in the 1-42 day old period.

In feed conversion, although difference was not reported between control diet and diets...
containing the antimicrobial AVI and MOS alone in the 1-42 day of life, period it was possible to verify that combination of different performance enhancers (AVI + MOS, AVI + HAL, and AVI + MOS + HAL) reduced (P<0.05) feed conversion in relation to the diet without the additives.

Reduction in feed conversion probably was possible due to the inclusion of performance enhancers in the diet and their synergistic action on different pathogenic microorganisms that can colonize the gastrointestinal tract. Pathogenic bacteria can present different degrees of virulence and resistance to antimicrobials, and addition to the diet of one or more enhancers with different mechanisms of action is necessary.

Avilamycin acts preferentially on Gram-positive bacteria and Halquinol have activity against a wide variety of both Gram-positive and Gram-negative bacteria. Mannan oligosaccharides have action on Gram-negative bacteria that express type-I fimbriae, such as Salmonella spp. and Escherichia coli. Thus, additives which act on different pathogenic microorganisms can promote a synergistic effect on development of broilers.

Regarding on the action of Mannan oligosaccharides MUNYAKA et al. (2012) stated that the differences between results reported in the studies may be due to differences between experimental protocols, production phase, conditions in which the experiment was conducted, source, type, and concentration of MOS used in the additive.

In the total period (1-49 days) of the experiment, effect (P>0.05) of feed on the weight gain and feed intake by the animals was not observed (Table 3). However, feed conversion, viability and PEI worsened (P<0.05) in birds fed with control diet compared to other treatments, which in turn were similar to each other.

The improvement in feed conversion in animals consuming MOS, antimicrobials or combinations may be due to a more efficient use of nutrients, since no significant difference was observed in weight gain and feed intake. According to MILES et al. (2006), improvement in animal performance with the use of performance enhancer antimicrobials is due to a greater availability of energy for growth and increase in absorption of nutrients, which, in turn, result from reduction in cell proliferation and smaller thickness of mucosa and lamina propria in the intestinal wall.

Effect (P>0.05) of feed on the carcass and cut yields was not observed (Table 4). Similar results were observed by BAURHOO et al. (2009) who compared feed without performance enhancer and with addition of virginiamycin (16.5mg kg⁻¹), bacitracin (55.0mg kg⁻¹) and MOS (0.2 or 0.5%) reported no difference between the yields of carcass, breast, fillet, drumstick, thigh, and wing at 38 days of age. According to the authors, the similarity between the responses to additives on the final weight of animals may have reduced the effects on carcass and cut yields. Even the improvement in feed conversion (P<0.05) of animals, which consumed diets with performance enhancers, was not enough to increase their muscle deposition. Similar results were reported by various authors (WALDRoup et al., 2003; PARKS et al., 2005; BAURHOO et al., 2009). Probably, adjustments in the nutritional levels (amino acids and energy) will be necessary to optimize the carcass and cut yields in animals receiving performance enhancers.

Effect (P>0.05) of titers of specific antibodies to avian infectious bronchitis on the treatments was not observed at the end of the experimental period (Table 4). This result is similar to that observed by KIM et al. (2011), who evaluated addition of AVI (6.0mg kg⁻¹) and MOS (0.025 or 0.050%) and reported no significant differences in the IgG and IgA production in birds with four weeks old.

According to SHASHIDHARA & DEVEGOWDA (2003), diet supplementation with MOS influences the immune system resulting in weight gain and feed intake. According to MILES et al. (2006), improvement in animal performance with the use of performance enhancer antimicrobials is due to a greater availability of energy for growth and increase in absorption of nutrients, which, in turn, result from reduction in cell proliferation and smaller thickness of mucosa and lamina propria in the intestinal wall.

**Table 4 - Effect of diets on carcass and cut yields, and antibody titers to avian infectious bronchitis at 49 days of age.**

<table>
<thead>
<tr>
<th>Variables</th>
<th>Without enhancer</th>
<th>AVI</th>
<th>MOS</th>
<th>AVI + MOS</th>
<th>AVI + HAL</th>
<th>AVI + MOS + HAL</th>
<th>P values</th>
<th>CV (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Carcass</td>
<td>73.1</td>
<td>72.9</td>
<td>72.5</td>
<td>73.0</td>
<td>72.4</td>
<td>72.6</td>
<td>0.145</td>
<td>2.32</td>
</tr>
<tr>
<td>Chest</td>
<td>36.9</td>
<td>37.8</td>
<td>36.9</td>
<td>36.0</td>
<td>36.9</td>
<td>38.2</td>
<td>0.063</td>
<td>7.45</td>
</tr>
<tr>
<td>Drumstick</td>
<td>15.0</td>
<td>14.5</td>
<td>15.0</td>
<td>14.9</td>
<td>14.9</td>
<td>14.7</td>
<td>0.062</td>
<td>6.09</td>
</tr>
<tr>
<td>Thigh</td>
<td>15.8</td>
<td>15.5</td>
<td>14.2</td>
<td>15.5</td>
<td>16.2</td>
<td>16.5</td>
<td>0.067</td>
<td>15.21</td>
</tr>
<tr>
<td>Titters</td>
<td>2.89</td>
<td>2.45</td>
<td>2.81</td>
<td>2.89</td>
<td>2.77</td>
<td>2.72</td>
<td>0.189</td>
<td>19.46</td>
</tr>
</tbody>
</table>

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in a significant increase in antibody production. According to these authors, improvement in immune system activation occurs due to an improvement in the absorption of some nutrients such as Zn, Cu, and Se or all of them both. MUNYAKA et al. (2012) attribute such improvement in the immune system to an increase in the activity of the T and B cells.

Despite the positive effects of the use of MOS on the immune system reported by several authors (SHASHIDHARA & DEVEGOWDA, 2003; MUNYAKA et al., 2012) in the present study was not observed significant difference (P>0.05) between treatments on the levels of IgG to infectious bronchitis. Probably the use of wood shaving beds between treatments on the levels of IgG to infectious bronchitis. Probably the use of wood shaving beds was not observed significant difference (P>0.05) in the production of immunoglobulins.

CONCLUSION

The supplementation of MOS and/or antimicrobial agents (AVI and HAL) alone or in combination no improves carcass characteristics and antibody production in broilers raised until 49 days of age. The feed conversion, viability and productive efficiency index improves with use of MOS and/or antimicrobial agents (AVI and HAL) alone or in combination in the period of 1-49 days of age.

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