Effect of yacon syrup on blood lipid, glucose and metabolic endotoxemia in healthy subjects: a randomized, double-blind, placebo-controlled pilot trial

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Abstract
We investigate the impact of yacon syrup consumption on glycemic, lipid and metabolic endotoxemia in healthy subjects. Thus, 32 healthy were randomized into two groups that consumed 40 g of yacon syrup/day (= 8.74 g FDS/day) or a placebo, for 2-weeks. Anthropometric assessments, gastrointestinal effects, glycemic and lipid profile, and LPS were performed at the beginning and at the end of the study. The results obtained shown that the 2-weeks consumption of yacon syrup does not promote serum LPS alteration in healthy volunteers. Moreover, the glycemic and lipid profile were also similar between the groups before and after the intervention. Yacon syrup did not affect blood lipid, glucose or metabolic endotoxemia. However, it is possible that positive effects of yacon syrup will turn evident only in a long-term intervention. Further studies are needed to assess the long-term effect of yacon syrup consumption, and its use in obese and diabetic individuals.

Keywords: Smallanthus sonchifolius; fructooligosaccharides; prebiotics; lipopolysaccharides.

Practical Application: In this work we evaluate the effects of a prebiotic product (yacon syrup) on human healthy.

1 Introduction

Yacon [Smallanthus sonchifolius (Poepp. et Endl.) H. Robinson] is a perennial herbaceous plant of the family Asteraceae, native to the Andean regions of South America and cultivated in other countries, such as Brazil (Castro et al., 2017). They store their carbohydrates in the form of β-1,2-fructooligosaccharides (FOS), which are able to resist the hydrolysis of enzymes from the upper gastrointestinal tract (Delgado et al., 2013), and selectively stimulate the growth of bifidobacteria, a gram-positive microorganism that play a regulatory role in the colon by inhibiting the growth of putrefactive bacteria (Caetano et al., 2016). Thus, FOS are considered as a prebiotic, which was recently defined as “[…] a substrate that is selectively utilized by host microorganisms conferring a health benefit” (Gibson et al., 2017, p. 493). The dairy foods products are the most famous carrier of prebiotics, such as reduced-fat bioyoghurt containing either short- or long-chain inulin (Teimouri et al., 2018), reduced-fat Labneh cheese with inulin and β-glucan fibre-based fat replacer (Aydinol & Ozcanc, 2018), and milk-sour cherry juice mixture containing inulin and hydrocolloid (Dabour et al., 2019). So, the use of yacon as a source of prebiotic compounds may be interesting to provide an alternative way to functionalize foods and beverages, including dairy products.

Several preclinical and clinical trials support the idea of intestinal bacteria involvement in host metabolism and potential prevention of prebiotic interventions (Nakamura & Omaye, 2012). According to these trials, FOS intake favors the modulation of the microbiota, stimulating the growth of health-promoting microorganisms and reducing pathogenic bacteria populations. Furthermore, the short chain fatty acids (SCFA), such as butyrate, propionate, and acetate – the endproducts of FOS fermentation by the intestinal microbiota – act as substrates or signaling molecules in the regulation of the glucose homeostasis and lipid metabolism. In this sense, the glycemic levels and body weight, for instance, can be reduced (Caetano et al., 2016; Kim et al., 2014; Nakamura & Omaye, 2012; Wanders et al., 2011). In addition, specific intestinal bacteria (gram negative) seem to serve as lipopolysaccharide (LPS) sources and increased intestinal permeability and to play a role in systemic inflammation (Nakamura & Omaye, 2012; Kim et al., 2014). Interestingly, the number of bifidobacteria was inversely correlated with the development of glucose intolerance and LPS level (Delzenne et al., 2011, 2013). Cani et al. (2007) were the authors who first proposed that a gram-negative bacterial outer membrane component known as lipopolysaccharide (LPS) was responsible for early onset of inflammation, insulin resistance, obesity, and diabetes. The authors observed that supplementation of high-fat diet (HFD) in mice for 4-weeks increased the plasma LPS levels by 2 to 3 times than those of control animals and called it “metabolic endotoxemia”.

Studies investigating the effects of prebiotics on metabolic parameters and inflammation in human showed different relevant findings (Costa et al., 2012, 2015), therefore, additional research...
is required to establish a more clear relationship between them. Focusing on studies using healthy subjects, Van Dokkum et al. (1999) showed no modifications of plasma lipids and glucose absorption, after the consumption of 15 g of FOS/d for three weeks. In contrast, Brighenti et al. (1999) found a striking reduction in the triglycerides (TG) levels and a moderate decrease in plasma cholesterol after the consumption of 9 g inulin/d for 4 weeks; and Letexier et al. (2003) observed a decrease in plasma TG (~16%) in subjects ingesting 10 g inulin/d for six weeks. Some studies indicated that xylo-oligosaccharide (XOS) in combination with inulin modulates both the intestinal environment and immune status in healthy subjects (Lecerf et al., 2012). Nilsson et al. (2008) shows that including indigestible carbohydrates in the evening meal of healthy subjects improves glucose tolerance, lowers inflammatory markers, and increases satiety after a subsequent standardized breakfast.

It can be observed that there is evidence that relates the consumption of prebiotics with beneficial effects in humans. However, the understanding of the prebiotic relationship with glycemic, lipid and endotoxemia in human subjects is still scarce, and the results are contradictory and based, mostly, in the effect of isolated compounds (FOS, XOS, inulin, and others) and not as a complex matrix food, such as yacon syrup. The yacon syrup represents an alternative and convenient product to human diet consumption, presents high levels of bioactive compounds, especially FOS (Silva et al., 2018a), and is obtained from a process which comprises an acid and enzymatic treatment, followed by microfiltration and concentration (Silva et al., 2018b). After a short and medium-term intervention of this yacon syrup in healthy subjects (~ 8.74 g FOS/d), the results showed important sex-based differences in appetite responses to foods, with effects observed only in a medium-term intervention (Silva et al., 2017). However, its metabolic effects are still unexplored and are an important step in positioning it as a nutraceutical product.

Therefore, this pilot study aimed to evaluate the impact of yacon syrup consumption on glycemic, lipid and endotoxemia (lipopolysaccharides, LPS) in healthy subjects through a randomized 2-weeks placebo-controlled intervention study.

2 Materials and methods

2.1 Yacon syrup preparation

The yacon syrup was produced in a pilot plant by Embrapa Tropical Agroindustry (Fortaleza - Ceará – Brazil). Initially, after acid treatment of the yacon (Dionisio et al., 2013), the juice is extracted from the root, treated with Celluclast® 1.5 L and Pectinex® Ultra SP-L (500 ppm of each enzyme, at 35 °C, 175 rpm, for 2 hours), and filtered in microfiltration system (Silva et al., 2018b). Thus, the clarified juice was concentrated under vacuum (< 60 °C) until it reached 71°Brix, and pH 3.7. The syrup was portioned into 40 g packs (corresponding to 8.74 g of FOS, 71.66 kcal or 301.61 kJ) and stored at 5 °C. The placebo was corn syrup, diluted up to 71 °Brix with water, acidified with 0.1% citric acid (final pH ~3.7) and added of 0.018% caramel colorant. The placebo was portioned into 40 g and stored at 5 °C, as well as the yacon syrup. The fructooligosaccharides (FOS) was determined using a Fructan Assay Procedure AOAC Method 999.03, AACC Method 32.32 (Megazyme International Ireland Ltda, 2018).

2.2 Study design and subjects

Subjects were studied over a 2-weeks period in a double-blind placebo-controlled experiment. The volunteers were randomly assigned to two groups. Group 1 received yacon syrup containing an intake level of 8.74 g FOS/day and group 2 included volunteers who received the placebo syrup. The randomization plan was conducted at the website (Randomization, 2017). The study protocol was approved by the Research Ethics Committee of the State University of Ceará (UECE) (nº 56094516.4.0000.5534), according to the National Commission of Ethics in Research of the Ministry of Health (CONEP/MS). All volunteers signed a free and informed consent form prior to the intervention. Inclusion criteria were included in the study: both sexes; between 20 and 59 years of age; do not present chronic self-reported diseases; have a body mass index greater than 18.5 kg m⁻²; not being in current use and in the last 30 days of hypoglycemic, lipid-lowering and anti-inflammatory drugs; not to participate in another clinical trial; not present current consumption and in the last 30 days of fiber supplementation and FOS. Exclusion criteria were: pregnancy; current smoker or in the last 30 days; daily ethanol consumption greater than 30 g for men and 15 g for women; use of hypoglycemic, lipid-lowering, antibiotic and anti-inflammatory drugs during the intervention.

Thirty out of a total of 32 volunteers completed this study with good compliance. The study was completed by 30 volunteers, distributed in the yacon syrup (n = 15) and placebo groups (n = 15) (see Figure 1). The volunteers consumed daily 40 g of yacon syrup or 40 g of placebo in the morning, along with foods commonly consumed or alone. The volunteers were instructed not to change their eating habits and lifestyle during the intervention.

2.3 Demographic and anthropometric evaluation

At the beginning of the study, all the volunteers were interviewed by a structured form, containing data referring to sex, age, race (self-referenced) and health status. A digital scale (Tanita® HD-313) was used to evaluate body weight, with a capacity of 150 kg, a sensitivity of 100 grams. The individuals were weighed in a lightweight and barefoot clothing, positioned in an upright posture, with the feet fully positioned on the platform of the scale, with arms along the body and looking at the horizon. To measure the height, was used a wall stadiometer (Seca 208®) with scale in millimeters. Measurement of waist circumference was performed with the tape measure positioned on the midpoint between the last costal arch and the iliac crest of the standing subject, with the reading made at the time of expiration. These data were collected by previously trained field researchers at the beginning and end of the study. Weight and height were used to calculate body mass index (BMI, kg m⁻²), as well as waist circumference (WC, cm) and waist-hip ratio (WHR).
2.4 Blood sampling

At the beginning of the study and on day 15, the blood samples were collected from volunteers who fasted overnight (12 h), and placed in tubes containing EDTA. Samples were transferred to sterilized centrifuge tubes and centrifuged for 10 min at 3000 g for serum separation. All serum and plasma aliquots for the biochemical determinations were kept frozen at -80 °C until analysis.

2.5 Biochemical measurements

The concentrations of total cholesterol, serum HDL cholesterol and triacylglycerol were analyzed using enzymatic colorimetric methods. The fasting glycemia was analyzed by enzymatic UV method (hexoquinase). The cholesterol content associated with LDL was estimated using Friedewald's formula (Friedewald et al., 1972). Serum plasma insulin was measured by chemiluminescence. Apolipoproteins B (Apo-B100) was determined by turbidimetric immunoassay method (Autokits APOB100, Randox Chemicals USA Inc., Richmond, VA, USA). The analysis of lipopolysaccharides - LPS was performed using Enzyme-Linked Immunosorbent Assay (ELISA) and ultra-sensitive C-reactive protein (hsCRP) was analyzed using Immuno-turbidimetric method, commercial kits from MyBioSource. All the biochemical analyzes were performed in serum before (t₀) and after the intervention (t₁).

2.6 Evaluation of gastrointestinal effects

Potential adverse effects including flatulence, bloated feeling, abdominal rumbling and abdominal pain were monitored daily using a diary. Alterations related to constipation were evaluated through a constipation scale (minimum score: 0 - normal, maximum score: 30 – severe constipation), according to a questionnaire proposed by Agachan et al. (1996).

2.7 Data analysis

An intention-to-treat analysis was performed. The data were compared by Student t-test or chi-square. The data are given as means ± standard deviation or with their 95% confidence intervals, after analysis of covariance (ANCOVA). The statistical analyzes were performed with the SPSS package version 20.0. P values of < 0.05 were considered statistically significant.

3 Results and discussion

Yacon syrup is a product obtained by enzymatic treatment, followed by microfiltration and vacuum concentration until it reaches a level of 71 °Brix (Silva et al., 2018a). This product contains high levels of FOS and phenolic compounds derived from quinic and trans-cinnamic acids (such as chlorogenic acid), and presents some mineral elements, such as K, Ca and P, and essential amino acids, such as tryptophan, valine, and threonine (Silva et al., 2018b). Moreover, we demonstrated in previous work, that yacon syrup administered as a diet supplement in women individuals showed a positive effect on appetite after a 2-week period of intervention (Silva et al., 2017). However, these effects were not observed when the yacon syrup was acute administered.

The effects of yacon syrup in colonic transit time of healthy volunteers (Geyer et al., 2008) and in biochemical parameters and appetite of obese and slightly dyslipidemic pre-menopausal women (Genta et al., 2009) were reported in the literature. These findings raise the interesting possibility of investing the effects of yacon syrup on human health. However, to the author’s knowledge, the effects on blood lipid, glucose and metabolic endotoxemia in healthy subjects were still unexplored.

On this sense, in the present study, we evaluate the effect on blood lipid, glucose and metabolic endotoxemia of the yacon syrup consumption, in healthy volunteers. The yacon syrup has 22% of FDS, corresponding to a daily portion of 8.74 g FDS/d, for each volunteer, for 2-weeks. Thirty-two volunteers were initially included in the study, but two dropped out and were not replaced. Thirty subjects completed the study (retention rate of 94%). The baseline characteristics of these subjects are presented in Table 1 and have shown no differences between the intervention groups. Body mass index (BMI) in both groups (~ 25 kg/m²) was similar. No change was observed in the placebo group for WC, BMI, and WHR after the intervention period (see Table 2).
At the end of the study, adverse effects, such as flatulence, abdominal rumbling, bloated feeling, and cramps, were reported in the yacon syrup on the first days during the treatment. None of these were considered serious or harmful to health, and they subsided with adaptation over time. The literature reports that FOS will be tolerated and do not cause gastrointestinal problems when taken at a minor dose of 15 g daily (Djansivu et al., 2011; Scheid et al., 2014; Williams & Jackson, 2002). Genta et al. (2009) report significant gastrointestinal adverse effects such as diarrhea, severe abdominal, distention, flatulence, and nausea when the patients consumed yacon syrup containing 20 g FOS/d. In contrast, the group treated with yacon syrup at a level intake of 10 g FOS/d went through the whole experimental period with no difficulties.

Results from the biochemical markers for glucose and lipid metabolism at baseline and post-intervention for both groups are summarized in Table 3. At baseline, there were no

### Table 1. Pre-treatment characteristics of subjects scheduled to participate in this study.

<table>
<thead>
<tr>
<th>Variables</th>
<th>Yacon syrup (n = 15)</th>
<th>Placebo (n = 15)</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>41.7 ± 8.9</td>
<td>40.1 ± 8.0</td>
<td>0.585</td>
</tr>
<tr>
<td>Female (%)**</td>
<td>10.0 (66.7%)</td>
<td>11.0 (68.8%)</td>
<td>1.000</td>
</tr>
<tr>
<td>Body weight (kg)</td>
<td>69.1 ± 14.0</td>
<td>71.2 ± 16.4</td>
<td>0.712</td>
</tr>
<tr>
<td>WC (cm)</td>
<td>82.9 ± 8.2</td>
<td>82.9 ± 11.7</td>
<td>0.999</td>
</tr>
<tr>
<td>BMI (kg m⁻²)</td>
<td>25.3 ± 3.3</td>
<td>25.0 ± 4.8</td>
<td>0.834</td>
</tr>
<tr>
<td>WHR</td>
<td>0.8 ± 0.1</td>
<td>0.8 ± 0.1</td>
<td>0.741</td>
</tr>
<tr>
<td>Glucose (mg dL⁻¹)</td>
<td>85.0 ± 6.0</td>
<td>84.0 ± 7.0</td>
<td>0.694</td>
</tr>
<tr>
<td>Insulin (µU mL⁻¹)</td>
<td>9.2 ± 4.6</td>
<td>7.8 ± 3.6</td>
<td>0.337</td>
</tr>
<tr>
<td>Total cholesterol (mg dL⁻¹)</td>
<td>192.0 ± 27.0</td>
<td>191.0 ± 22.0</td>
<td>0.967</td>
</tr>
<tr>
<td>HDL-C (mg dL⁻¹)</td>
<td>54.0 ± 13.0</td>
<td>56.0 ± 13.0</td>
<td>0.646</td>
</tr>
<tr>
<td>LDL-C (mg dL⁻¹)</td>
<td>111.0 ± 21.0</td>
<td>112.0 ± 25.0</td>
<td>0.961</td>
</tr>
<tr>
<td>Apo B (mg dL⁻¹)</td>
<td>93.7 ± 14.9</td>
<td>91.0 ± 16.6</td>
<td>0.633</td>
</tr>
<tr>
<td>Triacylglycerols (mg dL⁻¹)</td>
<td>132.0 ± 81.0</td>
<td>117.0 ± 60.0</td>
<td>0.555</td>
</tr>
<tr>
<td>LPS (pg mL⁻¹)</td>
<td>12.9 ± 4.2</td>
<td>15.3 ± 4.8</td>
<td>0.235</td>
</tr>
<tr>
<td>hsCRP (mg dL⁻¹)*</td>
<td>0.13 (0.03-0.41)</td>
<td>0.15 (0.02-1.62)</td>
<td>0.922</td>
</tr>
</tbody>
</table>

*Values expressed in median (minimum and maximum values); **Probability, no significant differences in a Mann-Whitney Test (p > 0.05); *Expressed in n (%). n = number of volunteers; WC = waist circumference; BMI = body mass index; WHR = waist-hip ratio; LDL-C = low-density lipoprotein; HDL-C = high-density lipoprotein; Apo B = Apo lipoprotein B; LPS = lipopolysaccharides; hsCRP = highly sensitive C-reactive protein.

### Table 2. Anthropometric parameters before the test (baseline, t₀) and after 2-weeks (t₂) of treatment (yacon syrup or placebo).

<table>
<thead>
<tr>
<th>Variables</th>
<th>Yacon syrup (n = 15)</th>
<th>Placebo (n = 15)</th>
<th>Mean difference*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight (kg)</td>
<td>69.1 ± 14.0</td>
<td>69.2 ± 14.0</td>
<td>0.07</td>
</tr>
<tr>
<td>WC (cm)</td>
<td>82.9 ± 8.2</td>
<td>82.3 ± 8.0</td>
<td>-0.16</td>
</tr>
<tr>
<td>BMI (kg m⁻²)</td>
<td>25.3 ± 3.3</td>
<td>25.3 ± 3.3</td>
<td>0.03</td>
</tr>
<tr>
<td>WHR</td>
<td>0.8 ± 0.05</td>
<td>0.8 ± 0.05</td>
<td>-0.01</td>
</tr>
</tbody>
</table>

*Results expressed as mean ± standard deviation; *Regression analysis (β values and 95% confidence interval (CI)). n = number of volunteers; β = regression coefficient; CI 95% = 95% confidence interval; WC = waist circumference; BMI = body mass index; WHR = waist-hip ratio.

### Table 3. Clinical parameters related to glycemic, lipid, and endotoxemia before the test (baseline, t₀) and after 2-weeks (t₂) of treatment (yacon syrup or placebo).

<table>
<thead>
<tr>
<th>Variables*</th>
<th>Yacon syrup (n = 15)</th>
<th>Placebo (n = 15)</th>
<th>Mean difference*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose (mg dL⁻¹)</td>
<td>85.0 ± 6.0</td>
<td>87.0 ± 6.0</td>
<td>0.43</td>
</tr>
<tr>
<td>Insulin (µU mL⁻¹)</td>
<td>9.2 ± 4.6</td>
<td>12.3 ± 7.5</td>
<td>1.90</td>
</tr>
<tr>
<td>Total cholesterol (mg dL⁻¹)</td>
<td>192.0 ± 27.0</td>
<td>191.0 ± 22.0</td>
<td>6.19</td>
</tr>
<tr>
<td>HDL-C (mg dL⁻¹)</td>
<td>54.0 ± 13.0</td>
<td>51.0 ± 13.0</td>
<td>2.44</td>
</tr>
<tr>
<td>LDL-C (mg dL⁻¹)</td>
<td>111.0 ± 21.0</td>
<td>112.0 ± 25.0</td>
<td>6.16</td>
</tr>
<tr>
<td>Apo B (mg dL⁻¹)</td>
<td>93.7 ± 14.9</td>
<td>93.2 ± 14.5</td>
<td>1.68</td>
</tr>
<tr>
<td>Triglycerides (mg dL⁻¹)</td>
<td>132.0 ± 81.0</td>
<td>117.0 ± 60.0</td>
<td>17.84</td>
</tr>
<tr>
<td>LPS (pg mL⁻¹)</td>
<td>12.9 ± 4.2</td>
<td>15.3 ± 5.2</td>
<td>0.03</td>
</tr>
<tr>
<td>hsCRP (mg dL⁻¹)</td>
<td>0.2 ± 0.1</td>
<td>0.4 ± 0.6</td>
<td>0.15</td>
</tr>
</tbody>
</table>

*Results expressed as mean ± standard deviation; *Regression analysis (β values and 95% confidence interval (CI)). n = number of volunteers; β = regression coefficient; CI 95% = 95% confidence interval; LDL-C = low-density lipoprotein; HDL-C = high-density lipoprotein; Apo B = Apo lipoprotein B; LPS = lipopolysaccharides; hsCRP = highly sensitive C-reactive protein.
significant differences in any biochemical variable between the placebo and the experimental group. Serum levels of glucose, insulin, total, Apo B, HDL, LDL, cholesterol, and triglycerides (TG) were within normal ranges for both groups. Therefore, the yacon syrup supplementation (~ 8.74 g FOS/d) ingested by subjects in 2-weeks may have been insufficient to influence the glucose and lipid metabolism, indicating that it need to be tested in higher doses and/or a more extended duration study. Moreover, several papers reports that the effect of prebiotics in glycemic and lipid metabolism are conflicting and inconsistent, especially in healthy subjects (Liu et al., 2017; Kellow et al., 2014; Costa et al., 2012; Beylot, 2005).

Kellow et al. (2014), in a systematic review of randomized controlled trials evaluating the metabolic benefits of dietary prebiotics in human subjects, conclude that the effects of prebiotics on insulin sensitivity, lipids, inflammatory markers and immune function were contradictory. The authors show that the association can be clearly associated with subjective improvements in satiety and reductions in postprandial glucose and insulin concentrations. Liber & Szajewska (2013), evaluated the effects of inulin-type fructans (ITF) on energy intake and body weight in children and adults, and concluded that the long-term administration of ITF may contribute to weight reduction. Sperry et al. (2018) evaluated the clinical effects of regular consumption of probiotic Minas Frescal cheese on hypertensive overweighted women, and showed an improvement in total cholesterol, low-density lipoprotein-cholesterol, high-density lipoprotein-cholesterol, triacylglycerides, diastolic and systolic pressure, hemoglobin, and hematocrit count of the hypertensive overweighted women. Wanders et al. (2011) show that different types of fiber affect subjective appetite, acute energy intake, long-term energy intake and body weight differently. The authors indicate that additional research (short-term and long-term intervention) is still necessary. More recently, Liu et al. (2017) reviewed the effect of ITF on blood lipid profile and glucose level. The authors have shown that the use of ITF may have benefits for LDL-cholesterol reduction across all study populations, whereas HDL-cholesterol improvement and glucose control were demonstrated only in the type 2 diabetes mellitus group. Thus, the authors conclude that an additional, well-powered, long-term, randomized clinical trials are required for a definitive conclusion. Williams & Jackson (2002) reviewed the effects of inulin and oligofructose on lipid metabolism, in human studies. The authors observed that the positive outcomes in lipid metabolism were observed more frequently in those studies conducted in subjects with moderate hyperlipidemia. In this sense, the use of yacon syrup supplementation in subjects presenting a low-grade systemic inflammation such as in obesity, metabolic syndrome and/or patients of type 2 diabetes will be considered in further studies.

Yamashita et al. (1984) studied the systemic effects of adding 8g/d FOS for 2-weeks to the diet of patients with type 2 diabetes mellitus whose serum glucose concentration was uncontrolled. Fasting blood test showed 8% reduction in serum glucose levels. In contrast, Alles et al. (1999) evaluated the effect of 15g/d FOS for 3-weeks in patients with type 2 diabetes mellitus and showed no significant changes in the glucose levels and lipid concentrations. Additionally, Luis et al. (2011) and Dewulf et al. (2013) showed differences in glucose and lipid metabolism after a 4-week to 3-month consumption of a prebiotic supplemented diet in obese patients.

In healthy subjects, Brighenti et al. (1999) found a striking reduction in the plasma TG levels and a moderate decrease in plasma cholesterol after the consumption of 9 g inulin/d for 4 weeks; and Letexier et al. (2003) observed a 16% decrease in plasma TG in subjects ingesting 10 g inulin/d for 6 weeks. Russo et al. (2008, 2010) showed positive effects on lipid and glycemic profile when inulin-enriched pasta was consumed by healthy male subjects for 5-weeks. In contrast, Van Dokkum et al. (1999) showed no modifications of plasma lipids and glucose absorption, after the consumption of 15 g of FOS/d after three weeks. Luo et al. (1996) showed that the chronic consumption of short-chain FOS decreased basal hepatic glucose production but had no effect on insulin-stimulated glucose metabolism.

Lipopolysaccharide (LPS) or endotoxin is a bacterial cell wall component found predominantly in gram-negative bacteria that stimulates an inflammatory response in a process that increases the levels of tumor necrosis factor α (TNF-α), a potent inflammatory cytokine linked to obesity and type 2 diabetes (Parnell & Reimer, 2012). This has been named as metabolic endotoxemia (Cani et al., 2007). Evidence from animal studies has shown that LPS levels negatively correlate with bifidobacteria numbers. Moreover, with a prebiotic treatment, the gut barrier was improved, and the LPS level was reduced (Cani et al., 2009). However, in our study, the effect on LPS after the 2-week consumption of yacon syrup (see Table 3) was not significant.

Significant reductions in circulating lipopolysaccharide (LPS) concentrations after 4-weeks of dietary prebiotic supplementation in healthy subjects were reported by Lecerf et al. (2012). Creely et al. (2007) shows that the endotoxin levels are higher (76%) in individuals with type 2 diabetes than in healthy individuals.

In studies with women with type 2 diabetes, Dehghan et al. (2014) showed a significant decrease in the LPS levels after a 8-week oligofructose-enriched inulin consumption. In a similar study, when women with type 2 diabetes received a daily supplement of 10 g resistant dextrin for 8 weeks, the LPS level was significantly reduced (Aliasgharzadeh et al., 2015). However, in these papers, the intervention period was longer than that used in our study and focused mainly in type 2 diabetes subjects. Utzschneider et al. (2016) reviewed the mechanisms linking the gut microbiome and glucose metabolism, and examined the role of gut permeability in circulating LPS concentrations. The authors conclude that, although the data in animal models are convincing, more data are needed in humans.

To our knowledge, this is the first study to evaluate the effect of yacon syrup consumption on glycemic, lipid and endotoxemia (lipopolysaccharides, LPS) in healthy subjects. Further studies are needed to understand the beneficial effect of yacon syrup in a long-term intervention, and the effects to specific target populations, such as obesity or diabetes, for example.

4 Conclusions

In conclusion, 2-weeks of yacon syrup (~ 8.74 g FOS/d) consumption did not affect blood lipid, glucose or metabolic endotoxemia. However, it is possible that positive effects of yacon
syrup will turn evident only after a long-term intervention. Further studies are needed to assess the long-term effect of yacon syrup consumption, and its use in obese and diabetic individuals.

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