A New Pentacyclic Triterpene Isolated from *Myroxylon balsamum* (syn. *Myroxylon peruiferum*)

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Um novo triterpeno pentacíclico, o 11α-metoxi-β-amirina (1) foi isolado de *Myroxylon balsamum* (L.) Harms (sin. *Myroxylon peruiferum* L.f.). A estrutura desta substância foi elucidada pela análise de seus dados de IV, EM, RMN 1H e 13C. RMN bidimensional foi também utilizada para definir a estrutura e atribuir os deslocamentos químicos dos átomos de hidrogênio e carbono do novo triterpeno.

The new pentacyclic triterpene, 11α-methoxy-β-amyrin (1), was isolated from *Myroxylon balsamum* (L.) Harms (syn. *Myroxylon peruiferum* L.f.). Its structure was established on the basis of IR, MS, 1H NMR, 13C NMR. 2D NMR experiments were also used to establish the structure and the hydrogen and carbon chemical shift assignments of the new triterpene.

**Keywords:** *Myroxylon balsamum* (Myroxylon peruiferum), Fabaceae, 3β-hydroxy-11α-methoxylean-12-ene

**Introduction**

*Myroxylon balsamum* (L.) Harms (syn. *Myroxylon peruiferum* L.f.), family Fabaceae, is a large size tree, with white flowers, winged and aromatic fruits. Its dispersion is wide, being spread from the South of Mexico until the North of Argentina. In Brazil, its distribution includes a vast range of the forest region of the country, being very common in the States of Bahia, Paraná and Mato Grosso. It is known as “cabriuna” and “cabriúna - vermelha”1.

The red-brown wood shows fine stripes, is a little rough and has a peculiar scent due to essential oil. It is a heavy wood and is resistant to deterioration, being widely used in carpentry. The hurtled log supplies a well-known exudate known as balsam of Peru or Tolu or a red oil which is quite rich in vanilla. The oil was used formerly in popular medicine as expectorant for breathing affections and as a sedative in cases of cystitis. Now its use is limited to the perfumery industry and as sedative pills for coughs1-2.

Chemically, the balsam is a mixture of free acids, especially benzoic and cinnamic, and benzyl benzoate. This plant also furnishes a resinous fraction containing monoterpenoids, sesquiterpenoids, alcohols and phenylpropanoids derivatives2. From trunk wood were isolated isoflavones, pterocarpan, coumestans, flavanone, isoflavanones and arylbenzofuran3,4.

In this paper we report the isolation and structural determination of a new pentacyclic triterpene I from the leaves of a specimen of *Myroxylon balsamum* (Fabaceae) collected in Espirito Santo State, Brazil. The structure was established by spectral analysis of I and its acetyl derivative 1a, mainly through 1H and 13C NMR spectra, including homonuclear 1H-1H COSY and heteronuclear 1H detected (inverse method) 1H-13C HMQC<sup>1</sup>CH and 1H-13C HMBC<sup>n</sup>CH (n=2 and 3) 2D shift-correlated spectra5.

**Results and Discussion**

The pentacyclic triterpene I was obtained as colorless crystals whose molecular formula C<sub>31</sub>H<sub>52</sub>O<sub>2</sub> (6 unsaturations) was deduced by comparative analysis of 13C NMR (31 singlet signals) and DEPT-13C NMR data6 (Table 1) [δ = 90° : 6 CH, including one sp<sup>2</sup> (δC 121.51, CH-12) and two sp<sup>3</sup> oxygenated (δC 78.61, CH-3, and 75.85, CH-11); θ = 135° : 9 CH<sub>2</sub> and 9 CH<sub>3</sub> including one methoxy group (δC 53.67, MeO-11)] and LREIMS [m/z 424, 100% (M-MeOH)]. The DEPT results [6 CH, 9 CH<sub>2</sub> and 9 CH<sub>3</sub> = C<sub>2</sub>H<sub>3</sub>11] indicated that one exchangeable hydrogen (hydroxy group) was present, in accordance with the absorption at ν<sub>max</sub> 3380 cm<sup>-1</sup> observed in the IR spectrum and a monoacetyl [δ<sub>H</sub> 2.03 (s)] derivative (1a) obtained by treatment with Ac<sub>2</sub>O in the presence of pyridine (see experimental). The
\(^{13}\)C NMR spectra of I revealed signals at \(\delta_C = 121.51\) (CH-12) and 149.67 (C-13), indicating the presence of a double bond. Because the unsaturation number is 6, I must therefore be a pentacyclic triterpene.

The location of the methoxy group at CH-11 (\(\delta_C = 75.85\) and \(\delta_H = 3.23\)) and not at CH-3 (\(\delta_C = 78.61\) and \(\delta_H = 3.84\)) was defined by heteronuclear long-range couplings between CH-11 (\(\delta_C = 75.85\) and the methoxy hydrogens (\(\delta_H = 3.23\), \(3C(\text{CH})\), CH-12 (\(\delta_C = 121.51\)) and H-11 (\(\delta_H = 3.84, 3C(\text{CH})\) and C-13 (\(\delta_C = 149.67\)) and H-11 (\(\delta_H = 3.84, 3C(\text{CH})\) observed in the HMBC spectrum (Table 1). Other heteronuclear long-range couplings are summarized in Table 1.

The 1H NMR spectrum of I (Table 1) showed nine singlet signals corresponding to one methoxy [\(\delta_H = 3.23\) (MeO-11)] and eight tertiary methyl groups [\(\delta_H = 0.80, 0.89, 0.91, 0.99, 1.00, 1.04, 1.08, 1.21\) (3H-28)] which revealed the signal corresponding to H-3 as a doublet \(J = 8.2\) (axial-axial interaction) and 5.0 Hz at \(\delta_H = 4.49\) [\(\Delta\delta_H = 4.49\) (Ia) – 3.23 (I) = 1.26 ppm].

\(^1\)H and \(^{13}\)C NMR assignments (Table 1) of the pentacyclic triterpene I were determined on the basis of HMBC orientation of the hydroxyl group at CH-3 (H-3 axial) was deduced by comparative analysis of the chemical shifts corresponding to CH-3 (\(\delta_C = 39.26\), CH-2 (\(\delta_C = 27.30\)), CH-3 (\(\delta_C = 78.61\)), CH-5 (\(\delta_C = 55.02\)), CH-24 (\(\delta_C = 15.44\)) of I and the model triterpenoids 2 and 3 supporting a HO-3\(^{\beta}\) (H-3\(\omega\)) and HO-3\(^{\omega}\) (H-3\(\gamma\))/2 (HO-3\(^{\beta}\))/3 (HO-3\(^{\gamma}\)); \(\delta_C = 39.5/33.5\) (CH-1, 2, 24), 78.7/76.0 (CH-3), 39.0/37.0 (C-4), 55.1/48.8 (CH-5) and 15.5/22.4 (CH-24)\(^{7}\), since superimposition of the signals was observed for H-3 (\(\delta_H = 3.23\)) and the methoxyl group (\(\delta_H = 3.23\)) in the \(^1\)H NMR. The shielding revealed by the chemical shifts corresponding to carbon signals of CH-2, 1, CH-5 and CH-24 of 3 can be justified by \(\gamma\)-effects attributed to the hydroxyl group at an axial position (H-3\(^{\beta}\) equatorial). This deduction was confirmed by \(^1\)H NMR and \(^1\)H-\(^1\)H COSY spectra (200 MHz) of the monoacetyl derivative Ia, which revealed the signal corresponding to H-3 as a doublet \(J = 8.2\) (axial-axial interaction) and 5.0 Hz at \(\delta_H = 4.49\) [\(\Delta\delta_H = 4.49\) (Ia) – 3.23 (I) = 1.26 ppm].

Table 1. \(^1\)H and \(^{13}\)C NMR spectral data for I (CDCl\(_3\)).

<table>
<thead>
<tr>
<th>C</th>
<th>(\delta_C)</th>
<th>(\delta_H)</th>
<th>(J_{CH})</th>
<th>HMBC</th>
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<tr>
<td>4</td>
<td>39.00</td>
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<td>8</td>
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<td>10</td>
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<tr>
<td>12</td>
<td>149.67</td>
<td>-</td>
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<td>-</td>
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<td>31.07</td>
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<td>CH</td>
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<td>9</td>
<td>51.56</td>
<td>1.69 ((J = 8.9))</td>
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<td>11</td>
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<td>5.34 ((J = 3.5))</td>
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<td>H-18</td>
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<tr>
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<td>46.76</td>
<td>1.99</td>
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<td>1</td>
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<td>1.91 ((td, J = 13.9, 3.3); 1.23)</td>
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<td>1.04 ((s))</td>
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<td>26</td>
<td>16.76</td>
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<td>0.89 ((s))</td>
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<td>53.67</td>
<td>3.23 ((s))</td>
<td>H-11</td>
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*Chemical shifts (\(\delta_C\) and \(\delta_H\)) and \(^1\)H coupling constants (\(J\) in Hz, in parenthesis) obtained from the one dimensional \(^1\)H and \(^{13}\)C NMR spectra. Multiplicity of signals of carbon atoms deduced by comparative analysis of HBBD- and DEPT-\(^{13}\)C NMR spectra. Homonuclear \(^1\)H-\(^1\)H COSY and heteronuclear \(^1\)H-\(^{13}\)C HMQC \(J_{CH}\) and \(^1\)H-\(^{13}\)C HMBC \(J_{CH}\) (n=2 and 3) 2D NMR spectra were also used for these assignments.
and HMBC data and comparison to the known triterpene 2 [11α-hydroxy-β-amyrin, isolated from callus tissues of Stauntonia hexaphylla (Lardizabalaceae)]8. The 13C NMR spectra of 1 (Table 1), the 11-0-methyl ether of 2, showed signals that matched closely with those of the triterpene 2 (recorded in CDCl3)8, revealing significant differences only in the chemical shifts of the methine CH-11 [δC 75.85 (1) and 81.70 (2)] and the quaternary C-13 [δC 149.67 (1) and 153.20 (2)] carbon atoms. These different chemical shifts may be justified by the presence of a hydrogen bond involving the hydroxy group at CH-11 of 2 and the solvent pyridine-d5, and not in CDCl3 as reported in the literature8. The 13C and 1H chemical shifts of the methine CH-11 [δC 76.6 and δH 3.87 (dd, J = 3.5 and 8.6 Hz)] of camaldulensis acid (3β,30-dihydroxy-11α-methoxyurs-12-en-28-oic acid, 4, recorded in CD3OD), isolated from leaves of Eucalyptus camaldulensis var. obtusa (Myrtaceae)9, and 3 (δC 68.4)7 were also used in this analysis.

Thus, the structure of the new triterpenoid isolated from Myroxylon balsamum was established as 3β-hydroxy-11α-methoxyolean-12-ene (1). The prominent peaks at m/z 255 (60 %) and 271 (16 %) observed in the mass spectrum, which could result from cleavage involving the rings B and D, respectively, in accordance with the formation of a diene after MeOH elimination10, were also used in this structural elucidation. To the best of our knowledge, this triterpene 1 is hitherto unreported.

### Experimental

#### General Experiments Procedures

EIMS were recorded on a VG Platform II mass spectrometer. 1H (400 MHz), 13C (100 MHz), 1H-13C HMBC and 1H-13C HMBC nJCH spectra were recorded using a Bruker ARX-400 spectrometer, in CDCl3 as solvent; residual CHCl3 (δH 7.24) and the central peak of the triplet of CDCl3 (δC 77.00) were used as internal references. The multiplicities of the carbon signals were deduced by comparative analysis of the HBBD- and DEPT-13 CNMR spectra. Heteronuclear 1H and 13C connectivities were deduced by 1H-13C HMOC nJCH [spin-spin coupling of carbon and hydrogen via one bond (nJCH 145.0 Hz)] and 1H-13C HMBC nJCH [n=2 and 3, spin-spin interaction of carbon and hydrogen via two (2JCH) and three (3JCH) bonds, optimized for m/z of 9 Hz]. IR spectra with KBr plates were obtained on a FTIR Perkin-Elmer 1600/1605 spectrometer.

#### Plant material

The leaves of Myroxylon balsamum were collected at Reserva Florestal de Linhares, Companhia Vale do Rio Doce (CVRD), Espirito Santo State, Brazil, during September 1996. A voucher specimen has been deposited in the CVRD Herbarium (voucher no CVRD-483).

#### Extraction and Isolation

The air dried and powdered leaves (328.0 g) of Myroxylon balsamum were successively extracted with hexane, EtOAc and MeOH at room temperature and the solvents removed under vacuum to yield 36.1 g (rich in waxes and carotenoids), 17.4 g and 5.25 g of residues, respectively. The residue (17.4) obtained from the EtOAc solution was chromatographed on a of silica gel column eluting with hexane/CH2Cl2 mixtures of increasing polarity; a total of 45 fractions (ca. 100 mL each one) were collected and combined of the basis of TLC comparison. The fractions 18-22, eluted with hexane-CH2Cl2 (1:1), furnished 1 (92.0 mg) after recrystallization from CHCl3.

3β-Hydroxy-11α-methoxyolean-12-ene (1)

Colorless crystals, mp 172-173°C, [α]D + 12.4° (c 0.73, CH2Cl2); IR (KBr): 3380, 1461, 1384, 1044, 815, 731 cm-1; EIMS m/z (rel. int.) 456 (M+· abs.), 425 (25, M-MeO), 424

Other 11-hydroxy or 11-methoxy pentacyclic triterpenes described in the literature are 28-oic acids: 2α,3β,23-trihydroxy-11α-methoxyurs-12-en-28-oic acid [isolated from resin of Shorea robusta (Dipterocarpaceae)]11, 11α-hydroxy-betulinic acid [isolated from leaves of Licania pyrifolia (Cryosbalanaceae)]12 and 11α-hydroxytormentic acid [2α,3β,11α,19α-tetrahydroyurs-12-en-28-oic acid, isolated from aerial parts of Rosa laevigata (Rosaceae)]13.
A sample of 1 (18.0 mg) was treated with Ac₂O (2.0 mL) and dry pyridine (1.0 mL) at room temperature overnight. After the usual work-up, the crude product was chromatographed on a silica gel column eluting with CHCl₃ to furnish the acetyl derivative 1a (16.0 mg) as colorless crystals, mp 149-151°C, [α]D +5.7° (c 0.965, CH₂Cl₂); EIMS (rel. int.) 498 (M +. abs.), 466 (100, M-MeOH), 451 (8, M – MeOH - Me₂), 391 (14, M – MeOH - Me – AcOH), 255 (42), 253 (10); ¹H NMR (200 MHz, CDCl₃) δH 5.30 (br s, H-12), 4.49 (dd, J = 8.2 and 5.0 Hz), 3.88 (dd, J = 8.8 and 3.6 Hz), 3.18 (s, MeO-11), 2.03 (s, AcO-3), 1.23 (s, 3H-23), 1.18 (s, 3H-14), 1.04 (s, 3H-25), 0.98 (s, 3H-26), 0.86 (s, 3H-27, 3H-28, 3H-29), 0.81 (s, 3H-30).

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