Diminution of Oxalate Induced Renal Tubular Epithelial Cell Injury and Inhibition of Calcium Oxalate Crystallization in vitro by Aqueous Extract of Tribulus terrestris

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ABSTRACT

Purpose: Recurrence and persistent side effects of present day treatment for urolithiasis restrict their use, so an alternate solution, using phytotherapy is being sought. The present study attempted to evaluate the antilithiatic properties of Tribulus terrestris commonly called as “gokhru” which is often used in ayurveda to treat various urinary diseases including urolithiasis.

Materials and Methods: The activity of Tribulus terrestris was investigated on nucleation and the growth of the calcium oxalate (CaOx) crystals as well as on oxalate induced cell injury of NRK 52E renal epithelial cells.

Results: Tribulus terrestris extract exhibited a concentration dependent inhibition of nucleation and the growth of CaOx crystals. When NRK-52E cells were injured by exposure to oxalate for 72 h, Tribulus terrestris extract prevented the injury in a dose-dependent manner. On treatment with the different concentrations of the plant, the cell viability increased and lactate dehydrogenase release decreased in a concentration dependent manner.

Conclusion: The current data suggests that Tribulus terrestris extract not only has a potential to inhibit nucleation and the growth of the CaOx crystals but also has a cytoprotective role. Our results indicate that it could be a potential candidate for phytotherapy against urolithiasis.

Key words: phytotherapy; urolithiasis; calcium oxalate; NRK 52E, Tribulus terrestris

INTRODUCTION

Nephrolithiasis is common, affecting up to 10% of the population at some point during their lifetime (1). Calcium-containing stones are the most commonly occurring to an extent of 75-90% followed by magnesium ammonium phosphate (Struvite) to an extent of 10-15%, uric acid 3-10% and cystine 0.5-1% (2). Calcium oxalate stones are found in two different varieties, calcium oxalate monohydrate (COM) or Whewellite, and calcium oxalate dihydrate (COD) or Weddellite. COM, the thermodynamically most stable form, is observed more frequently in clinical stones than COD and it has a greater affinity for renal tubular cells, thus responsible for the formation of stones in the kidney (3).

Various authors have suggested the role of crystal induced cell injury in the development of
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kidney stones by providing the sites for crystal attachment and retention within the kidneys (4,5).

Oxalate, a metabolic end product and a major constituent of the majority of renal stones, has been shown to be toxic to renal epithelial cells of cortical origin (6). It has been observed that exposure of renal epithelial cells to oxalate which is a constituent of most kidney stones leads to a disruption of the normal activities of the renal epithelial cells such as altered membrane surface properties and cellular lipids, changes in gene expression, disruption of mitochondrial function, formation of reactive oxygen species and decreased cell viability (7).

Various mechanisms have been proposed to explain crystal retention (8). As a result of crystal growth and agglomeration, particles may be formed that are too large to freely pass the renal tubules. Alternatively, relatively small crystals could be retained by adhering to the surface of the urothelial lining and then increase in size (8).

The surgical methods available to treat kidney stones like extracorporeal shock wave lithotripsy have serious side effects. Therefore, it is worthwhile to look for an alternative for the management of urolithiasis. Many medicinal plants have been employed during ages to treat urinary stones though the rationale behind their use is not well established through systematic and pharmacological studies, except for some composite herbal drugs and plants (9-12). Plant medicines are in great demand both in the developed as well as developing countries for primary health care because of their wide range of biological and medicinal activities, higher safety margin and low cost.

Fruits of Tribulus terrestris (Zygophyllaceae) locally named as “gokhru” in India are commonly used in folklore to treat urolithiasis. So far, its diuretic properties have been documented in literature and it is actively used in various drug formulations of kidney stone treatments.

The present study aimed at investigating the efficacy of Tribulus terrestris on calcium oxalate crystal nucleation and growth in vitro as well as further examining the potency of Tribulus terrestris on oxalate induced injury in NRK 52E (rat renal tubular epithelial) cells.

MATERIALS AND METHODS

Preparation of the Tribulus terrestris Extract

The dried and matured fruits of Tribulus terrestris were obtained from “Natural Remedies Pvt. Ltd.” at Bangalore in India. A collection of voucher specimens is available at the company.

The air-dried fine powdered plant fruits were boiled in distilled water. The extract was then filtered using Whatman No. 1 filter paper and the filtrate was evaporated in vacuum and dried using a rotary evaporator at 60°C (13). The final dried samples were stored in labeled sterile bottles and kept at -20°C. The various concentrations of the plant sample tested for their inhibitory potency were 25 µg/mL, 50 µg/mL, 100 µg/mL, 200 µg/mL, 400 µg/mL and 1000 µg/mL, which were prepared at the time of experiment and were referred to as aqueous extract of Tribulus terrestris.

For cell culture studies a stock solution of the dried aqueous Tribulus terrestris extract was dissolved in dimethyl sulfoxide (DMSO) [final concentration of the DMSO in the highest concentration of plant extract tested did not exceed 0.4% (v/v) and did not affect the cell proliferation]. Further dilutions of the stock were done using serum free DMEM (Dulbecco’s Modified Eagle’s Media) and filtered by 0.3 mm syringe filter (14).

Nucleation Assay

The method used was similar to that described by Hennquin et al. with some minor modifications (15). Solutions of calcium chloride and sodium oxalate were prepared at the final concentration of 3 mmol/L and 0.5 mmol/L, respectively, in a buffer containing Tris 0.05 mol/L and NaCl 0.15 mol/L at pH 6.5. Both solutions were filtered through a 0.22 µm filter; 33 mL of calcium chloride solution was mixed with 3.3 mL of the aqueous extract at different concentrations. Crystallization was started by adding 33 mL of sodium oxalate solution. The final solution was magnetically stirred at 800 rpm using a PTFE-coated
stirring bar. The temperature was maintained at 37°C. The absorbance of the solution was monitored at 620 nm after every 1 min. The percentage inhibition produced by the herb extract was calculated as \[1 - (T_{si}/T_{sc})\] X 100, where T_{sc} was the turbidity slope of the control and T_{si} the turbidity slope in the presence of the inhibitor.

**Growth Assay**

Inhibitory activity against CaOx crystal growth was measured using the seeded, solution-depletion assay described previously by Nakagawa and colleagues (16). Briefly, an aqueous solution of 10 mM Tris-HCl containing 90 mM NaCl was adjusted to pH 7.2 with 4 N HCl. Stone slurry (1.5 mg/mL) was prepared in 50 mM sodium acetate buffer (pH 5.7). CaOx monohydrate crystal seed was added to a solution containing 1 mM CaCl_2 and 1 mM sodium oxalate (Na2C2O4). The reaction of CaCl_2 and Na2C2O4 with crystal seed led to deposition of CaOx (CaC2O4) on the crystal surfaces, thereby decreasing free oxalate that is detectable by spectrophotometry at λ214 nm. When aqueous extract is added into this solution, depletion of free oxalate ions will decrease if the test sample inhibits CaOx crystal growth. Rate of reduction of free oxalate was calculated using the baseline value and the value after 30-second incubation with or without test sample. The relative inhibitory activity was calculated as follows:

\[
\text{Relative inhibitory activity} = \frac{(C-S)\times 100}{C}
\]

where C is the rate of reduction of free oxalate without any test sample and S is the rate of reduction of free oxalate with a test sample.

**Cell Culture**

Normal rat epithelial derived renal tubular epithelial (NRK 52E) cells were obtained from National Centre of Cell Sciences (NCCS, Pune). The cells were maintained as monolayers in Dulbecco’s Modified Eagle’s Medium (DMEM) with 2.0 mM L-glutamine adjusted to contain 3.7 g/L sodium bicarbonate, 4.5 g/L glucose. Media was supplemented with 1% Penicillin (100 units/mL)-Streptomycin (10,000 µg/mL) and 10% fetal bovine serum. Cells were cultured in 25 cm² tissue-culture treated flasks at 37°C and 5% CO² in humidified chambers.

**Oxalate-induced Cell Injury**

NRK 52E cells were incubated in DMEM containing 1 mM sodium oxalate in the presence of different concentrations of the aqueous extract of the test sample (10 µg/mL, 25 µg/mL and 50 µg/mL) for 72 hours (14,17). Cell injury was assessed by measuring the cell viability through trypan blue and monitoring the lactate dehydrogenase (LDH) leakage into the medium.

**Cytotoxicity - Trypan Blue Assay**

The cytotoxicity of the aqueous extract of T. terrestris was assessed by cell viability using trypan blue exclusion method. For the determination of cell viability, cells were plated at the density of 4 x 10^4 cells/well and cultured for 72 h. The medium was replaced with serum-free medium and the cells were treated with various concentrations of the plant extracts (10 µg/mL, 25 µg/mL and 50 µg/mL) for a further 72 h. The percentage viability for the cells was calculated as (live cells/total cells) x 100.

**LDH Leakage Assay**

LDH leakage assay was performed by the method described by Wagner et al. (18). Briefly, 6.6 mM NADH and 30 mM sodium pyruvate were prepared in Tris (0.2M, pH 7.3). Reaction was initiated with the addition of 50 µL of the test sample and the disappearance of NADH was monitored at 340 nm, for 5 min at an interval of 1 min. The percentage of LDH release was calculated by dividing the activity of LDH in the supernatant by the LDH activity measured after complete cell lysis achieved by sonication.

**Statistical Analysis**

Data were expressed as mean values of three independent experiments (each in triplicate) and ana-
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analyzed by the analysis of variance (p < 0.05) to estimate the differences between values of extracts tested.

RESULTS

Inhibition of Nucleation of CaOx Crystals by Tribulus terrestris Extract

Figure-1 displays the effect of the different concentration of the aqueous extract of Tribulus terrestris on the nucleation of calcium oxalate crystals. As regards control (with no plant sample), the percentage inhibition was constant at 71.4 ± 0.001 with increase in the concentration of Tribulus terrestris extract of 25 µg/mL, 50 µg/mL and 100 µg/mL. As the concentration of Tribulus terrestris extract was increased to 200 µg/mL, the percentage inhibition increased to 100 ± 0.001 but was reduced to 85.7 ± 0.002 for 400 µg/mL. The percentage inhibition was restored to 100 ± 0.001 with 1000 µg/mL of the extract.

Inhibition of CaOx Crystal Growth by Tribulus terrestris Extract

Figure-2 demonstrates the percentage inhibition shown by Tribulus terrestris on the calcium oxalate crystal growth. Tribulus terrestris extract showed inhibition in a concentration dependent manner. The percentage inhibition with 25 µg/mL of plant sample was 17.6 ± 0.004. With 50 µg/mL, 100 µg/mL and 200 µg/mL, the inhibition was almost constant in the range of 65-70% but inhibition increased significantly with 400 µg/mL and 1000 µg/mL of Tribulus terrestris extract to 126.4 ± 0.001 and 169.2 ± 0.001 respectively.

Diminution of Oxalate-induced Renal Tubular Epithelial Cell Injury by Tribulus terrestris Extract

Figure-3 depicts the protective effect of the aqueous extract of Tribulus terrestris towards the renal tubular epithelial cells. The oxalate induced a significant injury to the cells which could be ascertained by a decrease in viability from 100% in the controls (untreated cells) to 73.9%. However, the injury due to oxalate was significantly reduced in those cells treated with the Tribulus terrestris extracts. As the concentration of the extract increased from 10 µg/mL to 50 µg/mL, the percentage viability improved showing that the plant has an inhibitory activity towards the oxalate which caused injury to the renal cells in a concentration dependent manner. The plant extract alone (50 µg/mL, containing 0.4% DMSO) had no

![Figure 1 – Effect of Tribulus terrestris on nucleation of CaOx.](image-url)
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Effect on the cell injury in the absence of oxalate indicating that even at the highest concentration of DMSO used there was no cytotoxicity to the cells. The percentage viability with 10 µg/mL, 25 µg/mL and 50 µg/mL was 81.6 ± 6.9, 84.9 ± 1.9 and 89.1 ± 6.9 respectively.

Lactate dehydrogenase is a stable cytosolic enzyme that is released when the cell is lysed or there is any injury on the cell membrane. A significant increase in LDH release was seen when the NRK 52E cells were exposed to oxalate alone. When NRK 52E cells were treated with the plant extract at varying

Figure 2 – Effect of Tribulus terrestris on the growth of CaOx.

Figure 3 – Effect of Tribulus terrestris on the viability of NRK 52E. Data are mean ± SEM of three independent observations. * p < 0.05 versus untreated control, ** p < 0.05 versus oxalate control.
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concentrations (10, 25 and 50 µg/mL) along with oxalate (1 mM) for 72 h, a reduction in oxalate-induced cell injury was observed as assessed by a decreased LDH release (Figure-4). Again it was seen that the plant extract alone had no significant effect on the measures of cell injury in the absence of oxalate. The percentage LDH release for 10 µg/mL, 25 µg/mL and 50 µg/mL was observed to be 126.5 ± 4.2, 112.6 ± 5.2 and 109.8 ± 1.0 respectively after treatment with oxalate and the plant extract with respect to control.

COMMENTS

There is growing evidence that CaOx nephrolithiasis is associated with renal injury. Hyperoxaluria is a major risk factor for calcium oxalate nephrolithiasis, and calcium oxalate urinary stones are the most common type of urinary stone. High level of oxalate produced a variety of changes in the renal epithelial cells, such as an increase in free radical production and a decrease in antioxidant status, followed by cell injury and cell death. These changes are significant predisposing factors for the facilitation of crystal adherence and retention (5,14).

Due to significant side effects and failure to prevent recurrence by the present day treatment procedures for uroolithiasis, alternative treatment modalities using herbal products have assumed importance. A dramatic advancement in using phytotherapy for uroolithiasis treatments has been observed in recent years and many investigators have proposed to further scientific study on its efficacy. Many medicinal plants have been employed for centuries to treat urinary stones though the rationale behind their use is not well established.

In the present study, the anticalcifying properties of Tribulus terrestris commonly called “gokhru” were explored in vitro. The inhibitory potency of the plant was tested on the nucleation and growth of the most commonly occurring kidney stones, calcium oxalate monohydrate. A concentration dependent trend of inhibition was observed using Tribulus terrestris extract with maximum inhibition of 100% and 170% for CaOx nucleation and the growth assay respectively with 1000 µg/mL of the extract.

In our study with NRK 52E, Tribulus terrestris proved to have a protective effect towards the renal epithelial cells again in a concentration dependent manner. When NRK-52E cells were injured

*Figure 4 – Effect of Tribulus terrestris on the % LDH release. Data are mean ± SEM of three independent observations. *p < 0.05 versus untreated control, ** p < 0.05 versus oxalate control.
by exposure to oxalate for 72 h, the plant extract prevented the injury in a dose-dependent manner. The mechanism of inhibition/reduction in the injury needs to be studied further. Studies have shown that inhibition of the inflammatory response induced by injury due to crystal formation helps in restoring normalcy.

Beghalia et al. (19) have suggested in studies using certain Algerian medicinal plants that the herb extract may contain substances that inhibit the growth of COM crystals. This property of plant extracts could be important in preventing kidney stone formation; the agglomeration of particles is a critical step in urinary stone formation, as larger crystals are less likely to pass spontaneously in the urinary tract (8,20). They (19) further postulated that the plant extracts may contain substances that inhibit CaOx crystal aggregation and also the binding of the crystals to the renal epithelial surface. This could explain a decrease in LDH release as seen in the cells treated with the plant extract compared to those treated with oxalate alone.

Our studies are in agreement with the studies previously reported as regards the anti-urolithiatic potency of Tribulus terrestris on the growth COM crystals using double diffusion gel growth technique (21). The anti-urolithiatic ability of the plant is also currently being evaluated in animal models and has exhibited dose-dependent anti-urolithiatic activity and almost completely inhibited stone formation further supporting our results (22,23).

Recently several plants including Herniaria hirsuta (24), Phyllanthus niruri (25) and Bergenia linguata (26) are being explored for their anti-urolithiatic properties on the basis of their usage in the traditional medicine. Herniaria hirsuta, a plant from Morocco is also known to exhibit the antilithiatic activity. The adhesion of the radioactive COM crystals to the Madin Darby canine kidney cells was studied in the presence and the absence of the aqueous extract. COM crystal binding to the cells was inhibited by the extract in a concentration dependent manner (24). In vitro effect of an aqueous extract of Phyllanthus niruri L., a plant used in Brazilian folk medicine for the treatment of urolithiasis, on a model of CaOx crystal endocytosis by Madin-Darby canine kidney cells was investigated by Campos and Schor. The extract exhibited a potent and effective non-concentration-dependent inhibitory effect on the CaOx crystal internalization. This response was present even at very high (pathologic) CaOx concentrations and no Phyllanthus niruri L.-induced toxic effect could be detected (25). Bergenia ligulata is a widely used plant in South Asia, mainly India and Pakistan, as a traditional medicine for treatment of urolithiasis. The crude aqueous-methanolic extract of Bergenia ligulata rhizome was studied using in vitro and in vivo methods and the extract showed the anti-urolithic activity through CaOx crystal inhibition, diuretic, hypermagneseuric and antioxidant effects (26). Also in our laboratory, antilithiatic potency of Dolichos biflorus (27) and Trachyspermum ammi (28) has been evaluated in vitro and in vivo. The most active protein fraction was isolated from these plants and thus adds a new perspective to study plant fractions for their therapeutic use as antilithiatic proteins.

CONCLUSION

In conclusion, the aqueous extract of Tribulus terrestris has been shown to possess an ability to inhibit CaOx crystallization in vitro. In addition this extract has also shown cytoprotective properties towards the NRK 52E cells by lowering LDH leakage and increasing the cell viability. Our study suggests the possibility of using Tribulus terrestris as a therapeutic agent to treat urolithiasis and further characterization of its active compound(s) could lead to a new candidate drug for patients with urolithiasis.

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CONFLICT OF INTEREST

None declared.
REFERENCES


EDITORIAL COMMENT

Kidney stone disease is a major health problem in modern societies. As technology evolved, surgical options have gained more acceptance as they provide less invasive approaches, more efficacious results and lesser collateral effects. However, the costs involved are significant and an increasing effort should be continuously made in order to optimize prevention. The article presented by Aggarwal et al. clarifies the efficacy of the herbal Tribulus terrestris on the inhibition of calcium oxalate calculi formation. Herbal medicine has been long used to treat different health conditions including stone disease. However, only more recently efforts began to be made to determine the mechanisms involved and their objective efficacy. In the present evidence-based medicine era this is of utter importance. Herbal medicines may be an alternative to the currently existing medicines providing the additional advantage of minimal or inexistent collateral effects. Other herbal medicines should undergo evaluations in vitro to amplify the urologist’s clinical armamentarium to combat kidney stones.

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EDITORIAL COMMENT

In this paper, the authors addressed the potential use of Tribulus terrestris as the therapeutic agent to treat urolithiasis. Urolithiasis is characterized by high recurrence rate and among the treatments used are extracorporeal shock wave lithotripsy and drug treatment, although there is no satisfactory drug to use in clinical therapy. Thus the prevention of this disease or its recurrence would be of great interest. Phytotherapy is a common method used in folk medicine as an alternative for primary health care in many countries and particularly the potential effect of many plants to treat urolithiasis has been reported over the past years. The precipitation of calcium oxalate (CaOx) inside the renal tubules and the interaction between CaOx crystals and tubular epithelium plays an important role in the genesis and evolution of urolithiasis, since renal tubular cells selectively bind and uptake CaOx crystals, a phenomenon followed by a series of intracellular events that culminate in a cell damage and death. It was shown that aqueous extract of Tribulus terrestris was able to inhibit CaOx crystallization in vitro and showed cytoprotective properties increasing the cell viability.

The extract of plants with antilithiatic properties (Tribulus terrestris, Phyllanthus niruri, Herniaria hirsute, etc.) has been shown effective to prevent calculi development in the experimental models in vivo and in vitro, showing significant effects on many stages of stone formation including crystallization, aggregation, cellular adherence and adsorption of macromolecules into the calculi, however, its effects in lithiatic patients are much less clear. Many reasons can be raised for this difference such as the treatment onset, number of patients, time of treatment, adhesion to the treatment, etc. Moreover, it was previously shown (1) that rats with already formed vesical calculi, the administration of Phyllanthus niruri had no effect on the calculi size or elimination rate but it induced a shift in the calculi shape toward a smoother surface and probably more fragile form, which could contribute to elimination and/or dissolution of calculi. Overall the available data point to a useful therapeutic application of these plants, including Tribulus terrestris in lithiatic patients, mainly as prophylactic agent in those persons who are at high risk to develop stones since they can potentially interfere with the pathogenesis of urolithiasis and may represent an attractive alternative for the prevention of lithiasis of the urinary tract.

REFERENCE


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