Protective effect of carboxymethyl-glucan (CM-G) against DNA damage in patients with advanced prostate cancer

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Abstract

Carboxymethyl-glucan (CM-G) is a soluble derivative from Saccharomyces cerevisiae (1 → 3)(1 → 6)-β-D-glucan. The protective efficiency of CM-G against DNA damage in cells from patients with advanced prostate cancer (PCa), and undergoing Androgen Deprivation Therapy (ADT), was evaluated. DNA damage scores were obtained by the comet assay, both before and after treatment with CM-G. The reduction in DNA damage, ranging from 18% to 87%, with an average of 59%, was not related to the increased number of leukocytes in peripheral blood. The results demonstrate for the first time the protective effect of CM-G against DNA damage in patients with advanced PCa. Among smokers, three presented the highest reduction in DNA damage after treatment with CM-G. There was no observable relationship between DNA damage scores before and after treatment, and age, alcoholism and radiotherapy.

Key words: antioxidant, β-D-glucan, carboxymethyl-glucan, prostate cancer.

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β-D-glucan from the cell wall of Saccharomyces cerevisiae is a polymer composed of repeating glucose units, and organized into a central skeleton linked by β (1 → 3) glycosidic bonds, with side chains of variable sizes joined by β (1 → 6) linkages (Di Luzio et al., 1979). As the benefits arising from the biological activity of (1 → 3)(1 → 6)-β-D-glucan are limited by its insolubility in water, studies are under way with derivatives of this polymer in an attempt to improve its availability. Soluble derivatives have additional advantages when administered in vivo, such as the absence of toxicity and adverse effects (Miadaková et al., 2005). Moreover, derivatization may even enhance certain biological properties, such as antioxidant and immunomodulation activities (Chen and Seviour, 2007; Xu et al., 2009). Another important factor is the maintenance of bioactivity, since (1 → 3)(1 → 6)-β-D-glucan soluble derivatives can cross the gastrointestinal wall without causing damage to the digestive system, even when orally administered (Chorvatovicová et al., 1996; Rice et al., 2005).

Carboxymethyl-glucan (CM-G), one of the most studied among the soluble derivatives of S. cerevisiae (1 → 3)(1 → 6)-β-D-glucan, has been shown to possess significant bioprotective properties, such as antimutagenic, antigenotoxic, antioxidant, and anticancer activities (Chorvatovicová et al., 1996; Babincová et al., 2002; Slamenová et al., 2003; Miadaková et al., 2005). The main mechanism proposed for these protective effects of CM-G is the capability of scavenging reactive oxygen species (Kogan et al., 2005), even at concentrations as low as 30 μM (Babincová et al., 2002).

Prostate cancer (PCa) is the sixth most common cancer in the world, and represents about 10% of all cancers diagnosed. In Brazil, it is estimated that 49,530 new cases were diagnosed in 2009. Excluding non-melanoma skin cancer, it is the most frequent type of cancer in males and is prevalent countrywide, and counts with the second highest mortality rate in Brazil (National Cancer Institute of Brazil, 2010).

While a man with localized PCa is more likely to die from other comorbidities, the probability of death from PCa exceeds other causes when clinical metastases occur. At this stage, the aims of medical treatment are to increase the period of short-term survival of patients and, above all, to improve the quality of life, which often declines with progression of the disease. The standard treatment for advanced PCa is Androgen Deprivation Therapy (ADT), during which the administration of hormone analogues of a
gonadotropin-releasing hormone (GnRH) leads to receptor desensitization, thus inhibiting the release of LH and FSH, and resulting in chemical castration (Hellerstedt and Pienta, 2002).

The complications arising from chronic treatment, in addition to advanced age and fixed lifestyle habits, make patients with advanced PCAs even more vulnerable. Therefore, the use of innocuous, naturally bioactive compounds, capable of improving the quality of life of these patients can be a befitting alternative. Simulations of reactions that occur in the human body due to the administration of CM-G have been reported in several experimental models, although evaluation in humans is scarce. Our goal in this study was to evaluate the possible protective effects of CM-G against DNA damage in peripheral lymphocytes of patients with advanced PCAs. The nonrandomly selected target population expressed a clinical cancer scenario aggravated by age and lifestyle habits.

This study was undertaken after receiving approval from the Ethics Committee for Research Involving Humans, under registration number NTC406800007 in the records of the Brazilian Information on Ethics in Research Involving Humans (SISNEP). Patients at the Londrina Cancer Institute (ICL), with a histological diagnosis of stage T3 (n = 7) and T4 (n = 13) prostate adenocarcinoma according to TNM staging (AJCC/UIAC, 2010), and undergoing androgen deprivation therapy (ADT) with goserelin acetate, were included in this study, after signing a written informed-consent. Individual information was collected by a face-to-face questionnaire (Carrano and Natrajana, 1988). The patients were kept under observation in order to check for possible side effects from CM-G administration.

CM-G with substitution degree (DS) 0.8 was obtained from S. cerevisiae insoluble β-glucan (Magnani et al. 2009). 20 mg of CM-G, with starch as the excipient vehicle, were inserted into gelatin capsules. This procedure was carried out by Vico Farma® (National Health Surveillance Agency in Brazil number 136420-0). Bottles containing between 28 and 35 capsules were labeled with instructions for use. The variable number of capsules was to aid in controlling consumption by patients, return of the bottle being requested after the end of treatment. After daily administration of one capsule of CM-G (Demir et al., 2007), during a period of 28 days, the returned bottle should contain between zero and seven capsules, depending on the initial amount received by each patient.

Peripheral venous blood samples were collected, in a state of fasting, on the 1st day of treatment before ingestion of the first capsule of CM-G, and once again after 28 days of daily administration. The patients were arranged into couples, with no more than two patients having the same 1st or last day of ingestion. Vacutainer tubes of 4.5 mL containing EDTA were used for collecting samples. Leukocyte counts were obtained by an automated method (Abbott Cell Dyn 3200). Samples for comet assay were kept under refrigeration and protected from light during transport from the ICL to the Laboratory of Mutagenesis and Oncogenetics at the University of Londrina, Parana State, Brazil. Comet assays according Singh et al. (1988), with slight modifications, were conducted immediately after arrival of the samples in the laboratory. For the preparation of fresh-blood slides, 20 µL of peripheral blood was mixed with 180 µL of 0.5% low melting temperature agarose, and applied to microscope slides pre-coated with 1.5% normal melting temperature agarose. The slides were coverslipped and refrigerated for 5 min at 4 °C. Duplicate coded slides were prepared for each sample, and the samples were processed in a randomized manner. Cells attached to the agarose matrix underwent lysis by immersion in an ice-cold solution containing 1% Triton X-100, 10% DMSO, 2.5 M NaCl, 100 mM EDTA, and 10 mM Tris (pH 10.0), for at least 1 h. The DNA was subsequently denatured for 20 min in an alkaline buffer (300 mM NaOH and 1 mM EDTA, final pH > 13) and electrophoresed for 20 min at 25 V and 300 mA (1.25 V/cm) in the same buffer. Subsequently, the slides were neutralized (400 mM Tris, pH 7.5) and fixed with 100% ethanol for 10 min. For visual analysis, the slides were coded and stained with ethidium bromide (20 mg/mL), whereupon one hundred cells per patient were immediately scored at 400× magnification using a fluorescence microscope (Nikon-Brazil) with a blue (488 nm) excitation filter and yellow (515 nm) emission (barrier) filter. A single person person was assigned to analyzing the slides throughout the study by visual scoring. The cells were classified into four categories, each representing different degrees of DNA damage, these ranging from no visible migration (class 0, undamaged cells) to maximum length comet (class 3, maximally damaged cells (DC) (Kobayashi et al., 1995; Forchhammer et al., 2008). DC frequency was obtained by first determining the number of undamaged (class 0) and DCs from each of classes 1, 2 and 3, and then dividing this by the total number of cells analyzed in each treatment. The total score for 100 nucleoids was obtained by multiplying the number of cells in each class by the damage class (Manoharan and Banerjee, 1985): Total score = (0 × n₀) + (1 × n₁) + (2 × n₂) + (3 × n₃), where n = number of cells in each class analyzed. Thus, the total score could range from 0 (100 cells presenting no damage) to 300 (all cells presenting class 3 damage). The damage reduction rates for each patient were determined based on scores observed before and after CM-G.

The results were analyzed with MEDCAL 9.2 software using the non-parametric Mann-Whitney test for DNA damage scores before and after CM-G treatment, and the Wilcoxon test for leukocyte counts, considering significance at p ≤ 0.05.

After completing treatment with CM-G, the number of returned capsules patients corresponded to the use of 28 capsules in the period. Subsequently, and through weekly
contact, any side effects that could be associated with the use of CM-G were reported. This included typical symptoms of PCa under chronic treatment for testosterone inhibition, such as urinary symptoms and sexual dysfunction. The bone pain was reported by 90% of the patients. These effects of ADT occur due to the loss in bone mineral density (BMD), worsened by advanced age (Diamond et al., 2004).

As to diet, all men mentioned the eating of red meat and salad on an average of five and three times a week, respectively. Furthermore, 65% of the patients reported unintentional weight-loss. When questioned, they responded that the reduction in food intake was attributed to symptoms such as change in taste, dryness of the mouth, vomiting, constipation and nausea. Except for two patients, who reported a decrease in nausea, these symptoms persisted during the treatment with CM-G.

Data for each patient, obtained from medical records and the individual questionnaire (Table 1), show the various individual lifestyle characteristics, such as smoking, age, alcoholism, and exposure to chemicals and radiation, that exerted an influence on the baseline of DNA damage.

The analysis of data on DNA damage scores, obtained both before and after treatment with CM-G, showed significant differences ($p \leq 0.05$), with DNA damage reduction rates ranging from 18% to 87%, with an average of 59% (Figure 1). The number of DCs in classes 1, 2 and 3 decreased significantly ($p \leq 0.05$) after CM-G treatment. On the other hand, there was an increase in the number of undamaged cells for all patients (Figure 2). Based on reports of increased stimulation of hematopoiesis by CM-G (Possisil et al., 1991), the reduction in DNA damage could be attributed to an increase in the number of cells. In this way, the average numbers of white blood cells counted before $(1,789.25 \pm 766.1$ per mm$^3$) and after $(1,827.9 \pm 757.8$ per mm$^3$) CM-G administration were not statistically different ($p = 0.63$).

Five patients (25%) were classified as current smokers. Nicotine, the most toxic component of cigarettes, is known to cause oxidative stress by inducing the generation of reactive oxygen species (ROS) (Sener et al., 2007). It is known that CM-G is a potent antioxidant, which, besides being effective even at very low concentrations, plays an important role in protecting biological membranes against oxidative stress.

Table 1 - Data obtained from medical reports of 20 patients of the Cancer Institute of Londrina, Parana State, and from questionnaires completed after formal written consent and prior to administration of carboxymethyl-glucan (CM-G).

<table>
<thead>
<tr>
<th>Patient</th>
<th>Age (years)</th>
<th>Ethnicity</th>
<th>TNM</th>
<th>Goserelin use (months)</th>
<th>Life style</th>
<th>Agrochemicals *</th>
<th>Radiotherapy</th>
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</table>

*aPatients who had quit smoking for less than one year are considered smokers.

bPatients with behavior in alcohol-dependence.

*Yes = patients who have had contact or exposure to agrochemicals for $\geq 10$ years.

*Patients who reported intoxication by agrochemicals.
adverse effects of free radicals (Babincová et al., 2002). Sener et al. (2007) suggested that the oral administration of soluble β-D-glucan from S. cerevisiae is effective against chronic nicotine intoxication, thereby reducing oxidative damage. Our data is consistent with these findings, since among the patients appraised, three smokers (5, 19 and 20) showed the three highest rates of reduction in DNA damage after treatment with CM-G.

Eight men (40%) showed alcohol dependence. Ethanol itself does not induce lesions in the DNA of human lymphocytes, but its primary metabolite, acetaldehyde, evokes breaks in single- and double-stranded DNA (Singh et al., 1995). This, together with other factors that generate DNA damage, may have contributed to some of the highest rates of damaged DNA observed in these patients 1, 5, 8 and 10 before CM-G.

Six patients (30%) underwent radiotherapeutic treatment prior to starting ADT (Table 1) in order to control localized prostate tumors. The exposure of patients with cancer to therapeutic doses of radiation typically causes free-radical induced changes in the genomic DNA of lymphocytes, (Olinski et al., 1996). Moreover, the generation of free radicals in the tissues may continue after exposure for several months (Robbins and Zhao, 2004). The high radical scavenging activity of CM-G from S. cerevisiae, with DS 0.8, was confirmed by paramagnetic resonance spectroscopy (Kogan et al., 2005). Therefore, CM-G could be of help in decreasing the oxidative damage caused by radiotherapy.

Of the total number of patients, 85% had been occupationally exposed for at least 10 years previously, to a mixture of agrochemicals (Table 1), such as organophosphates, carbamates and organochlorines. This includes DDT and BHC, both banned in Brazil for over 20 years, due to high toxicity.

We believe that treatment with CM-G can be an efficient protection against DNA damage caused by mutagenic compounds through dismutagenic activity, either by its capability of blocking the action of damage-inducing agents, or by bio-antimutagenic activity, which exerts its effect by preventing damage or promoting DNA repair, since Oliveira et al. (2007) showed that this is a characteristic of (1 → 3)(1 → 6)-β-D-glucan from S. cerevisiae.

Prostate carcinogenesis may be related to chronic and acute prostate inflammation (De Marzo et al., 2007). The possible chemoprotective effect of antioxidants may be a form of protecting prostate cells from the DNA damage that normally occurs in the setting of inflammatory lesions (Smigel, 1998). Thus, there is a possibility that the DNA damage generated by the typical inflammation process was reduced through the administration of CM-G.

The results presented here highlight the protective effect of CM-G against DNA damage, which was shown for the first time in humans. These results suggest that CM-G is potentially useful for improving the short-term survival of patients with advanced PCa. Studies are underway on the further beneficial effects of CM-G administration, such as those related to immunomodulation, to assess the possibility of CM-G use improving prognosis in these patients.

Acknowledgments

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