Caudal vena cava thrombosis in a dairy cow (Bos taurus) in Argentina

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ABSTRACT: Caudal vena cava thrombosis (CVCT) is an acute disease secondary to ruminal lactic acidosis and rumenitis with fatal outcome in cattle fed with a high-grain diet. The aim of this paper is to describe CVCT in an adult Holstein cow from a herd in Balcarce, Buenos Aires, Argentina. Clinical signs included dyspnea, weakness, recumbence, and hemoptysis, followed by death in 3.5 days. Important necropsy findings included multiple hepatic abscesses, a septic thrombus (4.0cm x 2.0cm) in the vena cava adjacent to the liver, and a 15cm hematoma in the right lung. Histological lesions observed in the pulmonary parenchyma included suppurative pneumonia with embolic bacterial colonies and severe interstitial fibroplasia. Dissociation and degeneration of elastic fibers were observed in the pulmonary arteries, and necrotizing vasculitis was present in the caudal vena cava. We concluded that CVCT should be included in the differential diagnosis of diseases with acute respiratory signs. The preventive measures to control the CVCT include the reduction of lactic acidosis.

Key words: cattle, Holstein, thrombosis, vena cava, hemoptysis.

1 Pathology

CVCT is a disease that affect dairy (GUDMUNDSON et al., 1978) and feedlot cattle (JENSEN et al., 1976). A CVCT is usually diagnosed in adult cattle; although, it can occur in young animals, it is rare in calves under one year of age (BRAUN, 2008). The main predisposing factors are adult cattle fed with high-concentrate diets and the consequent ruminal lactic acidosis and rumenitis (WOOLUMS, 2015). Rumenitis allows penetration of bacteria such as Fusobacterium necrophorum, Trueperella pyogenes (IKAWA et al., 1987), Streptococcus spp., Staphylococcus spp. and Escherichia coli, among others (CONSTABLE et al., 2017a). Other conditions, such as deep digital sepsis (SIMPSON et al., 2012), mastitis, or metritis (BRAUN, 2008; WOOLUMS, 2015) can also result in CVCT. Omphalophlebitis (GUDMUNDSON et al., 1978) and abomasal ulcers (DIVERS & PEEK, 2008a) are the most common causes of CVCT in young animals.

CVCT is associated with rupture of hepatic abscesses, inflammation extending into the hepatic vein or the caudal vena cava, formation of a septic thrombus, and is the cause of multiple septic emboli in the pulmonary capillaries (MILLER & GAL, 2017).

Clinical signs include dyspnea, tachypnea, hyperpnea, cough, pale mucous membranes, tachycardia, rales, and expiratory wheezing; the clinical course of the disease typically extends 2-18 days. Hemoptysis and epistaxis indicate a poor prognosis; death usually follows within 24-48 hours (BREEZE
et al., 1976; MILLS & PACE 1990; BRAUN, 2008; WOOLUMS, 2015). During the postmortem examination, hepatic abscesses are present and CVCT is observed. Pulmonary hematomas can be observed around a ruptured aneurysm, along with pulmonary edema, emphysema, hemorrhages, and abscesses (BREEZE et al., 1976; GUDMUNDSON et al., 1978; BRAUN, 2008; WOOLUMS, 2015; CONSTABLE et al., 2017a). This paper described a case of CVCT in a Holstein cow in Argentina.

The case occurred in May 2014 in a 350-ha dairy farm with 350 Holstein cows located near Balcarce, in the Buenos Aires province, Argentina. Milking cows grazed alfalfa (Medicago sativa), winter fodder crops (Lolium multiflorum and Avena sativa), and were fed with ground corn and wheat grains, sunflower meal, and vitamin and mineral supplements. A sixth lactation cow in good body condition developed signs of weakness over 3 days. Subsequently severe dyspnea, hemoptysis, and epistaxis occurred. Death occurred 8-12h after developing hemoptysis-epistaxis. A postmortem examination was performed, and tissue samples were collected in 10% buffered formalin solution for histopathological analysis (hematoxylin and eosin staining). Only selected tissue sections were Gram stained.

At necropsy, the cow was in good body condition with significant fat deposition. Multiple random abscesses, 2.0 to 5.0cm in size, were disseminated throughout the liver. A 4.0×2.0cm septic thrombus was reported in the hepatic portion of the caudal vena cava (Figure 1A). A portion of the right lung was enlarged and firm, white petechiae and interlobar edema. A hematoma, 15.0cm in diameter, was reported in the cranial region of the middle right lobe, which was associated with a ruptured fusiform aneurysm 3.0cm in diameter (Figure 1B). The pulmonary parenchyma surrounding the hematoma had multiple randomly spaced abscesses 2.0 to 6.0cm in size, widely distributed hemorrhages, and interstitial edema. Multiple blood clots were reported in the oral cavity, nasal passages, trachea, and rumen. Microscopically, the caudal vena cava had focally extensive, necrosuppurative, chronic-active phlebitis in the intima and adventitia (Figure 1C). Lung had suppurative pneumonia with the presence of embolic thrombi and intra-alveolar colonies of gram negative bacilli and coccobacilli (Figure 1D); severe interstitial fibroplasia with mixed inflammatory infiltrate was observed. Focal areas of edema, emphysema, and hemorrhages were observed in the alveoli. The same necrosuppurative exudate was present in the bronchi. Elastic fibers in the adventitia of the pulmonary arteries were degenerate and dissociated.

In this case, clinical and gross findings, their anatomical location, and microscopic lesions were compatible with those reported in CVCT in cattle (BREEZE et al., 1976; GUDMUNDSON et al., 1978; BRAUN, 2008; CULLEN & STALKER, 2015; CONSTABLE et al., 2017a). No other lesions compatible with ruminal lactic acidosis or other conditions were observed during postmortem examination and histopathological analysis. However, focal rumenitis could not be completely ruled out because the entire ruminal wall was not evaluated and the cause of the pathology could have been missed. Nevertheless, the presence of liver abscesses and exposure of highly fermentable foods such as wheat grain and ground corn allowed us to presume that the animal had ruminal acidosis. (PLAIZIER et al., 2007; ENEMARK, GARRY & McCONNEL, 2015).

Additional tests, including ultrasonography between the eleventh and twelfth right intercostal space, chest X-rays, hematology analysis, and blood chemistry (BRAUN et al., 2002; BRAUN, 2008; SIGRIST et al., 2008) can be useful to confirm the ante mortem diagnosis, though they are rarely used. Hepatic abscesses occurred in feedlot cattle consuming high-concentrate diets; although, resultant clinical disease with CVCT is rare. However, in dairy herds where cows also received diets rich in grains, CVCT is more frequently reported. This may be due to more prolonged dietary exposure over the longer lifespan of the animal. However, due to its short clinical course, nonspecific clinical signs, and high lethality, CVCT may be underdiagnosed.

Treatment is generally unsuccessful (GUDMUNDSON et al., 1978; DIVERS & PEEK, 2008a; WOOLUMS, 2015); although, SIGRIST et al. (2008) reported a successful treatment in an affected cow. A better approach is prevention by reduction of the incidence of ruminal lactic acidosis in high-production dairy cows, including periodical control feeding practices such as amount, type, composition and structure of grains and fibers, feeding frequency and degree of adaptation. Similarity, the evaluation of the equipment used for mixing and distribution of the food is a strategy that should be also include. In addition, a gradual acclimatization of the rumen to the diets could be done, mainly in the early lactation cows, transition cows and pre-calving cows and heifers. Buffers such as sodium bicarbonate and magnesium oxide and antibiotics such as monensin or lasalocid have given good results too (DIVERS & PEEK, 2008b, ENEMARK, 2009; GARRY & McCONNEL, 2015, CONSTABLE et al., 2017b). Other approaches for detection of ruminal acidosis included the clinical evaluation of the history or sins
of diarrhea, anorexia, depression, dehydration, along with evaluation of conditions in which they could occur, as well as food intake and milk production.

In Argentina and Uruguay, the incidence and economic losses due to CVCT are unknown and should be determined by prospective studies. Finally, CVCT should be included in the differential diagnosis of clinically affected animals that show signs of acute respiratory disease (BREEZE et al., 1976; MILLS & PACE 1990; BRAUN, 2008, WOOLUMS, 2015).

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