Optimization of crude protein in diets for Nile tilapia reared in net pens: performance, hematology, and water quality

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ABSTRACT: Two experiments were conducted to evaluate the effects of reducing dietary crude protein (CP), based on the ideal protein concept for Nile tilapia reared in net pens. The experimental (isocaloric, isocalcium, and isophosphoric) diets were formulated to contain 270, 300, 330, and 360g kg⁻¹ CP. In experiment 1, 4320 Nile tilapia (13.5±0.82g) were used to evaluate the performance and hematological parameters. The experimental design was completely randomized, and the fish were distributed in 24 net pens (1.0m³), with four diets, six replicates, and 180 fishes per experimental unit. In experiment 2, 40 Nile tilapia (22.5±0.56g) were used to evaluate the ammonia excretion. Fish were distributed in 40 aquaria (3.0L), with one fish per aquarium (n=10). No protein reduction effect was observed on feed intake and the values of hematocrit and hemoglobin. Regarding the statistic models used in the present study, difference was observed between CP values. The optimal level estimated by the quadratic equation first interception with the linear response plateau (LRP) as a response to CP changes in the diet was determined for weight gain (324.3g kg⁻¹) and feed conversion (317.8g kg⁻¹). After reduction in the CP levels, a linear reduction was observed in the ammonia excretion in water. Based on the ideal protein concept for Nile tilapia reared in net pens, reducing the CP levels in the diets is possible, without change in the performance and hematological parameters, and with a reduction in the levels of ammonia excretion in water, since amino acids are supplemented.

Key words: amino acids, performance, statistic models, Oreochromis niloticus, environmental pollution.

RESUMO: Dois experimentos foram realizados para avaliar os efeitos da redução na quantidade de proteína bruta (PB) da ração, com base no conceito de proteína ideal para tilápias do Nilo criadas em tanques-rede. As rações experimentais (isoenergéticas, isocálcicas e isofosfóricas) foram formuladas para conter 270, 300, 330 e 360g kg⁻¹ de PB. No experimento 1, 4320 tilápias do Nilo (13.5±0.82g) foram usadas para avaliar os parâmetros de desempenho e hematológicos. O delineamento experimental foi inteiramente casualizado e os peixes foram distribuídos em 24 tanques (1.0m³), com quatro rações, seis repetições e 180 peixes por unidade experimental. No experimento 2, 40 tilápias do Nilo (22.5±0.56g) foram usadas para avaliar a excreção de amônia. Os peixes foram distribuídos em 40 aquários (3.0L), com um peixe por aquário (n=10). Nenhum efeito de redução proteica foi observado sobre o consumo de ração e os valores de hematocrito e hemoglobina. Em relação aos modelos estatísticos empregados neste estudo, uma diferença foi observada entre os valores de PB. O nível ótimo, estimado através da primeira intersecção da equação quadrática com o platô do modelo Linear Response Plateau (LRP) como resposta às variações na proteína da dieta, foi determinado para ganho em peso (324.3g kg⁻¹) e conversão alimentar (317.8g kg⁻¹). Após a redução nos níveis de PB, foi observada uma redução linear na excreção de amônia na água. Com base no conceito de proteína ideal para tilápias do Nilo criadas em tanques-rede, é possível reduzir os níveis de PB nas rações, sem alteração nos parâmetros de desempenho e hematológicos, e com redução nos níveis de excreção de amônia na água, mediante suplementação de aminoácidos.

Palavras-chave: aminoácidos, desempenho, modelos estatísticos, Oreochromis niloticus, poluição ambiental.

INTRODUCTION

Tilapia is a fish species that has best adapted to the intensive production system. In addition, tilapia has a quick development cycle, firm flesh, and good acceptance in the market. However, the challenges to improve production efficiency and reduce costs are constant.

In fish feed, protein is the nutrient that most influences the production cost. In addition, proteins can become an environmental problem if levels above the animal’s utilization capacity are used. Taking into account that fish requirements are not for protein but for a suitable amino acid ratio to form muscle protein and other body proteins, the amino acid excess will be removed by fishes in the
form of ammoniacal nitrogen, which may promote water eutrophication, reduction in growth rate, and increase in the production cost (FURUYA et al., 2005; BOTARO et al., 2007; RIGHETTI et al., 2011).

Usually, the protein levels in commercial diets for tilapia are above their needs, which means high participation of protein ingredients in the formulation (FURUYA et al., 2001). Source of proteins used in the formulation may be of vegetable or animal origin. However, the nutrients should promote optimal performance, reducing both costs and environmental pollution (PEZZATO et al., 2009; FURUYA & FURUYA, 2010b). To achieve this goal, determining the levels and availability of the amino acids present in each ingredient is necessary. Therefore, diets should be formulated based on amino acid requirements (STOREBAKKEN et al., 2000; BOTARO et al., 2007; BOMFIM et al., 2008; FURUYA & FURUYA, 2010b).

Reduction in crude protein content and supplementation with synthetic amino acids are possible strategies for better balanced amino acid formulations in diets (FURUYA & FURUYA, 2010b). Several authors have confirmed that this strategy is effective in reducing nitrogen excretion without impairing the general performance (BALLESTRAZZI et al., 1994; FURUYA et al., 2005). However, most of these studies were conducted with fish in laboratory conditions, and experiments evaluating crude protein reduction in net pens and intensive production systems are scarce. Under field conditions, factors such as density, stress, age, size, temperature, feeding, and oxygen level can affect reduction in the amount of protein diet, which either alone or together can affect fish performance.

Therefore, based on the ideal protein concept, the objective of this study was to evaluate the effect of reducing the amount of crude protein provided to Nile tilapia on the performance, hematological, and water-quality parameters using different mathematical models.

MATERIALS AND METHODS

Two experiments were conducted with Nile tilapia (Oreochromis niloticus; Thailand strain) to evaluate the performance, hematological, and water-quality parameters of fish fed diets containing different amounts of crude protein. Treatments consisted of four experimental isocaloric, isocalcium, and isophosphoric diets containing 270, 300, 330, and 360 g kg\(^{-1}\) of crude protein (CP). The diets (Table 1) were formulated according to FURUYA (2010a) to meet the minimum nutritional requirements (except for CP), as recommended for Nile tilapia (≤100 g). The CP values of the experimental diets were obtained by adjusting the amount of fish and corn flour. Synthetic amino acids L-lysine, DL-methionine, L-threonine, and L-tryptophan were provided as a supplement when their ratios with digestible lysine were lower than those obtained in the diet containing a higher CP level.

After the ingredients were mixed, the mixture was extruded in a company (Nutriave Alimentos) using a single screw extruder (Ferraz, E-200 model; engine power: 200 hp; nominal capacity: 4.5 ton h\(^{-1}\)) followed by drying in a horizontal belt dryer (140±12°C; 22 min).

In experiment 1, the performance and hematological parameters were analyzed. The study was conducted at the Acquapeixe farm, which is installed in the Aguiar lagoon, in the Baixo Quartel district of Linhares city, Espírito Santo State (ES), Brazil. A total of 4320 tilapia (initial weight: 13.5±0.82 g) were distributed in 24 net pens (1.0 m\(^3\); 1x1x1 m). The experimental design was completely randomized, with four diets, six replications, and 180 fish per experimental unit. Diets were weighed and provided daily in three feeds (8:00, 12:00, and 16:00) for 50 days. Thus, each meal was offered in small amounts, with repeated administration (by manual feeding) until apparent satiation, when pellet capture and regurgitation by the fish was no longer observed. Animals were weighed on a digital scale (precision: 0.01 g) at the beginning and end of the experiment to determine both weight gain and feed conversion.

Water temperature in the net pens was measured daily (8:00 and 18:00) and pH and dissolved oxygen (mg L\(^{-1}\)) were measured (deep: 50 cm) every two weeks with a portable digital multiparameter meter (Akso; model AK 87).

At the end of the experimental period, six fish per replicate were used for hematological analysis. Fish were anesthetized (benzocaine; 1 g 15 L\(^{-1}\) of water) and, after complete desensitization, blood was collected by puncture of the caudal vein with a syringe (3 mL) coated with 3% ethylenediamine tetra-acetic acid (EDTA). Samples were sent to the laboratory of veterinary clinical pathology (Biolabor Veterinário\(^{b}\)), where the values for hemoglobin, hematocrit, and plasma protein were determined.

In experiment 2, ammonia excreted by fish was determined in the Laboratory of Applied Ichthyology, Universidade de Vila Velha (UVV). A total of 40 tilapia of the Thai strain (weight: 22.5±0.56 g) were used. Fish were acclimatized for a week in a tank (100 L) under constant aeration, being

fed (to satiation; once daily) a diet containing 270g kg\(^{-1}\) protein. After this period, they were anesthetized (benzocaine solution; 0.1g L\(^{-1}\)), weighed, and transferred to 40 individual three-liter aquariums; (n=10) being kept under constant aeration and fed the experimental diets (270, 300, 330, and 360g kg\(^{-1}\) CP) for a week. After this period, they were subjected to fasting (48h) for excreta disposal. After fasting, fish were again fed with the experimental diets (to satiation) and then transferred to aquaria containing fresh water. To calculate the amount of ammonia, water was analyzed before receiving the fish and 24h after they were received in the new tank. Ammonia was determined by the indophenol method, and measured using a spectrophotometer (Biospectro; SP-220; Curitiba, PR, Brazil; \(\lambda = 640\)nm) according to the American Public Health Association (APHA, 1998). Total excretion was calculated using the ALTINOK & GRIZZLE (2004) equation: Total excretion = \{[(NH\(_f\)/U\(_f\) - NH\(_i\)/U\(_i\)] / g\(_{total}\)\} / 3, where NH\(_i\)/U\(_i\) is the calculated sample final ammonia, NH\(_i\)/U\(_i\) is the initial ammonia, which is divided by the mean biomass of

<table>
<thead>
<tr>
<th>Ingredients</th>
<th>Crude protein, g kg(^{-1})</th>
<th>Crude protein analyzed</th>
<th>Digestible energy (kcal kg(^{-1}))</th>
<th>Ether extract</th>
<th>Starch</th>
<th>Crude fiber</th>
<th>Total calcium</th>
<th>Available phosphorus</th>
<th>Digestible lysine</th>
<th>Digestible methionine + cystine</th>
<th>Digestible threonine</th>
<th>Digestible tryptophan</th>
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<tr>
<td>Soybean meal</td>
<td>225.26</td>
<td>225.26</td>
<td>3100</td>
<td>312.6</td>
<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
<td>3.0</td>
<td>3.0</td>
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<td>312.6</td>
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<td>16.0</td>
<td>15.3</td>
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<td>14.7</td>
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<td>137.44</td>
<td>3100</td>
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<td>16.0</td>
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<td>10.1</td>
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<td>10.1</td>
<td>14.7</td>
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<td>14.7</td>
<td>3.0</td>
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<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
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<td>14.7</td>
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</tr>
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<td>0.90</td>
<td>3100</td>
<td>312.6</td>
<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
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<td>3.0</td>
<td>3.0</td>
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<tr>
<td>L-Tryptophan</td>
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<td>0.10</td>
<td>3100</td>
<td>312.6</td>
<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
<td>3.0</td>
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<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
<td>3.0</td>
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<tr>
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<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
<td>3.0</td>
<td>3.0</td>
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<tr>
<td>Vitamin premix(^1)</td>
<td>2.00</td>
<td>2.00</td>
<td>3100</td>
<td>312.6</td>
<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
<td>3.0</td>
<td>3.0</td>
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<tr>
<td>Mineral premix(^1)</td>
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<td>2.00</td>
<td>3100</td>
<td>312.6</td>
<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
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<td>Vitamin C</td>
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<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
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<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
<td>3.0</td>
<td>3.0</td>
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<tr>
<td>Antifungal agent(^3)</td>
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<td>0.50</td>
<td>3100</td>
<td>312.6</td>
<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
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<td>3.0</td>
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<tr>
<td>Antioxidant agent (BHT)(^4)</td>
<td>0.30</td>
<td>0.30</td>
<td>3100</td>
<td>312.6</td>
<td>20.9</td>
<td>16.0</td>
<td>15.3</td>
<td>14.7</td>
<td>10.1</td>
<td>14.7</td>
<td>3.0</td>
<td>3.0</td>
</tr>
</tbody>
</table>

\(^1\)Content (kg\(^{-1}\)): Vitamin A: 1,333,333UI; Vitamin D3: 333,333UI; Vitamin E: 10,000mg; Vitamin K3: 666mg; Vitamin B1: 1666mg; Vitamin B2: 1666mg; Vitamin B6: 1333mg; Vitamin B12: 3.33µg; Vitamin C: 35,333mg; Pantothenic acid: 2500mg; Niacin: 5200mg; Folic acid: 500mg; Biotin: 83.3mg; Choline: 83,333mg; \(^1\)Content (kg\(^{-1}\)): Iron: 10,000mg; Copper: 1333mg; Manganese: 2500mg; Iodine: 83.3mg; Zinc: 13,333mg; Selenium: 50.0mg; Mineral oil: 1.67mg; Si dioxide: 200g; \(^3\)Calcium propionate; \(^4\)Butylated hydroxytoluene.

Table 1 - Percentage and chemical composition of experimental diets.
the tank, and finally divided by 3 which corresponds to the volume of water in each aquarium.

Performance data were submitted to analysis of variance. Then the data set was subjected to regression analysis, considering the performance parameters as the dependent variable and the CP level of the diet as the independent variable. The following models were used: quadratic polynomial model, where \( Y_i = \beta_0 + \beta_1 PB + \beta_2 PB^2 + e_i \), where \( Y_i \) is the dependent variable; \( \beta_0 \) = intercept; \( \beta_1 \) = linear-component parameter; and \( \beta_2 \) = quadratic-component parameter. Linear response plateau (LRP): \( Y_i = \beta_0 + \beta_1 PB \), to estimate the straight line and \( Y_i = \beta_2 \) to estimate the plateau; where: \( \beta_1 \) = estimate of the performance parameters. Point of intersection of the straight lines was obtained by: \( PB = (\beta_1) / (\beta_2) \).

The estimate of CP optimal level needed to improve performance based on the LRP was obtained by the intercept of the straight line with the plateau. Regarding the quadratic polynomial, its estimate was obtained by equalizing to zero the first derivative of the equation. The optimal level, as estimated by the exponential model, was obtained considering 95% of the asymptotic response as calculated by the expression (ln 0.05)/\( \beta_0 \) (SAKOMURA & ROSTAGNO, 2007). The first intersection of the quadratic equation with the LRP plateau was obtained by the equation \( L = \beta_0 + \beta_1 PB + \beta_2 PB^2 \), and the CP level corresponding to the intersection was calculated by the expression \((-\beta_1 + (\beta_1^2 - 4 \beta_2 (L - \beta_0))^{1/2}) / (2\beta_2)\).

Hematologic data were analyzed by comparing the mean values while using the Tukey test. Data were analyzed using the procedures for analysis of variance and regression contained in the System for Statistical and Genetic Analysis (SAEG; Universidade Federal de Viçosa, UFV, 2000; v. 9.0).

Probability values less than 0.05 were considered significant in all statistical procedures.

RESULTS AND DISCUSSION

Mean values were calculated for temperature (24.5±2.6°C), dissolved oxygen in the water (5.15±1.2mg L\(^{-1}\)), and pH in the lagoon (6.7±0.3). These values remained within the range recommended by POPMA & GREEN (1990) for proper performance of tilapia.

The CP requirements for different performance parameters varied depending on the mathematical models used in the study. Regarding weight gain, an increase (P<0.05) was observed in the value estimated for CP (353.1g kg\(^{-1}\)) when the quadratic regression model was used (Table 2). However, other values were estimated for a greater weight gain as a function of CP when the LRP model (303.2g kg\(^{-1}\)) and LRP quadratic model (324.3g kg\(^{-1}\)) were used (Table 3). In this study, the quadratic model applied to the LRP allowed a better fit to the data, resulting in an estimate intermediate between those obtained with the quadratic and LRP models.

The LRP model alone tends to underestimate the optimal level because it does not take into account the biological law of decreasing return, and the plateau is then obtained once the maximum response is achieved. Conversely, quadratic model tends to overestimate the optimum level. Therefore, a combined (LRP and quadratic) model, which provides intermediate values between the models alone, is recommended to determine requirements in dose-response experiments (SAKOMURA & ROSTAGNO, 2008).

The CP value (324.3g kg\(^{-1}\)) estimated for weight gain indicated that an improvement

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Table 2 - Performance, ammonia excretion, and values for hematocrit, hemoglobin, and plasma protein.

<table>
<thead>
<tr>
<th>Variables</th>
<th>270</th>
<th>300</th>
<th>330</th>
<th>360</th>
<th>P values</th>
<th>VC¹</th>
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</thead>
<tbody>
<tr>
<td>Initial weight, g</td>
<td>13.25</td>
<td>13.59</td>
<td>13.42</td>
<td>13.16</td>
<td>0.45</td>
<td>5.66</td>
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<tr>
<td>Weight gain², g</td>
<td>32.48</td>
<td>41.33</td>
<td>40.27</td>
<td>44.27</td>
<td>0.03</td>
<td>4.23</td>
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<tr>
<td>Feed intake, g</td>
<td>39.63</td>
<td>40.50</td>
<td>40.67</td>
<td>41.17</td>
<td>0.35</td>
<td>5.51</td>
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<td>Feed conversion²</td>
<td>1.22</td>
<td>0.98</td>
<td>1.01</td>
<td>0.93</td>
<td>0.02</td>
<td>6.49</td>
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<tr>
<td>Ammonia excretion³, mg L⁻¹</td>
<td>0.0141</td>
<td>0.0157</td>
<td>0.0158</td>
<td>0.0195</td>
<td>0.00</td>
<td>27.91</td>
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<tr>
<td>Hematocrit, %</td>
<td>35.00</td>
<td>33.00</td>
<td>33.00</td>
<td>37.00</td>
<td>0.47</td>
<td>10.15</td>
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<tr>
<td>Hemoglobin, g dL⁻¹</td>
<td>8.34</td>
<td>7.63</td>
<td>8.10</td>
<td>8.57</td>
<td>0.54</td>
<td>11.64</td>
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<tr>
<td>Plasma protein, g dL⁻¹</td>
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<td>3.20</td>
<td>2.97</td>
<td>3.43</td>
<td>0.02</td>
<td>10.85</td>
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</table>

Mean values labeled with different letters in the same row are different (P<0.05) by the Tukey test.

¹Variation coefficient; ²Significant effect (quadratic and LRP); ³Linear effect.

Table 3 - Equations adjusted for weight gain and feed conversion as a function of crude protein levels.

<table>
<thead>
<tr>
<th>Models</th>
<th>Weight gain</th>
<th>Feed conversion</th>
<th>Ammonia excretion</th>
</tr>
</thead>
<tbody>
<tr>
<td>Quadratic</td>
<td>Y=143.378 + 105.849 PB – 0.149869 PB²</td>
<td>0.86</td>
<td>353.1</td>
</tr>
<tr>
<td>LRP</td>
<td>422.689 = -471.204 + 2.948 PB</td>
<td>1.00</td>
<td>303.2</td>
</tr>
<tr>
<td>Quadratic + LRP</td>
<td>422.689 = -143.378 + 105.849 PB – 0.149869 PB²</td>
<td>-</td>
<td>324.3</td>
</tr>
<tr>
<td>Linear</td>
<td>Y=0.81204625 + 0.000577317 PB</td>
<td>0.86</td>
<td>343.3</td>
</tr>
<tr>
<td></td>
<td></td>
<td>9.687974=34.23457 - 0.08151412 PB</td>
<td>1.00</td>
</tr>
</tbody>
</table>

1 Estimated level of crude protein.

occurred in protein efficiency, with a reduction in protein content from the maximum value (360g kg⁻¹) as used in the experiment (Figure 1). This effect can be attributed to the decrease in the amount of amino acids exceeding fish requirements, which did not participate in body protein formation and were then excreted (BOMFIM et al., 2008). The CP value estimated in the present study (297.3g kg⁻¹; for tilapia post-reversion up to 100g of body weight) were above those reported by FURUYA (2010a). However, BOTARO et al. (2007) evaluated the effect of different protein concentrations in the diet (291-245g kg⁻¹) based on the concept of ideal protein for tilapia, and they observed no significant difference between the effects on weight gain after reducing protein in the diet.

As expected, reduction in the CP values had no effect on feed consumption, since the diets were isocaloric and the fish were fed to satiation. This result confirmed the study of BONFIM et al. (2008), who observed that feed consumption in fish was regulated by energy intake.

In the present study, feed conversion was influenced (P<0.05) by reduction in the CP levels in the feed. With regard to the best feed conversion, the best CP value was estimated by the quadratic regression (343.3g kg⁻¹), and a different value (301.1g kg⁻¹) was estimated by the LRP model. However, an intermediate value (317.8g kg⁻¹) was obtained for the best feed conversion when the quadratic regression was combined with the LRP plateau. As feed conversion is directly related to effectiveness in deposition of nutrients ingested and feed intake was not different (P>0.05) between the treatments, it can be concluded that improvement in feed conversion (P<0.05) occurred due to weight gain provided by the levels of protein in the feed.

Ammonia excretion (Table 2) was significantly influenced (P<0.05) by the CP values in the diet. Regression analysis showed that ammonia excretion decreased linearly (Y=-0.00210625+0.000577317x; r²=0.81) with the decrease in the CP value. As difference between feed intake (P>0.05) was not observed in the treatments, reduced ammonia excretion occurred by a decrease in nitrogen consumption as the CP levels were reduced. FURUYA et al. (2005) also reported a lower nitrogen excretion with reduction in the amount of protein in the feed.

Reduction in the CP value in the diet associated with supplementation of synthetic amino acids allowed the reduction in nitrogen excretion by the fish without any negative effect on the weight gain until value of 324.3g kg⁻¹ CP. In intensive production systems, such as net pens, this is an important finding because the possibility of reducing ammonia excretion without impacting performance results is important for a sustainable tilapia culture.

Difference was not reported (P>0.05) between the respective hemoglobin and hematocrit values. However, the values obtained in the present study remained within the reference range (7.0-9.8g dL⁻¹ and 27-37%, respectively), as described by HRUBEC & SMITH (2010). A statistical difference (P<0.05) was observed between the lowest and highest values for plasma protein as a response to treatments containing the lowest and highest protein values. According to STOCKHAM & SCOTT (2011), most plasma proteins are composed of albumins and globulins, and they can be decreased in conditions...
such as severe hepatic dysfunction or poor nutrition. According to MARTINS (2011), protein restriction in the diet can cause hypoproteinemina in monogastric animals. The lowest level of plasma protein seen after intake of the diet containing 260g kg$^{-1}$ CP may be related to a deficiency in amino acids that were not supplemented, especially the non-essential ones, that are also important for the synthesis of plasma proteins.

CONCLUSION

The values of 324.3 and 317.8g kg$^{-1}$ for crude protein in the diets promote the best results in weight gain and feed conversion, respectively, for Nile tilapia (ca. 14g) bred in net pens. Reduction in crude protein to the level estimated in this study and supplementation with synthetic amino acids, based on the ideal protein concept, reduce the ammoniacal nitrogen excretion in water and did not alter the hematological parameters in fish.

BIOETICS AND BIOSECURITY COMMITTEE APPROVAL

This study was submitted to and approved by the Ethics Committee (Process N. 004-3/2012; UVV).

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REFERENCES


