Comparative analysis of *Monilinia fructicola* and *M. laxa* isolates from Brazil: monocyclic components of peach brown rot

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**ABSTRACT:** Brown rot is the most important disease of peaches in Brazil. The objective of this study was to compare the brown rot monocyclic components from *Monilinia fructicola* and *M. laxa* isolates from Brazil on peaches, due to the detection of *M. laxa* in the São Paulo production area. Conidia germination and pathogen sporulation were assessed in vitro under a temperature range of 5-35°C and wetness duration of 6-48h. Incubation and latent periods, disease incidence, disease severity and pathogen reproduction on peach fruit were evaluated under 10, 15, 20, 25 and 30°C and wetness duration of 6, 12 and 24h. Six of seven parameters of a generalised beta function fitted to conidia germination of *M. fructicola* and *M. laxa* were similar. Only the shape parameter was higher for *M. fructicola* indicating that the range of temperatures and wetness periods favourable for germination is wider for *M. laxa* than for *M. fructicola*. The optimum temperature for brown rot development caused by *M. fructicola* was 24.5°C and for *M. laxa* was 19.8°C. At 10°C, *M. laxa* lesions produced more conidia than *M. fructicola*, and the opposite occurred at 30°C. The estimated maximum temperature for lesion development was also higher for *M. fructicola* than for *M. laxa*. *M. fructicola* is favored by warmer weather than *M. laxa* and the presence and impact of this specie in Brazil must be investigated especially in the South states.

**Key words:** epidemiology, Prunus; *Monilia*, disease monocycle.

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**INTRODUCTION**

Brown rot is the most important disease of *Prunus* species in Brazil (MAY DE MIO et al. 2011). The main *Monilinia* species related to the disease are *M. fructigena* Honey, *M. laxa* (Aderh. & Ruhland) Honey and *M. fructicola* (G. Winter) Honey. *M. fructigena* occurs mainly in pome and stone fruits in the European Union territory (EU) and has been eradicated in the United States (OGAWA et al., 1995). *Monilinia laxa* is common in the EU and can also be reported in South Africa, Chile, Iraq, United States (EFSA, 2011). *Monilinia fructicola* is reported to be a serious pathogen in North and South America, Japan, New Zealand and Australia (OGAWA et al., 1995; EFSA, 2011). In addition, it has been spreading in the EU since 2001 (EFSA, 2011). In Brazil, *M. fructicola* was reported in all stone fruit production areas (MAY-DE MIO et al., 2011; LICHTEMBERG et al., 2016) on peaches, plums and nectarines. Detection of *M. laxa* in São Paulo State was performed in 2007 (SOUZA et al., 2008). From 2008 to 2011, the frequency of *Monilinia* species was monitored annually in the peach producing areas in Brazil and there was no spread detected of *M. laxa* (LICHTEMBERG et al., 2016). However, *M. laxa* has been detecting in imported fruit from 2013 to 2015 (unpublished data) and the impact and relevance of these remains unknown and should be investigated.
A shift in the population of Monilinia spp. from *M. laxa* to *M. fructicola* was observed in the USA during the 1980s on diseased prunes and apricots (MICHAILEDES et al., 1987). In Spain a shift in the populations of *Monilinia* spp. was observed in 2008 when *M. fructicola* replaced *M. laxa* in some orchards (VILLARINO et al., 2012). In Brazil, the detection of *M. laxa* in orchards where *M. fructicola* is already present can lead to a change in disease behaviour. Conditions favourable to *M. fructicola* infection are not necessarily the same as those for *M. laxa*. Scattered information on the effect of temperature on monocyclic components of this disease has been published (PHILLIPS, 1984; BIGGS & NORTHOVER, 1988a; WATSON et al., 2002), but no comparative study of the effect of different temperature and wetness duration across different *Monilinia* peach pathogens from Brazil has been performed. The objective of this study was to assess the effect of different wetness periods and incubation temperatures on the development of brown rot monocycle caused by *M. fructicola* and *M. laxa* on peaches.

**MATERIALS AND METHODS**

Isolation of *Monilinia* spp. was performed by aseptic transfer to culture media (PDA) of conidia produced by symptomatic peaches collected from naturally infected fruit from commercial orchards in São Paulo State, Brazil. The isolates were collected from the main production area of São Paulo (Paranapanema, Jundiaí and Jarinu municipalities). Isolates ISMf1 and ISMf2, collected in orchards at Jundiaí municipality, SP, Brazil (23°8′23″S, 46°42′55″W), and the isolates ISMf3 and ISMf4, collected at Paranapanema municipality, SP, Brazil (23°23′16″S, 48°43′36″W), were identified as *M. fructicola* by morphological characteristics and by PCR using species-specific primers (CÔTÉ et al., 2004). The isolate ESALQ1 was obtained from rotted fruit from an orchard located at Jundiaí municipality, SP, Brazil (23°04′35″S, 46°47′14″W) and identified as *M. laxa* by SOUZA et al. (2008). The PCR reactions were performed with a common reverse primer (MO368-S) and the species-specific forward primers MO368-10R (*M. fructicola*) and laxa-R2 (*M. laxa*) (CÔTÉ et al., 2004). As the population of *M. laxa* remained restricted to the area where it has been detected, the isolate named as ESALQ1 represents this population. Isolates of *M. fructicola* represented the main area of SP production and were chosen by geographic distance.

All isolates were cultivated on potato-dextrose-agar (PDA) at 20°C under a 12h photoperiod.

**Conidia germination and sporulation of *Monilinia* isolates under different temperatures and wetness duration**

Conidia suspensions were obtained from 7-day-old colonies grown on PDA under a photoperiod of 12h and prepared with sterile distilled water. Three drops (0.05 µL each) of 10^5 conidia mL^-1 suspension from each culture were placed separately in polystyrene Petri dishes into a Gerbox® with moistened filter paper. The Gerbox® were kept in growth chambers (Eletrolab®, São Paulo, SP, Brazil) at 5, 10, 15, 20, 25, 30 and 35°C for 6, 12, 24, 36 and 48h of wetness duration. After each interval, 20 µL of lactoglycerol was placed on each drop of the spore suspension, and conidal germination was estimated. The number of germinated conidia was counted by observing 100 conidia in each drop under an optical microscope (400x). Three replications were used in each treatment, and the experiment was performed twice.

Sporulation of each isolate was assessed from 7-day-old colonies on PDA which were previously incubated at 5, 10, 15, 20, 25, 30 and 35°C. Mycelial discs of 0.5 cm diameter were removed from within 1 cm of the colony centre, randomly selected, and transferred to glass tubes with 5 mL of sterile water. Tubes were vortexed for 30 seconds, the mycelial discs were removed, and 30 µL of lactoglycerol was added to the conidia suspension. Sporulation of two isolates (ISMf1 and ESALQ1) was also assessed in 7-day-old lesions on fruit, following the same methodology described for the colonies in PDA. Conidia concentration was estimated using a hemocytometer. Experiments had 5 replications each and were performed twice.

**Incidence and severity of brown rot under different temperatures and wetness duration.**

The *in vivo* experiments were performed with isolates ISMf1 (*M. fructicola*) and ESALQ1 (*M. laxa*). Peaches of cultivar Dourado, the most important variety grown in São Paulo State, were surface-sterilized by immersion in a 0.5 % sodium hypochlorite solution for 5 minutes. Fruits were rinsed twice, air dried for 24h, and then placed in plastic containers. Each replicate consisted of a container holding 6 fruit. The fruit were wounded at a single point with a sterile needle (0.2mm). For each isolate, 30 µL of 10^5 conidia mL^-1 suspensions collected from 7-day-old colonies were placed on fruit wounds. Fruit containers were sealed with
Compared to the incubation and latent periods of the disease caused by isolate ESALQ1 of *M. laxa* by ANOVA and Tukey’s test after square root transformation. Numbers of conidia produced by both isolates were also compared by ANOVA and Tukey’s test.

A generalized beta function (BASSANEZI et al., 1998) described by *Y(T) = \{Y_{opt}\left[(T-T_{min}) / (T_{opt}-T_{min})\right]^{\left[3(T_{max}-T_{opt})/(T_{max}-T_{opt})\right]}\}^{b_{3}} \left\{\left[(T_{max}-T_{opt})/(T_{max}-T_{opt})\right]^{b_{3}} \right\} (where Y_{opt} is the lesion diameter at the optimal temperature; T_{min}, T_{opt}, and T_{max} are the lowest, optimal, and highest temperature for lesion development, respectively; and b_{3} is the shape parameter) was fitted to the final lesion diameters estimated at each temperature for each isolate by non-linear regression analysis using STATISTICA 6.0 (Statsoft, Tulsa). Equation parameters were compared by *t* test.

**RESULTS**

Conidia germination of *Monilinia* isolates under different temperatures and wetness duration

*M. fructicola* conidia germinated at a wide range of temperatures, ranging from 10 to 30ºC. High germination rates were observed for 12h and longer wetness periods. Low germination rates (less than 25%) were observed at 10 and 30ºC for the 6h wetness period. The beta-monomolecular model provided a good fit to the data (Table 1). The minimum germination temperatures estimated by the model for *M. fructicola*

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### Table 1 - Estimated parameters and standard errors (in parenthesis) for the surface curves of germinated conidia (proportion) of *Monilinia fructicola* (isolates ISMf1, ISMf2, ISMf3 and ISMf4) and *M. laxa* (isolate ESALQ1) as a function of temperature and wetness periods fitted to a beta-monomolecular model.

<table>
<thead>
<tr>
<th>Model parameters</th>
<th><em>M. fructicola</em></th>
<th><em>M. laxa</em></th>
</tr>
</thead>
<tbody>
<tr>
<td><em>b</em>&lt;sub&gt;1&lt;/sub&gt;</td>
<td>0.06 (0.013)</td>
<td>0.21 (0.173)</td>
</tr>
<tr>
<td><em>b</em>&lt;sub&gt;2&lt;/sub&gt;</td>
<td>4.71 (1.170)</td>
<td>A</td>
</tr>
<tr>
<td><em>b</em>&lt;sub&gt;3&lt;/sub&gt;</td>
<td>0.49 (0.041)</td>
<td>A</td>
</tr>
<tr>
<td><em>b</em>&lt;sub&gt;4&lt;/sub&gt;</td>
<td>35.01 (0.018)</td>
<td>A</td>
</tr>
<tr>
<td><em>b</em>&lt;sub&gt;5&lt;/sub&gt;</td>
<td>0.52 (0.037)</td>
<td>A</td>
</tr>
<tr>
<td><em>b</em>&lt;sub&gt;6&lt;/sub&gt;</td>
<td>0.83 (0.215)</td>
<td>A</td>
</tr>
<tr>
<td><em>b</em>&lt;sub&gt;7&lt;/sub&gt;</td>
<td>0.20 (0.041)</td>
<td>A</td>
</tr>
<tr>
<td><em>R</em>&lt;sup&gt;2&lt;/sup&gt;</td>
<td>0.76</td>
<td>0.82</td>
</tr>
</tbody>
</table>

\[ Z = \left(b_{1}^{2} + b_{2}^{2}\right) / \left(b_{3}^{2} + b_{4}^{2}\right)(1-b_{5}^{2} \exp(-b_{6}^{2} \cdot M)), \]

where *Z* represents the proportion of germinated conidia, *T* is the temperature (ºC), *M* is the wetness period (hours), *b*<sub>2</sub> and *b*<sub>4</sub> are the minimum and maximum temperatures, respectively, *b*<sub>3</sub> is a shape parameter, that influences the temperature range around the optimum in which the curve stays near to maximum germination, *b*<sub>6</sub> is a rate parameter, and *b*<sub>5</sub>, *b*<sub>6</sub>, and *b*<sub>7</sub> are parameters from the model with no biological meaning. *Parameters followed by the same letter in the row are not different by *t* test (*P*<0.05).

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**Data analysis**

The combined effect of temperature and wetness period on conidial germination was analyzed using the beta-monomolecular model: \[ Z = \left(b_{1}^{2} + b_{2}^{2}\right) / \left(b_{3}^{2} + b_{4}^{2}\right)(1-b_{5}^{2} \exp(-b_{6}^{2} \cdot M)), \] in which *Z* represents the proportion of germinated conidia; *T* is the temperature (ºC); *M* is the wetness period (h); *b*<sub>2</sub> and *b*<sub>4</sub> are the minimum and maximum temperatures, respectively; *b*<sub>3</sub> is a shape parameter, that influences the temperature range around the optimum in which the curve stays near to maximum germination; *b*<sub>6</sub> is a rate parameter; and *b*<sub>5</sub>, *b*<sub>6</sub>, and *b*<sub>7</sub> are parameters from the model with no biological meaning (HAU & KRANZ, 1990; BASSANEZI et al., 1998).

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At the same temperature until typical symptoms and sporulation were observed.

Disease incidence (number of fruit with symptoms) and disease severity (lesion size) were evaluated daily. Lesion diameter was estimated by the average of two perpendicular measurements of lesion size. Incubation period (time between inoculation and expression of symptoms in 50% of fruit) and latent period (time between inoculation and sporulation in 50% of fruit) were estimated. The entire experiment was performed twice.
and *M. laxa* were 4.7°C and 0°C, respectively. The maximum temperature was 35°C for both species (Figure 1, Table 1). Curve parameters were similar for both species, except for $b_5$, which was lower for *M. laxa*.

**Incidence and severity of brown rot for different temperatures and wetness duration**

The disease incidence was greater than 80% for both pathogens in all combinations of temperature and wetness duration (data not shown). Brown rot incubation periods ranged from 2.0 (at 25°C) to 5.7 (at 10°C) days for *M. fructicola* and from 2.5 (at 25°C) to 4.3 (at 10°C) days for *M. laxa*. Incubation periods of brown rot caused by *M. fructicola* were only higher than the incubation periods for *M. laxa* at 10°C (Table 2). At all other temperatures, there was no difference in incubation periods between pathogens. The brown rot latent periods ranged from 2.0 (at 25°C) to 5.8 (at 30°C) days for *M. fructicola* and from 2.5 (at 25°C) to 8.5 (at 10°C) days for *M. laxa*. Incubation periods of brown rot caused by *M. fructicola* were only higher than the incubation periods for *M. laxa* at 10°C (Table 2). At all other temperatures, there was no difference in incubation periods between pathogens.

Lesion diameters of brown rot caused by *M. fructicola* and *M. laxa* were significantly influenced by temperature and showed a good fit to the generalized beta function (Table 3). Lesion diameters at the optimum temperature ($Y_{opt}$) were higher for the *M. laxa* isolate than for the *M. fructicola* isolate when 6 and 12h of wetness duration were applied after inoculation. However, these differences in lesion diameters were less than 1cm, and no difference was detected between isolates under 24h wetness duration (Table 3). Similarly, the estimated maximum temperature ($T_{max}$) for lesion development was lower for the *M. laxa* isolate than for the *M. fructicola* isolate by 6 and 12h wetness periods after inoculation. With a 24h wetness period, the maximum temperature for lesion development was similar (approximately 33°C) for both isolates. Optimum temperature for lesion development under the three wetness periods was significantly lower for the *M. laxa* isolate than for the *M. fructicola* isolate. Other parameters were similar for both isolates.

**Sporulation of Monilinia isolates under different temperatures and wetness duration**

The number of conidia produced *in vitro* was highly variable for both *Monilinia* species, but a difference between species was detected only at 10°C. At this temperature, *M. fructicola* produced fewer conidia than *M. laxa* (data not shown). High conidia production was observed on peach lesions from both species between 15 to 25°C (Table 2).
Comparative analysis of Monilinia fructicola and M. laxa isolates from Brazil assessing monocyclic components of peach...

Table 2 - Average incubation and latent periods (days) for brown rot isolates and number of conidia produced (x 10⁷) mL⁻¹ by Monilinia fructicola (isolate ISMf1) and M. laxa (isolate ESALQ1) on peaches at different temperatures.

<table>
<thead>
<tr>
<th>Temperature (ºC)</th>
<th>M. fructicola (ISMf1)</th>
<th>M. fructicola (ESALQ1)</th>
<th>M. laxa (ISMf1)</th>
<th>M. laxa (ESALQ1)</th>
<th>Number of conidia</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>5.7 B⁺</td>
<td>4.3 A</td>
<td>8.5 B</td>
<td>2.8 B⁺</td>
<td>14.6 A</td>
</tr>
<tr>
<td>15</td>
<td>3.2 A</td>
<td>5.7 A</td>
<td>2.9 B</td>
<td>2.2 A</td>
<td>22.5 A</td>
</tr>
<tr>
<td>20</td>
<td>2.2 A</td>
<td>4.0 A</td>
<td>15.9 A</td>
<td>20.2 A</td>
<td>22.5 A</td>
</tr>
<tr>
<td>25</td>
<td>2.0 A</td>
<td>3.7 A</td>
<td>19.5 A</td>
<td>19.8 A</td>
<td>22.5 A</td>
</tr>
<tr>
<td>30</td>
<td>2.7 A</td>
<td>5.8 A</td>
<td>5.1 A</td>
<td>2.6 B</td>
<td>22.5 A</td>
</tr>
</tbody>
</table>

*Means in each row followed by the same letters do not differ according to ANOVA and Tukey test (P<0.05). Data are means of two sets and are original but for the mean test they were transformed by square root. *Sporulation was detected in less than 50% of the inoculated fruit; latent period was greater than 9 days.

However, variability in the number of conidia produced was very high in this temperature range. The M. laxa isolate produced more conidia than the M. fructicola isolate at 10°C and fewer at 30°C. At other temperatures, there was no difference in conidia production between species (Table 2).

**DISCUSSION**

Results of this study demonstrated that the optimum temperature estimated for brown rot development caused by M. laxa is lower than that estimated for M. fructicola. Additionally, at low temperatures, the number of conidia produced in brown rot lesions was higher for M. laxa than for M. fructicola. Conversely, at high temperatures M. fructicola lesions produced more conidia than M. laxa lesions, and the estimated maximum temperature for lesion development was higher for M. fructicola than for M. laxa. This occurred despite the fact that no differences in conidia germination were detected between species across different temperature and wetness settings.

For both Monilinia species, high conidia germination rates (>50%) were observed for 6h of wetness in the range of 15-25°C, and the maximum rates occurred for 12h. The optimal temperature and wetness duration ranges for germination were similar to those previously observed for M. laxa (Tamm & Flückiger, 1993) and M. fructicola (Weaver, 1950; Phillips, 1982; Casals et al., 2010). Parameters estimated by the beta generalized function were similar for both Monilinia species, except for the shape parameter (b⁺), showing that the

Table 3 - Parameters and standard errors (in parenthesis) from a generalized beta function¹ fitted to brown rot lesion diameter for Monilinia fructicola (ISMf) and M. laxa (ESALQ1), with three wetness duration (hours).

<table>
<thead>
<tr>
<th>Temp (ºC)</th>
<th>6</th>
<th>12</th>
<th>24</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>M. fructicola</td>
<td>M. fructicola</td>
<td>M. fructicola</td>
</tr>
<tr>
<td></td>
<td>ISMF1</td>
<td>ESALQ1</td>
<td>ISMF1</td>
</tr>
<tr>
<td>Y₀</td>
<td>7.46 A (0.251)</td>
<td>8.27 B (0.324)</td>
<td>7.22 A (0.289)</td>
</tr>
<tr>
<td>Tₘᵦ</td>
<td>8.36 A (1.286)</td>
<td>7.99 A (0.002)</td>
<td>7.93 A (1.419)</td>
</tr>
<tr>
<td>Tₓₒ</td>
<td>24.48 B (0.488)</td>
<td>19.84 A (1.406)</td>
<td>23.23 B (0.531)</td>
</tr>
<tr>
<td>b⁻</td>
<td>0.36 A (1.105)</td>
<td>0.50 A (0.177)</td>
<td>0.43 A (0.116)</td>
</tr>
<tr>
<td>Tₑₒ</td>
<td>33.00 B (0.001)</td>
<td>31.32 A (0.748)</td>
<td>33.00 B (0.001)</td>
</tr>
</tbody>
</table>

¹Y(T) = (1/[Yₑₒ(Tₑₒ)+Yₓₒ(Tₓₒ)]^2[b⁻(Tₓₒ-Tₑₒ)(Tₑₒ-Tₒ)]/[Tₑₒ-Tₓₒ]))]. Where Yₑₒ is the lesion diameter at the optimal temperature, Tₑₒ, Tₓₒ, and Tₒ are, respectively, the lowest, optimal, and highest temperature for lesion development, and b⁻ is the shape parameter. ²Parameters followed by the same letter in the row within each wetness duration are not different by t test (P<0.05).
range of temperatures and wetness periods favourable for germination is greater for *M. laxa* than for *M. fructicola*. In Brazil, a 6 h wetness period at 15-25°C occurs frequently during peach season in most areas of peach production (GARRIDO et al., 2011). Consequently, no restraint will be imposed on the conidia germination of *M. laxa* in Brazilian conditions, and forecast systems for brown rot based on these variables (range of temperatures and wetness periods favourable for germination) should not be useful.

For all monocyclic components of brown rot assessed in this study, high temperatures favoured *M. fructicola*, and low temperatures favoured *M. laxa*. The incubation and latent periods of brown rot under optimum conditions (20-25°C) were short for both pathogens, as already stated by BIGGS & NORTHOVER (1988a, b), but at 10°C, the incubation and latent periods were shorter for *M. laxa* than for *M. fructicola*. Similarly, the estimated temperatures for the maximum lesion diameter of *M. laxa* were 3-5°C below those of *M. fructicola*, regardless of wetness duration. Under the optimum wetness duration (24 h), the brown rot lesion diameters were similar for both species, as observed by PIZZUOLO et al. (2006). Low temperatures inhibited *M. fructicola* lesion development four-fold more than to *M. laxa*.

Although, there are many reports concerning different monocyclic components of *M. laxa* (PIZZUOLO et al., 2006) and *M. fructicola* (BIGGS & NORTHOVER, 1988a; 1988b; NORTHOVER & BIGGS, 1990; FOURIE & HOLZ, 2003; LUO & MICHAILEIDES, 2003; PIZZUOLO et al., 2006), none of them analyzed how environmental variables affect each monocyclic component. Methodologies used in different reports are highly variable. For instance, the disease has been described on different hosts, such as cherry (BIGGS & NORTHOVER, 1988b; NORTHOVER & BIGGS, 1990), nectarine (FOURIE & HOLZ, 2003), peach (BIGGS & NORTHOVER, 1988a; PIZZUOLO et al., 2006) and prune (LUO & MICHAILEIDES, 2003), and in different plant organs (BIGGS & NORTHOVER, 1988a). The inoculation method with (LUO & MICHAILEIDES, 2003) or without (PIZZUOLO et al., 2006) wounds and the environmental conditions are distinct in the different reports. The comparison performed in our research showed that *M. laxa* is as aggressive as *M. fructicola* on peach fruit under optimal conditions and can be favoured by temperatures lower than 15°C. The population shift of *Monilinia* spp. observed in the USA (MICHAILEIDES et al., 1987) was most likely due to the benomyl resistance of *M. fructicola* and somewhat the environmental preferences of *M. laxa* rather than its aggressive behaviour.

As observed in our study; although, *M. fructicola* shares some features with *M. laxa*, differences in ecological requirements and host plant preferences are reported from areas where these species co-exist (EFSA, 2011). In these areas, *M. fructicola* is mostly reported on fruit, whereas *M. laxa* is mostly prevalent on blossoms and twigs (EFSA, 2011). This differentiation could be explained by differences in weather conditions during flowering and fruit ripening, as the former occurs at spring at low temperatures, and the latter occurs in summer when temperatures are high. Peach production in Brazil is concentrated in the Southern region, where low temperatures are frequent (GARRIDO et al., 2011) and *M. laxa* could cause epidemic during blooming in this region. The impact of *M. laxa* in Brazil should be investigated by monitoring the species entrance in the country and prevalence of the species in São Paulo State, where *M. laxa* where first reported.

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