Renal injury in female dogs with pyometra

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ABSTRACT: Pyometra is a common disease in intact female dogs and can cause glomerulopathy and tubular injury. This study aimed to evaluate kidney injury in female dogs with pyometra, as well as progression of the injury during treatment and the markers of this condition. This study analyzed 20 intact female dogs with both clinical and sonographic diagnosis of pyometra. Dogs were treated with intravenous fluids and antibiotics, and an ovariohysterectomy was performed. The following parameters were assessed at eight separate time points: blood pressure; serum creatinine, phosphorus, and urea levels; urinalysis and urinary biochemical parameters (urinary gamma-glutamyl transferase (uGGT) and urinary protein-to-creatinine ratio (UCPR)); glomerular filtration rate (GFR); and urine output. All dogs showed some degree of kidney injury at the time of pyometra diagnosis. This was transient in most animals, resolving with treatment of the pyometra. Measurement of uGGT and UPCR identified renal parenchymal injury, helping to determine the prognosis of the animals analyzed in the present study.

Key words: kidney disease, kidney function, urinary GGT, urinary protein-to-creatinine ratio, glomerular filtration rate.

INTRODUCTION

Female dogs with pyometra may also manifest decreased renal perfusion, immune-mediated glomerulonephritis, low ability for urinary concentration, interstitial tubular disease and/or decreased GFR (STONE et al., 1988; HEIENE et al., 2001; MASTROCINQUE, 2002; OLIVEIRA, 2007; FOSSUM, 2008; CHEW et al., 2011). These changes are caused by injuries to different parts of the kidney, resulting in varying degrees of renal impairment. The evolution of these changes is gradual and can result in kidney failure.
or, more precisely, by the UPCR. Proteinuria, or more specifically albuminuria, indicates glomerulonephropathy. Tubular cell lesions can be identified by measuring uGGT. Decreased GFR is another indicator of impending renal failure. GFR is directly related to the number of functional nephrons and is the gold standard for assessing kidney function. Serum phosphorus concentration is a sensitive marker of excretory function and is directly related to GFR. An elevation in serum phosphorus concentration precedes azotemia with several kidney injuries (GRECO et al., 1985; ROCHA & VEADO, 2005; CHEW et al., 2011; NABITY, 2011; HARLEY & LANGSTON, 2012; FREITAS et al., 2014).

Both pyometra and surgical and/or anesthetic complications associated with its treatment can cause kidney injury. A late diagnosis of pyometra, when kidney failure has already occurred, may result in irreversible damage to the kidneys (FOSSUM, 2008). This study aimed to evaluate kidney injury at the time of diagnosis of pyometra as well as progression of kidney injury during treatment for pyometra.

MATERIALS AND METHODS

The present study evaluated 20 intact female dogs of varying breeds. All animals were between 6 and 15 years of age and had been diagnosed with pyometra. Animals were treated at the Veterinary Hospital of the Universidade Federal de Minas Gerais (UFMG), Brazil, between May and December 2012. Treatment consisted of ovariohysterectomy and antibiotics, with all animals receiving IV fluid therapy (Lactated Ringer’s solution at 20mL kg⁻¹ h⁻¹ for fluid replacement over 3 to 4h and then 60mL kg⁻¹day⁻¹ for maintenance) beginning immediately upon diagnosis. Parenteral fluid therapy was maintained for up to 48h after surgery. The animals were medicated with amoxicillin-potassium clavulanate (20mg kg⁻¹ BID x 10 days). Ovariohysterectomy was performed following clinical stabilization.

The anesthetic protocol consisted of premedication with tramadol hydrochloride (4mg kg⁻¹ IV) followed by induction with 1% propofol (5mg kg⁻¹ IV). Isoflurane in O₂ was used for maintenance. Postoperative analgesia was provided by tramadol hydrochloride (3mg kg⁻¹ IV TID x 3 days).

Physical examinations and laboratory tests were performed at eight time points: T0 (diagnosis and hospitalization), T1 (period after fluid replacement), T2 (1-2h of fluid therapy for maintenance after T1), T3 (12h after surgery), T4 (24h after surgery), T5 (48h after surgery), T6 (10 days after surgery), and T7 (60 days after surgery).

Diagnostic tests performed included oscillometric blood pressure measurement (CONTEC 08A Veterinary Digital Blood Pressure & Heart Beat Monitor & SpO₂ Monitor, Contec, China); serum creatinine, phosphorous, and urea measurements; urinalysis and urine biochemistry (uGGT, UPCR) by enzymatic, kinetic and colorimetric methods on Cobas Mira Plus (Roche®) automated spectrophotometer according to reagent kit specifications (Synermed®); urine output (UO) and endogenous creatinine clearance (GFR). UO and GFR were calculated using a urethral catheter connected to a closed urinary drainage system and measured from T4 due to the minimum amount of time necessary for the calculation (24h). The following formulas were used: UO = urine volume (mL) weight⁻¹ (kg) 24h⁻¹ and GFR = [urine creatinine (mg dL⁻¹) x urine volume (mL)]/[serum creatinine (mg dL⁻¹) x weight (kg) x time (min)]. Systolic blood pressure (SBP)>150mmHg was consistent with hypertension.

Statistical analysis was performed using a completely randomized design and the differences were considered to be significant when P<0.05. Parametric data were tested for normality (Kolmogorov-Smirnov test) and homoscedasticity and subjected to analysis of variance. Either Duncan’s test or the Kruskal-Wallis test was used to compare different time points. Dispersion frequency was evaluated by the Fisher’s exact test and interactions were evaluated by Spearman’s correlation for data not normally distributed (creatinine, phosphorous, and GFR; SBP and UPCR; UPCR and leukocyturia, pyuria, bacteriuria, and uGGT; uGGT and renal epithelial (kidney cells and pelvis cells) cells in the urinary sediment).

RESULTS AND DISCUSSION

Varying degrees of azotemia (increased urea and/or creatinine) were observed among the female dogs at the different time points as shown in table 1. At T0, fewer of the dogs in this study were azotemic than has been previously reported. STONE et al. (1988) analyzed 27 female dogs with pyometra and found 26% of them to be azotemic at the time of diagnosis. ALBUQUERQUE (2010) analyzed 170 female dogs with pyometra and found 17.6% had increased creatinine measurements and 34.1% had increased urea measurements at the time of diagnosis. In analyzing 20 female dogs with pyometra, EVANGELISTA et al. (2010) reported that 30% of them had increased creatinine measurements and 75% had increased urea measurements whereas MIRA (2010), in analyzing the same number of affected dogs, reported that 30%

of them had increased creatinine measurements. Preexisting kidney damage and time elapsed between the development of pyometra and treatment start are relevant factors for this variation, but it is difficult to ascertain this information in the clinical setting.

The presence of azotemia in the animals in this study at T6 and T7 suggests either preexisting kidney injury or the development of chronic kidney disease secondary to a major insult (pyometra). These two conditions could not be distinguished because neither ultrasonography nor kidney biopsy was performed in the present study. EVANGELISTA et al. (2010) also reported increased creatinine measurements in female dogs 10 days after ovariohysterectomy were performed as treatment for pyometra, including those without azotemia at the time of diagnosis. This points out the importance of postoperative evaluation of kidney function in dogs treated for pyometra.

Regarding serum phosphorus levels, some of the animals in the present study had elevated measurements at all time points, including T6 and T7 (Table 1). At T0, the mean values were similar to those reported by YU (2012), who studied 49 female dogs with pyometra (phosphorus 6.3±0.6μg dL⁻¹). Phosphorus levels were greater than those reported by ZARAGOZA et al. (2004) at this same time point (phosphorus 5.05μg dL⁻¹), who studied 15 female dogs with the same condition. The evaluation of serum phosphorus levels at different time points revealed that eight of the animals in this study (40%) had hyperphosphatemia without azotemia, and subsequently showed increased creatinine measurements; however, this was not statistically significant (P=0.05). Hyperphosphatemia may result from reduced renal excretion secondary to decreased GFR. This often precedes azotemia (ROCHA & VEADO, 2005; CHEW et. el., 2011). However, a statistically significant correlation between phosphorus level and GFR was not observed (P=0.17). A high percentage of the animals were hyperphosphatemic at T6; although, there was no reduction in GFR. Uncommonly, other factors such as diet, acidosis, and hormonal changes can interfere with serum phosphorus levels. These factors were not evaluated in the present study.

Hypertension was noted at all time points (Table 1) and no correlation was observed between systolic blood pressure and proteinuria (P=0.54). Of the four animals that were persistently hypertensive at more than five time points, only two (50%) also had proteinuria at T6 and T7. FINCO (2004) and STEPIEN & ELLIOTT (2007) reported that persistent hypertension was always associated with progressive kidney damage; however, this was only observed in some of the animals in the present study.

Regarding urine specific gravity (Table 2), among the five animals noted to be azotemic at T0, 3 (60%) were also isosthenuric. This finding indicates the
existence of a renal or prerenal disorder resulting in low ability for urinary concentration. This may be due to endotoxemia which is liable to occur with pyometra. The kidneys of patients with prerenal azotemia are commonly functional and should respond to hypoperfusion by developing hypersthenuria (MASTROCINQUE, 2002; OLIVEIRA, 2007; FOSSUM, 2008). This was not observed in the animals in this study. Isosthenuria can occur in animals subjected to rehydration procedures and in animals whose kidneys have lost the ability to concentrate urine (STOCKHAM & SCOTT, 2011). Of the four isosthenuric animals at T0, two (50%) demonstrated polydipsia, explaining the decrease in urine concentration noted. The primary differential for the remaining two animals is chronic kidney injury because polydipsia was not observed and they had not yet received intravenous fluid therapy. A high frequency of animals with reduced GFR, isosthenuria, and polyuria was evident at T4 and T5 (Tables 2 and 3), suggesting an inability by the kidneys to concentrate urine caused in many of the dogs by pyometra.

Hyposthenuria indicates that the renal tubules maintain the ability to dilute urine. Hyposthenuria is not expected to be reported in dehydrated animals. However, it was observed in one dehydrated dog, which may have resulted from desensitization to antidiuretic hormone (ADH or vasopressin) in the renal tubules caused by bacterial endotoxins. This characterizes nephrogenic diabetes insipidus (MASTROCINQUE, 2002; FOSSUM, 2008). Hyposthenuria observed at T1 was probably due to fluid therapy.

Table 2 - Mean, standard deviation, and relative frequency of urine concentrating ability at various time points for 20 female dogs with pyometra.

<table>
<thead>
<tr>
<th>Time (h)</th>
<th>Mean and standard deviation</th>
<th>Frequency of Hyposthenuria (Density&lt;1.008) (%)</th>
<th>Frequency of Isosthenuria (Density 1.008-1.014) (%)</th>
<th>Frequency of animals (Density 1.015-1.045) (%)</th>
<th>Frequency of Hypersthenuria (Density &gt;1.045) (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T0</td>
<td>1.027 cd (0.017)</td>
<td>5</td>
<td>21</td>
<td>63</td>
<td>11</td>
</tr>
<tr>
<td>T1</td>
<td>1.014 a (0.009)</td>
<td>21</td>
<td>50</td>
<td>29</td>
<td>0</td>
</tr>
<tr>
<td>T2</td>
<td>1.016 ab (0.011)</td>
<td>10</td>
<td>60</td>
<td>25</td>
<td>5</td>
</tr>
<tr>
<td>T3</td>
<td>1.021 bc (0.011)</td>
<td>5</td>
<td>15</td>
<td>75</td>
<td>5</td>
</tr>
<tr>
<td>T4</td>
<td>1.017 abc (0.008)</td>
<td>5</td>
<td>40</td>
<td>55</td>
<td>0</td>
</tr>
<tr>
<td>T5</td>
<td>1.018 abc (0.008)</td>
<td>0</td>
<td>50</td>
<td>50</td>
<td>0</td>
</tr>
<tr>
<td>T6</td>
<td>1.033 de (0.012)</td>
<td>5</td>
<td>0</td>
<td>90</td>
<td>5</td>
</tr>
<tr>
<td>T7</td>
<td>1.043 e (0.020)</td>
<td>0</td>
<td>0</td>
<td>59</td>
<td>41</td>
</tr>
</tbody>
</table>

Values followed by different letters (column) were significantly different (P<0.05). Time points: T0 (diagnosis and hospitalization), T1 (period after fluid replacement), T2 (1-2h of fluid therapy for maintenance after T1), T3 (12h after surgery), T4 (24h after surgery), T5 (48h after surgery), T6 (10 days after surgery), and T7 (60 days after surgery).
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Table 3 - Mean, standard deviation, and relative frequency of increased UPCR, increased GGT, decreased GFR and decreased UO at different time points for 20 female dogs with pyometra.

<table>
<thead>
<tr>
<th>Time</th>
<th>M and SD UPCR&gt;0.2(%)</th>
<th>M and SD (U/L)</th>
<th>GGT&gt;92 (U/L) (%)</th>
<th>M and SD mL kg⁻¹ min⁻¹</th>
<th>GFR&lt;2mL kg⁻¹ min⁻¹ (%)</th>
<th>M and SD mL kg⁻¹ h⁻¹</th>
<th>Oliguria UO&lt;1mL kg⁻¹ h⁻¹ (%)</th>
<th>Polyuria UO&gt;2mL kg⁻¹ h⁻¹ (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T0</td>
<td>0.76 bc (0.81)</td>
<td>65</td>
<td>339.87 e (531.43)</td>
<td>70</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
</tr>
<tr>
<td>T1</td>
<td>1.21 c (0.89)</td>
<td>85</td>
<td>153.81 ab (244.94)</td>
<td>39</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
</tr>
<tr>
<td>T2</td>
<td>0.83 bc (0.51)</td>
<td>90</td>
<td>167.12 abc (318.20)</td>
<td>55</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
</tr>
<tr>
<td>T3</td>
<td>0.70 bc (0.41)</td>
<td>95</td>
<td>151.34 bc (138.67)</td>
<td>50</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
<td>NM</td>
</tr>
<tr>
<td>T4</td>
<td>0.54 b (0.44)</td>
<td>80</td>
<td>91.06 ab (99.53)</td>
<td>32</td>
<td>2.33 ab (1.67)</td>
<td>55</td>
<td>1.78 bc (1.07)</td>
<td>20</td>
</tr>
<tr>
<td>T5</td>
<td>0.61bc (0.47)</td>
<td>90</td>
<td>93.24 ab (77.87)</td>
<td>42</td>
<td>2.04 a (1.40)</td>
<td>63</td>
<td>1.87 c (1.08)</td>
<td>25</td>
</tr>
<tr>
<td>T6</td>
<td>0.23 a (0.28)</td>
<td>32</td>
<td>54.55 a (33.48)</td>
<td>20</td>
<td>2.55 b (1.33)</td>
<td>26</td>
<td>1.12 ab (0.58)</td>
<td>45</td>
</tr>
<tr>
<td>T7</td>
<td>0.14 a (0.19)</td>
<td>21</td>
<td>50.41 a (23.18)</td>
<td>8</td>
<td>2.66 ab (1.23)</td>
<td>42</td>
<td>0.81 a (0.33)</td>
<td>76</td>
</tr>
</tbody>
</table>

Values followed by different letters (column) were significantly different (P<0.05). Time points: T0 (diagnosis and hospitalization), T1 (period after fluid replacement), T2 (1-2h of fluid therapy for maintenance after T1), T3 (12h after surgery), T4 (24h after surgery), T5 (48h after surgery), T6 (10 days after surgery), and T7 (60 days after surgery). M = Mean; SD = Standard Deviation; NM = Not Measured.

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(Table 3). uGGT was also elevated at T0, and then decreased in the subsequent evaluations (Table 3). ZARAGOZA et al. (2004) studied 15 female dogs with pyometra and observed a higher mean UPCR (1.13) and a higher frequency of animals with proteinuria (93%) than was observed in the present study. At T0, the frequency of proteinuria was higher than the frequency of azotemia, which was also reported by MADDENS et al. (2011). This demonstrated the importance of conducting tests that can identify early kidney injury, as it is often a silent condition. A higher frequency of animals with proteinuria was observed at early time points, suggesting glomerular damage (immune-mediated glomerulonephritis), which is often transitory and can be reversed after removal of the diseased uterus (MASTROCIQUE, 2002; OLIVEIRA, 2007; FOSSUM, 2008). In the present study, there was no correlation between increased UPCR and the presence of urinary sediment components (leukocytes, erythrocytes, and bacteria). This suggested that increased UPCR indicates proteinuria of renal origin that is little influenced by inflammation and infection of other segments of the urinary tract (ELLIOTT & GRAUER, 2007; CHEW et al., 2011; ROSA et al., 2016).

Animals with persistent proteinuria may develop severe kidney damage according to HEINNE et al. (2007). Some animals continued to have proteinuria 60 days after surgery (Table 3), demonstrating that kidney function evaluation remains necessary in the late postoperative period.

Regarding uGGT, the frequency of enzymuria observed in this study at T0 (70%) was higher than that reported by DE SCHEPPER et al. (1989) (49% of 75 female dogs). In addition, at T0, the number of animals with enzymuria was higher than the number of animals with increased urea and creatinine, suggesting that the female dogs developed tubular injury before manifesting azotemia. This supports an early diagnosis of kidney injury (ROCHA & VEADO, 2005). Urinary GGT is primarily located in the brush border of proximal tubule cells and is released when the cellular membrane is damaged. Serum GGT is too large of a molecule to cross the selective wall of the glomerulus and, therefore, increased levels of uGGT suggested a renal tubular cell origin. In the present study, uGGT was significantly correlated with increased UPCR (P=0.01; r=0.22), suggesting tubular damage caused by albuminuria (GRAUER, 2005). In addition, the significant correlations between uGGT and the presence of renal epithelial cells (kidney cells) (P<0.01; r=0.26) and granular casts (P<0.01; r=0.25) in the urine confirmed tubular injury.

Granular casts were the most common type reported in the present study (Table 4). Granular casts are formed in the renal tubules and the presence

of increased amounts indicates tubular injury (STOCKHAM & SCOTT, 2011), which was confirmed in the present study by the correlation with uGGT. The decline in the number of renal epithelial cells (kidney cells and pelvis cells) observed in the urine at later time points suggested resolution of kidney injury in most of the animals. The presence of casts is influenced by urine concentration. Casts are degraded more rapidly in dilute urine which underestimates the actual number. In general, there was a higher frequency of animals with cylindruria when the urine was more concentrated; however, a significant correlation existed only for cellular casts (P=−0.09).

CONCLUSION

All of the female dogs with pyometra in this study presented with glomerular injury, tubular injury, or both. The injury to the kidneys was transient in most animals; however, it persisted up to 60 days post-surgery in some cases, emphasizing the importance of continued monitoring of kidney function. Increased uGGT, UPCR, and serum phosphorus levels were earlier markers of kidney injury than increased serum creatinine levels, and should be used to evaluate female dogs with pyometra for possible acute kidney failure.

RESEARCH ETHICS COMMITTEE AND BIOSAFETY

This study was approved by the Ethics Committee on Animal Research (CEUA/UFMG), protocol 198/2011.

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