Fruit yield and bacterial wilt symptoms on eggplant genotypes grown in soil infested with *Ralstonia solanacearum*

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**ABSTRACT**

Fruit yield and bacterial wilt symptoms of eggplant genotypes CNPH006, CNPH171, CNPH658, CNPH778, CNPH783, and CNPH785 were quantified in soil naturally infested with *Ralstonia solanacearum* (race 1, biovar 3) in Brasília, Brazil. Except for CNPH778, all genotypes developed at least one typical wilted plant. Besides wilt, other symptoms considered for assessing levels of tolerance/resistance among genotypes were plant death (CNPH006, CNPH171, CNPH658 and CNPH783), leaf chlorosis (CNPH785) and plant dwarfing (all genotypes). The occurrence of bacterial ooze on cut stems was observed in all plants grown in infested beds, including those that did not exhibit wilt symptoms. Plants grown simultaneously in noninfested beds of the same dimensions and soil characteristics allowed the comparison of yield losses due to the pathogen. The genotypes were grouped according to their ability to maintain fruit production in the infested area. CNPH785 was the most tolerant genotype, with no significant loss due to the disease, to maintain fruit production in the infested area. CNPH785 was the most tolerant genotype, with no significant loss due to the disease, followed by CNPH783, CNPH778 and CNPH171, with mean losses of 19.3%, 11.4%, and 10.1%, respectively. The genotypes CNPH658 and CNPH006 were the most susceptible, with average losses of 99.53 and 99.32%, respectively.

**Keywords:** *Solanum melongena*, resistance, losses.

**Palavras-chave:** *Solanum melongena*, murcha bacteriana, perdas.

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is dominated by hybrids due to their productivity characteristics, resistance to diseases and pests, uniformity and fruit quality (Antonini et al., 2002).

The occurrence of bacterial wilt, caused by *Ralstonia solanacearum*, is one of the limiting factors for growing solanaceous crops in warm and humid climates, such as in the Amazon Region. The local production is practically restricted to smallholder’s gardens which, after a few successive crops, are no longer suitable for growing many vegetables, especially solanaceous. For instance, losses of up to 40% due to bacterial wilt were reported in a commercial eggplant field in the state of Amazon (Coelho-Netto et al., 2004). The disease also has a surmount importance for the cultivation of solanaceous in the Northeastern and Mid-Western lowland regions (Takatsu & Lopes, 1997).

The most typical symptom of bacterial wilt of eggplant is a wilt starting from the upper parts of the plant, observed initially in the warmer periods of the day, with leaf turgidity often recovering overnight in cooler or rainy days. If favorable environmental conditions prevail, diseased plants complete wilt and die (Ribeiro, 2007; Lopes, 2009). Leaf epinasty and adventitious root formation due to the imbalance in the levels of auxin and ethylene eventually occur (Buddenhagen & Kelman, 1964).

*Ralstonia solanacearum* enters the plant through mechanical wounds or micro-wounds caused by the growth of secondary roots, followed by xylem invasion (Vasae et al., 1995). Xylem colonization induces partial or complete mechanical blockage of the water transport from the roots to the top of the plant. The formation of tyloses and the increased levels of ethylene and abscisic acid are associated to the infection process (Kelman, 1953). Xylem vessels can hold concentration above $10^{10}$ cells per centimeter of susceptible tomato stem which, with extracellular polysaccharides produced by the pathogen, increase the sticky consistency of the xylem fluid, hindering its flow toward the leaves, thus causing wilting and the death of the plant (Genin & Boucher, 2002). After plant death, the bacterium infects the soil where it survives saprophytically and infects new plants (Genin & Boucher, 2002). Extracellular polysaccharides produced by the pathogen, besides forming biofilms on the surface of the xylem during disease development, also prevent desiccation or antibiotic in the absence of a host plant, what partially explains the long survival of the pathogen in the soil (Milling et al., 2011; Hikichi et al., 2007; Genin & Boucher, 2002).

When compared to other solanaceous hosts, eggplant is not as susceptible to bacterial wilt as potatoes, tomatoes and bell peppers. Under less favorable conditions for the plant, plants may not wilt, but grow more slowly than the healthy ones (Ribeiro, 2007). Therefore, when classical symptoms of bacterial wilt are not observed, diagnosis of the disease and the selection of resistant genotypes can be complicated.

Despite the large amount of work on bacterial wilt, reports that estimate yield losses due to this disease under natural soil infection are scarce, especially considering the variability of resistance on host plants. The criterion mostly used to quantify the resistance is the survival rate under infection pressure, but, in almost all cases, the latent infection or colonization is not taken into account (Boshou, 2005). In greenhouse tests, eggplant genotypes reported as resistant to bacterial wilt based on the wilting characteristic, displayed different symptoms, including dwarfism, suggesting that yield losses in the field are real also for nonwilting plants (Morgado, 1991; Oliveira, 2011).

The objective of this work was to evaluate, in a soil naturally infested by *R. solanacearum*, the plant growth, the bacterial wilt symptoms and the yield capacity of eggplant genotypes previously identified as resistant to bacterial wilt in the greenhouse.

### MATERIAL AND METHODS

The experiments were carried out from September 1999 to March 2000 at Embrapa Hortaliças, Brasilia, Brazil, where the average monthly rainfall was 154.7 mm and mean temperature of 23.9°C. The experimental plots consisted of a set of 10x1 m beds surrounded by a 15 cm high borders. Part of the beds, originally used for seedling production, was built on a soil spot naturally infested with *R. solanacearum* race 1, biovar 3. In order to improve the uniform infestation of the soil of each of five beds, three rows of tomato seedlings ‘Ponderosa’, susceptible to bacterial wilt, were transplanted spaced 25x25 cm. Prior to planting, the soil of the five infested and five noninfested beds were analyzed, treated with a soil insecticide, corrected with dolomitic lime and fertilized with NPK (10-10-10) and cow manure, according to the recommendation for tomato (CFSEMG, 1989). After 30 days of transplanting, all the tomato plants in the infested beds showed symptoms of bacterial wilt and were mechanically incorporated into the soil.

Thirty days after bed preparation, which included weekly sprinkler irrigation to maintain the soil slightly humid, seedlings of eggplant grown in polystyrene trays with sterilized substrate, 35 days after sowing, were transplanted in rows 50 cm apart and 80 cm between plants. In each infested and noninfested bed, four plants of six genotypes with different levels of susceptibility to *R. solanacearum* were transplanted (Table 1). The plants were irrigated with a micro sprinkler to keep the soil moisture close to field capacity.

The experimental design consisted of randomized blocks with five replications, each bed being a block. Analyses of variance were used to compare the performance of the six eggplant genotypes in the infested beds with the statistical program Winstat 1.0 (Machado & Conceição, 2005). In this same application, the residual analysis to verify the assumptions of the analysis of variance was also performed: box plot, normal distribution and dispersion error.

Mean yield of genotypes were compared by Tukey test ($p<0.05$). The yield reduction of each genotype was calculated in relation to the mean yield of the respective genotype upon simultaneous cultivation in the four
noninfested beds. The Scott-Knott group test (p≤0.05) was performed for this comparison using GENES software (Cruz, 2006).

The ordinary bacterial wilt symptoms expected for eggplant infection, i.e. yellowing, dwarfism, wilting, and death, were observed during crop cycle upon weekly monitoring up to 130 days after transplanting. Fruit production (kg/plant) of each plot was taken in five harvests, started 70 days after transplanting with an interval of 15 days between harvests. Wilted plants were sampled in order to certify the causal agent of the disease. After the last harvest, all the remaining plants of the two areas were cut and stem fragments were removed 5 cm above the soil line for observing the bacterial flow oozing out from the stem in a glass, to confirm the bacterial etiology (Lopes & Quezado-Soares, 1997). Bacterial isolation was performed in Kelman’s culture medium with tetrazolium, in which *R. solanacearum* colonies can be partially identified after cultivation for 48 hours at 28°C (Kelman, 1954). Fluidal colonies with red centers and white edges, typical of this bacterium in Kelman’s medium, were transferred to new plates containing the same medium devoid of tetrazolium to allow faster growth. Putative colonies of *R. solanacearum* were submitted to biochemical tests to determine the biovar, according to the ability to oxidize sole carbon sources in a minimal medium (Hayward, 1991). The isolates obtained and identified were stored in tubes with sterile water at room temperature.

**RESULTS AND DISCUSSION**

Symptoms associated to bacterial wilt developed only in plants cultivated in infested beds. The typical wilting was first observed 10 days after transplant in genotypes CNPH006 and CNPH658. Except for CNPH778, all genotypes yielded at least one wilted plant (Table 2). Evolution from wilting to plant death was observed in genotypes CNPH006, CNPH171, CNPH783 and CNPH658, and leaf chlorosis in CNPH785. Plant dwarfism was present in all genotypes when compared to controls cultivated in noninfested beds.

The bacterial exudation tests indicated the presence of xylem-associated bacteria in the remaining

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**Table 1. Set of Solanum melongena genotypes and their previously known reactions1 to Ralstonia solanacearum strains (procédência e reações3 à três estirpes de Ralstonia solanacearum de genótipos de Solanum melongena). Brasília, Embrapa Hortaliças, 2000.**

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Origin</th>
<th>R. solanacearum strains</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>CNPH562</td>
</tr>
<tr>
<td>CNPH006</td>
<td>Agroflora (Brazil-SP)</td>
<td>R</td>
</tr>
<tr>
<td>CNPH171</td>
<td>INRA (France)</td>
<td>HR</td>
</tr>
<tr>
<td>CNPH658</td>
<td>Topseed (USA)</td>
<td>S</td>
</tr>
<tr>
<td>CNPH778</td>
<td>AVRDC (India)</td>
<td>nd</td>
</tr>
<tr>
<td>CNPH783</td>
<td>AVRDC (Indonesia)</td>
<td>nd</td>
</tr>
<tr>
<td>CNPH785</td>
<td>AVRDC (India)</td>
<td>nd</td>
</tr>
</tbody>
</table>

1HR: high resistance; R: intermediate resistance; S: susceptible; nd: not determined (HR: resistência elevada; R: resistência intermediária; S: suscetível; nd: não determinado); Biovar 3. Fonte: Morgado, 1991; 3Biovar 1. Fonte: Oliveira, 2011)

**Table 2. Wilt Incidence (W) and plant death (D) in eggplant genotypes at 10, 50, 90 and 130 days after transplanting to beds naturally infested with Ralstonia solanacearum, race 1, biovar 3 [incidência (%) de sintoma de murcha (W) e morte (D) em plantas de genótipos de berinjela, aos 10, 50, 90 e 130 dias após transplante em área infestada naturalmente com Ralstonia solanacearum, raça 1, biovar 3]. Brasília, Embrapa Hortaliças, 2000.**

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Days after transplant</th>
<th>10</th>
<th>50</th>
<th>90</th>
<th>130</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>W</td>
<td>D</td>
<td>W</td>
<td>D</td>
<td>W</td>
</tr>
<tr>
<td>CNPH658</td>
<td>5</td>
<td>0</td>
<td>55</td>
<td>5</td>
<td>100</td>
</tr>
<tr>
<td>CNPH006</td>
<td>10</td>
<td>0</td>
<td>85</td>
<td>0</td>
<td>100</td>
</tr>
<tr>
<td>CNPH171</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>5</td>
</tr>
<tr>
<td>CNPH778</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>CNPH783</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>15</td>
</tr>
<tr>
<td>CNPH785</td>
<td>0</td>
<td>0</td>
<td>5</td>
<td>5</td>
<td>10</td>
</tr>
</tbody>
</table>

**Table 3. Yields (kg/plant) of eggplant genotypes cultivated in soils noninfested and infested with Ralstonia solanacearum, race 1, biovar 3 {produção (kg de frutos/planta sobrevivente) de genótipos de berinjela em área livre e em área infestada por Ralstonia solanacearum, raça 1, biovar 3]. Brasília, Embrapa Hortaliças, 2000.**

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Yield1</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Noninfested</td>
</tr>
<tr>
<td>CNPH658</td>
<td>3.66 b c</td>
</tr>
<tr>
<td>CNPH006</td>
<td>6.16 a</td>
</tr>
<tr>
<td>CNPH171</td>
<td>3.14 c</td>
</tr>
<tr>
<td>CNPH778</td>
<td>5.28 a b</td>
</tr>
<tr>
<td>CNPH783</td>
<td>2.70 c</td>
</tr>
<tr>
<td>CNPH785</td>
<td>1.98 c</td>
</tr>
<tr>
<td>CV (%)</td>
<td>22.12</td>
</tr>
</tbody>
</table>

1Means of five replicates; Means followed by the same letter on columns do not differ by Tukey test (p≤0.05) [médias de 5 repetições; Valores na coluna seguidos pelas mesmas letras não diferem pelo teste de Tukey (p≤0.05)].
Figure 1. Fruit yield maintenance (%) on eggplant genotypes grown in soil naturally infested with *Ralstonia solanacearum*, race 1, biovar 3, in relation to yield mean in noninfested soil. Columns topped by the same letters formed a group that does not differ by Scott-Knott clustering test (p ≤ 0.05) (manutenção na produção de frutos (%) de genótipos de berinjela cultivados em solo naturalmente infestado por *Ralstonia solanacearum*, raça 1, biovar 3, em relação à média de produção em solo livre do patógeno. As colunas com mesmas letras, no topo, formam grupos que não diferiram entre si, segundo o teste de agrupamento de Scott-Knott (p≤0.05). Brasília, Embrapa Hortaliças, 2000.

The genotype CNPH658 (Florida Market) was confirmed as a good susceptible control for studies on eggplant bacterial wilt resistance or on the pathogen’s isolate virulence, as suggested by Morgado (1991). In contrast, CNPH006, which, in greenhouse tests by Morgado (1991) and Oliveira (2011) showed variable results and strain-dependent response, was as susceptible as the genotype CNPH658 in our experiments. However, its high yield, dark color and elongated fruit, which are the characteristics demanded by the Brazilian market, makes this genotype be considered as a recurrent parent in a future breeding work.

CNPH778 was the most resistant genotype in our trials given by disease incidence readings, but presented yield reduction in the infested as compared to noninfested beds (Table 2). Despite this reduction, it was the most productive genotype in the infested beds. The genotype CNPH785, despite being infected by *R. solanacearum* and displaying wilting, leaf chlorosis and plant dwarfism when grown in the infested beds, yielded the same fruit weight as in the noninfested beds, therefore indicating tolerance to the disease. Bacterial wilt tolerance was also described by Gousset et al. (2005) in *Solanum torvum* genotypes collected in Java.

Gousset et al. (2005), in studies on resistance in *S. torvum*, conjectured that the resistance of some genotypes to *R. solanacearum* could occur by limiting the multiplication or spread of the bacterial population in the stem. In fact, Oliveira (2011) quantified the multiplication rate of two *R. solanacearum* isolates in the stem of the CNPH778 and CNPH785, and found that the colonization in these genotypes was lower when compared to more susceptible genotypes, CNPH658 and CNPH171. The two resistance-related mechanisms, by supporting higher population of the pathogen and eggplant is dependent on biovar and isolate of the pathogen (Lopes & Boiteux, 2004; Lopes et al. 1994; Wicker et al., 2007; Coelho Netto et al., 2004; Gopalakrishnan et al., 2005; Gousset et al., 2005; Grimault & Prior, 1994; Milling et al., 2011).

Based on differences in yield reduction, the genotypes were separated in three groups: susceptible (CNPH 006 and CNPH 658), moderately resistant (CNPH 171, CNPH783 and CNPH778) and resistant (CNPH785), according to Scott-Knott test (Figure 1). The low reduction in fruit production observed for CNPH171 is in accordance with results obtained for biovar 1 in a greenhouse by Morgado (1991), when this genotype was indicated as the most promising among a set of accesses. This genotype, however, showed to be susceptible in a test in the greenhouse when inoculated with the strain CNPH19 of *R. solanacearum*, which belongs to biovar 1 (Oliveira, 2011). This behavior is explained in terms of the interaction already observed among eggplant genotypes and strains of the pathogen (Oliveira, 2011). This fact is reinforced by previous studies indicating that resistance to bacterial wilt in tomato, *Capsicum* peppers

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without yield loss (tolerance) or by depriving the bacterium to multiply or spread in the host, both of quantitative genetic control, can be exploited by plant breeders to fight the bacterial wilt disease in eggplants.

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