ABSTRACT

Objective: To evaluate ertapenem disk performance to predict Klebsiella pneumoniae carbapenemase production by Gram-negative bacilli.

Methods: All Gram-negative bacilli isolated between January 2010 and June 2011 were tested by disk diffusion (Oxoid™) for sensitivity to ertapenem, meropenem and imipenem. Resistant or intermediate sensitivity strains (diameter ≤22 mm for ertapenem) were also tested for the bla<sup>KPC</sup> gene by polymerase chain reaction. Disk predictive positive value for Klebsiella pneumoniae carbapenemase and specificity were calculated.

Results: Out of the 21839 cultures performed, 3010 (13.78%) were positive, and Gram-negative bacilli were isolated in 708 (23.52%) of them. Zone of inhibition diameter for ertapenem disk was ≤22 mm for 111 isolates, representing 15.7% of all Gram-negative isolates. The PCR assay for bla<sup>KPC</sup> detected 40 Klebsiella pneumoniae carbapenemase-producing strains. No strains intermediate or resistant to meropenem and imipenem were sensitive to ertapenem. The ertapenem disk presented a positive predictive value of 36% to predict bla<sup>KPC</sup> and 89% specificity.

Conclusion: The resistance of Gram-negative bacilli detected by disk diffusion against ertapenem does not predict Klebsiella pneumoniae carbapenemase. Other mechanisms, such as production of other beta-lactamases and porin loss, may be implicated. The need to confirm the presence of the bla<sup>KPC</sup> is suggested. Therefore, ertapenem was a weak predictor for discriminating strains that produce Klebsiella pneumoniae carbapenemase.

Keywords: Klebsiella; Carbapenems; Drug resistance, multiple, bacterial
INTRODUCTION

Ertapenem (ERT) is a beta-methyl-carbapenem that is active against Gram-negative bacteria producing extended-spectrum beta-lactamases (ESBL) or AmpC, and it is widely used in clinical practice since 2001. In the United States, the emergence of resistance against carbapenems is usually related to the production of carbapenemases, such as metallo-beta-lactamases and Klebsiella pneumoniae carbapenemase (KPC). However, other mechanisms may be involved in the resistance to this antibiotic class, including porin loss associated with ESBL or AmpC production.

KPC is a class-A carbapenemase that inactivates all beta-lactam antibiotics. It was first described in Klebsiella pneumonia, but it has also been occasionally detected in other enterobacteria. This enzyme is coded by sequences related to transposons and are present in conjugative plasmids with high dissemination power.

The identification of KPC-producing Gram-negative bacilli (GNB) is mandatory, as they may cause severe infections, and carbapenems (imipenem – IMP and meropenem – MER) are the therapy of choice in many nosocomial infections. In addition, the identification of carrier individuals allows controlling the dissemination of those agents. No precise phenotypic tool for their identification has yet been described, and the available tools are not able to differentiate resistance mechanisms.

The Brazilian Health Surveillance Agency (ANVISA) currently recommends disk-diffusion with carbapenems for screening and determination of minimum inhibitory concentrations (MIC), followed by confirmation using molecular biology to identify the gene \( \text{bla}_{KPC} \). The ERT disc is considered a very sensitive marker, but data published in Brazilian and international literature on its specificity are still subjective and conflicting.

OBJECTIVES

To evaluate the positive predictive value and the specificity of the ertapenem disk to predict the presence of KPC-producing Gram-negative bacilli.

METHODS

Gram-negative bacilli (GNBs) isolated in biological material cultures collected and processed at Hospital Municipal Dr. Moisys Deautch, in M’Boi Mirim, SP, Brazil, between January 2010 and June 2011, were tested by disk-diffusion (Oxoid™) for ERT, MER, and IMP in Mueller-Hinton agar (BioMérieux®). The interpretation criteria applied were those suggested in the Technical Note published by ANVISA in 2010.

RESULTS

Out of a total of 21839 cultures from different biological materials performed during that period, 3010 (13.78%) were tested positive, with GNB isolated in 708 (23.52%) of these positive cultures.

ERT disk zone of inhibition was ≤22mm in 111 (15.67%) of the isolates, out of which 96 (86.48%) were considered resistant and 15 (13.51%) intermediate. Strains were submitted to molecular testing. The \( \text{bla}_{KPC} \) gene tests characterized 40 KPC-producing strains. Klebsiella pneumoniae was biochemically identified in 38 cases and Enterobacter cloacae in 2 cases. The bacteria were isolated in the urine in 16 cases, rectal/anal swab in 17 cases, tracheal secretion in 4 cases, blood in 2 cases and pancreatic abscess in one case. Eleven of the infected patients died. In all gene-positive cases, the zone of inhibition diameter of the ERT disk was considered resistant. No MER or IMP intermediate or resistant samples were sensitive to ERT.

The results showed that there were 40 \( \text{bla}_{KPC} \)-positive strains out of the 111 strains found to be resistant or intermediate to ERT among 708 isolated GNB. Calculated ERT disk PPV and specificity for KPC production were 36% and 89%, respectively.

DISCUSSION

The study of Anderson et al. (7), considering the potential clinical impact, stresses the importance of evaluating the sensitivity to ERT because this is the most sensitive indicator of KPC, independently of the method applied.
However, Woodford et al.\(^8\) mentioned that the \textit{in-vitro} resistance to that carbapenem is not specific for KPC production, particularly when considering centers where carbapenemase-producing bacteria are rare.

The present study shows objective data on PPV and specificity of the ERT disk to predict KPC in our center. It was observed that ERT-disk resistance does not determine that the isolated GNB produces KPC. Other resistance mechanisms, such as the production of other beta-lactamases and porin loss, may be implicated. Therefore, ERT was a weak predictor for the identification of KPC-producing strains in our cohort.

\textbf{CONCLUSION}

Despite the efforts to find an ideal phenotypic method to trace KPC production, there are still no results supporting the individual use of these tools. There are many groups working to find better methodologies, but no definite conclusions have been published yet. Consistent with ANVISA guidelines, the data obtained in the present study also indicate the need of confirming the presence of the \textit{bla}\textsubscript{KPC} gene to determine KPC production using molecular biology.

\textbf{REFERENCES}


