Mapping Ethanol Production Sources in Brazil Through Stable Isotopes

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Ethanol is a biofuel produced in Brazil through fermentation of sugarcane, requiring vast plantation areas and water availability. The present work aimed at testing isotopic markers as tools for ethanol source appointment or certification of origin. For this, oxygen and hydrogen isotopic patterns were determined in plant-water, soil-water, rainwater, and water from reservoirs and some rivers in four sugarcane crop areas. The isotopic fingerprint of carbon and hydrogen in ethanol produced in the respective mills was also determined. Samples were collected in 2011 and 2012 in crop areas of the state of Amazonas (North), Mato Grosso (Center-West), São Paulo (Southeast) and Rio Grande do Sul (South). The substantial and complex influence of the hydrological cycle on the ethanol δD and the small δ^{13} C variations constrain the use of isotopes for the intended objectives.

Keywords: continuous flow isotope ratio mass spectrometry (CF-IRMS), isotope ratio infrared spectroscopy (IRIS), ethanol, water, isotopic ratios

Introduction

Ethanol is an important biofuel in the scenario of renewable and sustainable sources of energy. In Brazil it is produced via fermentation of sugarcane, a plant of the botanical group C4, (family Gramineae and genus *Saccharum*), which has a high economic importance since it also yields sugar. Native to tropical climates, this plant grows under different and even adverse environmental conditions due to the development of several improved varieties as to give the best response and allow extended harvest over most part of the year.¹ The typical sugarcane biomass structure is composed of water, fiber and sugar.²

Sugarcane crops require large amounts of water, which represents 70% of its weight, mostly absorbed through the roots.³ Soil-water is closely connected to the plant-water and also to rainwater. Rainwater infiltrates into soil subsurface where it is retained to rebuild the moisture and recharge the phreatic. Ethanol is formed during fermentation of sugars, and in this process both sugar and water medium

contribute as sources of hydrogen,^{4,5} while plant-water and sugar are the sources of oxygen.⁶ The hydrogen atoms in the ethanol methylene group derive from water present in the fermentative medium, composed basically of plant-water, so that the differentiation of ethanol sources involves the understanding of water cycle and its interactions in the ecosystem. In terms of carbon isotope ratios, ethanol should reflect the isotopic composition of original sugar since fermentation is not a fractionating reaction.⁷

¹⁸O/¹⁶O and ²H/¹H of water are important tracers in hydrogeological studies, since phase changes, like evaporation and condensation (precipitation), determines the enrichment or depletion of the heavier isotope in a water reservoir. Although gradients in the isotopic composition of soil-water may arise from differences in seasonal moisture inputs, evaporation in the uppermost surface layer as well as from differences between bulk soil moisture and groundwater, there is no isotopic fractionation during water uptake by roots and transportation to leaves.⁸⁻¹⁰ The isotopic evaluation provides, therefore, information about the water source (irrigation, rainwater, groundwater, and others) and usage by plants. Oxygen, hydrogen and also carbon isotopic

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composition of ethanol has been used for: differentiation of botanical and geographical origins;^{4,11,12} evaluation of isotopic fingerprinting in contaminated sites;¹³ assessment of ethanol sources in the atmosphere¹⁴ and investigation of alcoholic beverages adulteration.^{15,16} Despite the widespread ethanol production in Brazil, in sites of diverse climatic properties, no evaluation has been reported of regional conditions on the isotopic fingerprinting of ethanol.

Brazil is one of the most important ethanol producers^{17,18} with the mean annual production of 21.3 billion m³ (for the 2003-2012 period)¹⁹ carried out in at least 378 registered active sugarcane mills^{20,21} and has the potential to be the most important exporter of this commodity. Because the expansion of agricultural frontier may represent a threat to forested ecosystems and protected areas,²² it is pertinent to identify tools that can be used to detect the provenance of ethanol and to certify its origin.

The specific objectives of this work were: (*i*) to assess oxygen and hydrogen isotopic patterns in plant-water, soil-water, rainwater, and water from reservoirs and rivers associated to four Brazilian sugarcane crop areas; (*ii*) to determine carbon and hydrogen isotopic fingerprint of ethanol produced in the same areas as to evaluate regional isotopic patterns, seasonal variations, and the influence of hydrological cycle/climatic conditions on ethanol isotopic ratios. Relationship between carbon isotope ratio of ethanol and sugarcane biomass was also investigated. The general aim of the work was testing tools for identification of ethanol geographical origin and source appointment.

Experimental

Study area

All samples in this study were collected in sugarcane mills and nearby not irrigated crops distributed in four different locations covering distinct geographical regions of Brazil (Figure 1): Amazonas (AM, North), Mato Grosso (MT, Center-West), São Paulo (SPA, Southeast) and Rio Grande do Sul (RS, South). Reference spots for rainwater isotopic pattern and precipitation amount were selected among the nearest stations of the Global Network for Isotopes in Precipitation (GNIP) database²³ and the National Institute of Meteorology (INMET) database,²⁴ respectively.

Sampling campaigns in AM were performed in Agropecuária Jayoro mill (1° 59' 17.85'' S 60° 8' 27.39'' W, altitude 144 m), in Presidente Figueiredo municipality. The average annual temperature in this location is 25 °C and the annual average rainfall of 2,500 mm.^{25,26} Presidente Figueiredo lies on the basin of Uatumã River, in the limits of the hydrographic sub-regions of Negro and Trombetas rivers, which belong to the hydrographic basin of the Amazon River.²⁷ Soil type is dystrophic yellow latosol.²⁶

Sampling campaigns in MT were performed in Coprodia mill (13° 47' 17.33" S 57° 50' 31.31" W, altitude 528 m), in Campo Novo do Parecis municipality. Average temperature ranges between 24 and 40 °C in this location, where two climatic types predominate: Equatorial hot and humid, and tropical hot and sub-humid. The first climate



Figure 1. Map showing sampling locations; typical rain amount and isotopic pattern in reference stations nearest to sampling sites. Lower plot: monthly accumulated precipitation (1961-1990) in the INMET stations. Upper plot: rainwater isotopic ratios (1965-1987; SP only in 1996-1998) in the GNIP stations.

type includes three dry months, from June to August, and intense rainfall from January to March dominated by the Equatorial continental air mass; the second climate type includes four dry months, from June to September, and a rainfall period from December to February, dominated by continental tropical air mass.²⁷ The mean annual precipitation is 1,796 mm.²⁸ Water provision for this mill derives from an affluent of Do Sangue River in the Juruena-Arinos River basin, located in the Tapajós River hydrographic sub-region, which, in turn, fits in the Amazon River hydrographic region.^{27,29} Three soil types are found in this location: quartz sand, red and dark red latosols.²⁷

Sampling campaigns in SPA were performed in Pau D'Alho mill (22° 46' 4" S 50° 6' 43" W, altitude 458 m), located in Ibirarema municipality. The climate is tropical with annual isothermal temperature in the range of 20 to 22 °C and mean annual precipitation of 1,511 mm; the rainy season occurs from November to February. In general, weather conditions in this hydrographic region depends on a series of factors, such as tropical Atlantic mass control, invasion of cold fronts, incursions of the continental tropical air mass associated with Chaco Low (typical low pressure system), presence of south Atlantic convergence zone (SACZ) and disturbances caused by the relief. This place lies in the hydrographic region of Paraná River, in Paranapanema sub-basin. Structured purple ground soil predominates in this location.^{30,31}

Sampling campaigns in RS were performed in Coopercana mill (27° 54' 18.35" S 55° 9' 37.92" W, altitude 125 m), located in Porto Xavier municipality, near the Argentinean border, and only 2 km away from Uruguay River. In this location climate is temperate, average annual temperatures ranges from 16 to 20 °C increasing from May to September; intra-annual rainfall distribution is regular with an annual average of 1,784 mm. Atmospheric circulation is governed by tropical and polar air mass systems, with tropical Atlantic (Ta) and polar Atlantic (Pa) predominating alternately in all seasons.^{31,32} This location lies in the so-called medium Uruguay area, in Ijuí River hydrographic sub-region which is part of the Uruguay River hydrographic region.³² Dystrophic red latosol soil predominates in this location.

GNIP reference stations for rainwater isotopic ratios located in Manaus (AM_{GNIP}), Cuiabá (MT_{GNIP}) and Porto Alegre (RS_{GNIP}) has data survey for the period from 1965 to 1987. Because results for São Paulo State are scarce, existing GNIP data for the period 1996 to 1998²³ obtained in Santa Maria da Serra, Campinas, Bragança and Piracicaba stations (SPA_{GNIP}) were used. Historical data for mean monthly and annual accumulated precipitation were available for the INMET stations located in Manaus (AM_{INMET}) , Cuiabá (MT_{INMET}) , Porto Alegre (MT_{INMET}) , Santa Rita and Campinas (SPA_{INMET}) in the period from 1961 to 1990.²⁴

Sampling

Sampling campaigns carried out in August and November, 2011, included ethanol, sugarcane and rainwater samples (pilot study), and those in July and October-November, 2012 included ethanol, sugarcane, rainwater, surface waters and soil samples. Additional surface and groundwater were sampled in MT on September, 2014 for confirmation of groundwater isotopic profile. Ethanol and water samples were stored refrigerated at -4 °C while plant and soil samples were kept frozen at -18 °C.

Ethanol samples were collected from the reservoirs of the respective mill in clear glass flasks fitted with screw caps faced with polytetrafluoroethylene (PTFE), and protected from light with aluminum foil.

Rainwater samples were collected during individual rain events, a couple of days before sampling of the other matrices, using glass bottles fitted with funnel or large aluminum recipients, then transferred to clear glass flasks fitted with screw caps faced with PTFE. The only exception was the sample collected in MT, which integrated all precipitations throughout October, 2012. Surface and groundwater were collected from open water systems and artesian wells, respectively, directly into clear glass flasks fitted with screw caps faced with PTFE.

Plant samples were collected from the crop, in the neighborhood of the mills, by cutting a transverse section at half height of the plant. In 2011, sampling involved the collection of a single plant in each spot, which was involved in polyethylene film and stored in a polyethylene flask sealed with PTFE film, while in 2012 plants were collected from every 10 m along the perimeter of a square of 20×20 m, plus one in the center, amounting to 9 sub samples that were gathered as a final composite sample and kept in an aluminum flask sealed with PTFE film.

Soil samples were collected only in 2012, from 0-5 cm and 20-25 cm depths, near the plant root system, in the center of the referred square described for plant samples, and kept in an aluminum flask sealed with PTFE film.

Analysis

Analyses of water were carried out through the technique of isotope ratio infrared spectroscopy (IRIS) using a water analyzer model DLT-100, based on off-axis integrated cavity output spectroscopy (OA-ICOS; Los Gatos Research, Mountain View, CA, USA). All water

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samples were filtered through 0.45 µm filters and injected 6 times, discharging the 3 first results to avoid memory effect. Plant- and soil-water were previously extracted by vacuum distillation in a specially designed apparatus that allowed the processing of six-sample batches twice a day. Water samples extracted from plants were also treated over activated charcoal for several days to eliminate any co-extracted organic contaminant. Analytical precisions, expressed as standard deviations, were better than $\pm 0.60\%$ for δ^{18} O and $\pm 2.80\%$ for δ D for water and soil-water results, while for plant-water they were better than $\pm 0.74\%$ for δ^{18} O and $\pm 3.60\%$ for δ D.

An inter-laboratory comparison was performed for the plant-water samples using an analyzer model L2130-I, based on wavelength-scanned cavity ring-down spectroscopy, fitted with a vaporizer model A0211 and a micro-combustion module (WS-CRDS; Picarro Inc., Sunnyvale, CA, USA). The procedures are detailed by Godoy *et al.*³³ and analytical precisions were better than $\pm 0.25\%$ for δ^{18} O and $\pm 0.60\%$ for δ D.

Determinations of δ^{13} C and δ D in ethanol were performed via continuous flow isotope ratio mass spectrometer (CF-IRMS). Before analysis, all samples were dehydrated using molecular sieve UOP type 3A (Sigma-Aldrich, Fluka, St Louis, USA) according to Monsallier-Bitea *et al.*¹⁵ and Jamin *et al.*³⁴ The molecular sieve was previously activated at 300 °C for 4 h, added in small amounts to the vials containing samples and then kept refrigerated overnight. Dehydrated samples were diluted to 1 mmol L⁻¹ in isooctane and either injected in 1:100 split for carbon analyses or without dilution in splitless mode for hydrogen analyses. Pro analysis grade ethanol (VETEC, Duque de Caxias, RJ, Brazil) was used as working standard to evaluate the performance of the analytical system at every 20 injections. Analytical precisions were better than ± 0.15‰ for δ^{13} C and ± 4.8‰ for δ D.

Analytical system was composed of a CF-IRMS Deltaplus V (ThermoFinnigan, Bremen, Germany) with open split, connected in-line via Conflo 4 system to a gas chromatograph Trace GC Ultra interfaced by an Isolink combustion oven (Thermo Electron S.p.A, Milan, Italy). GC was equipped with a capillary column HP DB-624 $(30 \text{ m} \times 0.45 \text{ mm i.d.}, 2.55 \text{ }\mu\text{m film})$ and the analysis conditions were as follows: injector temperature, 250 °C; split injection mode with ratio 100:1 (for carbon analysis) or splitless (for hydrogen analysis); split flow at 150 mL min⁻¹; carrier gas in constant flow mode, 1.5 mL min⁻¹; oven temperature program: 45 °C for 5 min, raising from 45 °C to 190 °C at 35 °C min-1 and 190 °C for 1 min. Injection volumes were 2 µL. Isolink ovens were set at 1000 °C and 1400 °C for carbon and hydrogen isotope ratios, respectively.

Confirmation of isotopic ratios for the ethanol working standard as well as the determinations of δ^{13} C in sugarcane samples were performed in an elemental analyzer Flash EA 1112 (ThermoQuest, Milan, Italy) equipped with combustion and thermal conversion reactors and coupled to the IRMS (EA-IRMS). The combustion reactor (conversion to CO₂ at 1020 °C) was prepared with quartz tube filled with chromium oxide, 50 mm; reduced copper, 110 mm; silvered cobaltous/ cobaltic oxide, 30 mm, and 1 mm quartz wool layer on top, bottom and between layers. The thermal conversion reactor (conversion into H₂ at 1450 °C) was prepared with a ceramic tube containing a glassy carbon granulate, 100 mm; silver wool, 5 mm; glassy carbon granulate, 15 mm, quartz wool, 15 mm, and silver wool, 5 mm.

Sugarcane samples were freeze-dried and ground in an analytical mill A11 Basic (IKA Works Inc., North Carolina, USA) before analyses. Ethanol standard was introduced in the analytical system through the autosampler for solids, using adsorbents to avoid evaporation and consequent fractionation before conversion in the reactor. For δ^{13} C analysis ethanol was previously supported over Chromosorb W, as described by Adami *et al.*,¹¹ while for δ D analyses ethanol was supported over activated charcoal previously treated by heating at 170 °C for 4 h. Activated charcoal was chosen because it does not contain hydrogen atoms and is a cheap and readily available adsorbent. No previous citation was found in the consulted literature about the application of active charcoal for isotope analysis purposes.

Stable isotope ratios are expressed in δ notation (%) and were calculated according to the following equation:

$$\delta = \left(\frac{R_{\text{sample}}}{R_{\text{standard}}} - 1\right) \times 1000$$

where R_{sample} and $R_{standard}$ represent the isotope ratios (¹³C/¹²C and D/H) of samples and standard material, respectively. The reference materials used for calibration were: USGS 40, L-glutamic acid ($\delta^{13}C_{VPDB-LSVEC} = -26.39 \pm 0.04\%$) and IAEA-CH-7, polyethylene ($\delta D_{VSMOW} = -100.3 \pm 2.0\%$), and Vienna standard mean ocean water (VSMOW).

All isotopic ratios are reported relative to VSMOW and VPDB.

Evaluation of ethanol results used of the Statistica software, version 12.0 from Stat Soft Inc. Outliers values for each distilleries were removed based on Chauvenet, Dixon and Grubbs statistical tests. The remaining data set were used to run analysis of variance (ANOVA), which involved tests of homogeneity of variances based on Cochran, Hartley, and Bartlett. Normal distribution of data was confirmed by evaluation of residuals of the statistical model using Shapiro-Wilk and Kolmogorov Smirnov tests.

Results and Discussion

Sampling campaigns were carried out during the months of August and November, 2011 and July and October/ November, 2012. The 2011 campaign was a pilot study comprising only sugarcane, rainwater and ethanol samples. Obtained data revealed a complex relation between these matrices, so that new sampling campaigns were planned and carried out in 2012 including surface- and soil-water, aiming at a better understanding. Sampled sugarcane varieties were the most representative in the production of each mill. Since in Brazil there is a continuous overturn of varieties cultivated along the year, it was not possible to keep a control area or collect a specific variety along subsequent sampling campaigns.

Results of δ^{18} O and δ D for rain and surface water obtained in the present work are presented in Table 1, followed by those for plant- and soil-water in Table 2. Results of δ D and δ^{13} C for ethanol and sugarcane are presented in Table 3.

The statistical tests with ethanol results indicated data normal distribution therefore variance analysis based on ANOVA was applied.

Interlaboratorial comparison of plant-water results

Plant-water analysis by IRIS can be biased due to the presence of residual methanol or ethanol after distillation procedure,^{35,36} resulting in discrepancies in $\delta^{18}O$ and δD values of up to 7.97% and 13.70% for water extracted from stem samples.35 This problem can be corrected by use of a post-processing software in OA-ICOS equipments (not applied in this work), as long as the expected amount of these contaminants is known, what requires comparison of results with those from CF-IRMS or another technique to eliminate the interference. All water samples in this study were analyzed in the Los Gatos DLT-100 equipment, and to check for possible bias in the obtained results, these samples were re-analyzed in a Picarro L2130-I equipment doted of a micro-combustion module, thereby eliminating any organic interference. Isotope differences between mean results varied between 0.58 to 2.25% (median 0.20%) for oxygen and 0.25 to 7.25% (median 2.57%) for hydrogen. Statistical comparison of their variances (F test) and means (*t*-test), showed that respectively 83% and 78% of the δ^{18} O and δD results were not statistically different at significance levels of 0.05 to 0.02 (Tables S1 and S2 in Supplementary Information (SI) section).

Rainwater, surface and groudwater

Results for both δ^{18} O and δ D in rain and surface waters were found in a broad range, although precipitation showed the largest variability (Table 1). As mentioned in the Study area section, hydrological patterns for these locations were estimated by comparison with historical data from GNIP stations and INMET stations (Figure 1).

Rain amount has important influence upon isotopic ratios of rainwater due to the so-called amount effect, resulting in enriched rainwater during the dry season,^{10,37} as shown in Figure 1, for all reference stations. In AM, the largest rain amount occurs by the beginning of the year; in MT an important drought follows in the middle of the year similarly, but attenuated, in respect to SPA; and RS has a regular distribution of rain along the entire year. In MT, virtually no rain occurs in the crop area during all the harvest period from May to October, and the influence of this climatic condition is revealed in the plant-water and soil-water isotopic ratios, as it will be discussed later. Isotopic fluctuations in the rainfall in tropical regions seem to be governed by the rain-out process in the surrounding region instead of depending on the rain-out history,³⁸ so that the comparison with GNIP and INMET data can be considered a good approach for isotopic amount effects in this study.

Except for MT, surface water results were close to δ^{18} O and δ D mean values of -4.54% and -27.65% for 2011-2012 samplings (Table 1). These values are in good agreement with the weighted average of δ^{18} O and δ D for annual rainfall in the GNIP reference stations (Table 1), which should represent groundwater isotopic fingerprint in these hydrographic basins.^{10,39} The differences observed for MT samples may be attributed to the severe drought which occurs regularly from May to October in that location. Comparison of river and groundwater MT results with the weighted average for $\delta^{\scriptscriptstyle 18}O$ and δD in the reference MT_{GNIP} during the rainy season (Table 1) revealed isotopic depletion in this location resulting from the recharge period in the catchment basin. This evaluation strongly indicates that aquifers and water reservoirs observed in this work reproduce groundwater isotopic fingerprint due to phreatic recharge by rainwater. The two vinasse samples, by-products from sugarcane milling process collected from dikes in MT and SPA, seem to be highly enriched in relation to fresh surface water, probably due to evaporation.

Craig⁴⁰ reported a global relationship between δ^{18} O and δD in precipitation known as the global meteoric water line (GMWL), which can be expressed by the regression $\delta D = 8\delta^{18}$ O + 10 and provides important information about a certain site, since isotopic shifts are basically a

Sample	Campaign	Source	δ ¹⁸ O / ‰	δD / ‰
AM rain 1	Aug/2011	Rainwater	-6.4	-29.2
AM rain 2	Nov/2011	Rainwater	-4.6	-22.7
AM rain 3	Oct/2012	Rainwater	-1.02	1.17
AM rain 4	Oct/2012	Rainwater	-1.49	0.89
AM rain 5	Nov/2012	Rainwater	-0.03	5.29
AM water 1	Oct/2012	Igarapé (stream)	-4.20	-23.41
AM water 2	Oct/2012	Igapó (stream)	-4.67	-27.68
AM water 3	Nov/2012	Igarapé (stream)	-5.26	-29.95
MT rain 1	Oct/2012	Rainwater	-1.60	-1.80
MT water 1	Jul/2012	Do Sangue River (reservoir)	-6.53	-43.36
MT water 2	Jul/2012	Vinasse effluent	-4.56	-26.74
MT water 3	Sep/2014	Do Sangue River (reservoir)	-7.44	-42.64
MT water 4	Sep/2014	Do Sangue River	-7.95	-42.96
MT water 5	Sep/2014	Verde River	-7.62	-44.15
MT water 6	Sep/2014	Artesian well 1	-7.42	-43.05
MT water 7	Sep/2014	Artesian well 2	-7.39	-42.74
SPA rain 1	Nov/2011	Rainwater	-3.5	-16.5
SPA water 1	Ago/2012	Bom Retiro Lagoon	-4.12	-29.01
SPA water 2	Ago/2012	Process water	-4.20	-27.91
SPA water 3	Ago/2012	Vinasse effluent	-2.55	-13.6
RS rain 1	Ago/2011	Rainwater	-2.9	0.3
RS rain 2	Nov/2011	Rainwater	-9.6	-65.9
RS rain 3	Jul/2012	Rainwater	-1.13	7.97
RS rain 4	Oct/2012	Rainwater	-2.73	-7.52
RS rain 5	Oct/2012	Rainwater	-2.74	-7.89
RS water 1	Jul/2012	Uruguay River	-4.90	-27.95
RS water 2	Oct/2012	Uruguay River	-4.38	-26.85
	Surface water mean value	-4.54	-27.65	
AM_{GNIP}^{a}	1965-87	Rain, Manaus	-5.15	-27.68
MT _{GNIP} ^a	1965-87	Rain, Cuiabá	-5.40	-34.56
MT _{GNIP} ^b	1965-87 (Nov-Apr)	Rain, Cuiabá	-6.15	-40.65
SPA _{GNIP} ^a	1996-98	Rain, São Paulo	-5.15	-27.95
RS _{GNIP} ^a	1965-87	Rain, Porto Alegre	-4.83	-25.84

Table 1. Isotopic ratios for surface water and GNIP reference stations

^aWeighted average for annual rainfall isotopic ratios calculated from GNIP database, representing the isotopic fingerprint of groundwater;^{10,39} ^bweighted average of rainfall isotopic ratios between November and April (rainy season), calculated from GNIP database.²³

function of local temperature, altitude and distance from ocean. Reference local meteoric water lines (LMWL_{GNIP}) were drawn using GNIP rainwater data and compared with respective LMWL obtained from water samples collected in the present work (Figure 2). In these plots, slope values below that of LMWL_{GNIP} indicate enrichment, typical for water from reservoirs, e.g., lakes, which are more likely to evaporate, while slopes above LMWL_{GNIP} should represent a depletion, which may have several causes, such as the altitude and continental effects.¹⁰ AM and MT showed LMWL slopes similar to each other and both smaller than the respective LMWL_{GNIP} station. The smaller slope in LMWL for AM samples in relation to the respective LMWL_{GNIP} may be due to the contribution of isotopically enriched water vapor released by evapotranspiration from

the Amazon forest, while the steaper line in LMWL for RS samples must be related to the orographic complexity and latitudinal positioning⁴¹ of that site. Due to the reduced number of samples no reliable LMWL could be drawn for SPA location.

Climatic events capable of causing alterations in the isotope ratios in this study, such as the El Niño and La Niña, were considered. These events are disruptions of the ocean-atmosphere system in the tropical Pacific characterized by unusually warm or unusually cold ocean temperatures, respectively, in the Equatorial Pacific, and which have important consequences for weather around the globe including changes in rain amount in Brazil, which could affect isotopic patterns of rainfall influencing results obtained in this work.⁴² Evaluation of oxygen and hydrogen



Figure 2. Local water lines and comparison with GNIP rainwater data.

isotopic ratios from reference GNIP stations showed that rain water becomes usually isotopicaly enriched for both elements during El Niño episodes in comparison to regular periods, while no strong tendency is observed during La Niña events (Figure S1 in the SI section). The GNIP data shows that La Niña has little or no impact on the isotopic fingerprint. The November 2011 sampling campaign carried out during such event may then be evaluated under the same conditions as those used for the other campaigns occurring in normal periods.

Plant-water and soil-water

In general, isotope ratios for water extracted from soil and plant samples showed considereable spread (Table 2). Plant-water was in the range of -9.98 to -1.54% for δ^{18} O and -82.15 to -8.81% for δ D, while soil-water showed δ^{18} O in the range of -12.09 to 3.89% and for δ D from -83.71to 1.78%, all relative to VSMOW. Despite the variability of plant-water results, the proximity with the weighted average for rainfall (Table 1), representing the isotope fingerprinting of groundwater, reflects the connection among this compartment, soil- and plant-water, since soil water, which recharges groundwater and is absorbed by plant roots, is fed by rainwater. In general, the most depleted plant-water both in ¹⁸O and D were found in MT samples, while RS samples showed the least variation. Having in mind the considerable natural variability of the studied system, the results point to a reasonable parity between soilwater (20-25 cm) and the respective plant-water average isotopic ratios, while this does not occur for soil-water from the top layer (0-5 cm). Differences between soil- and plant-water in both cases may derive from: (i) fractionation due to evaporation and/or mixing of waters from the vadose zone with water from recent rain events; (ii) water uptake by sugarcane from deeper soil layers then better reflecting the groundwater isotopic fingerprint. Studies^{9,10,43} on the isotopic fractioning of water at different soil depths (Figure 3) show kinetic isotopic effects increasing as soil moisture decreases, so that strong fractionation may be expected for samples near surface, while there is a tendency for water in vadose zone to be isotopically similar to that from the phreatic. Indeed the 0-5 cm soil-water samples gave results dramatically enriched in respect to soil samples collected at 20-25 cm depth. Despite most of sugarcane root system being found close to surface (about 63% on the top 30 cm), it can eventually reach a maximum depth of 1.5 to 2 m (Figure 3).44

Amazonas

Plant-water results for August and November, 2011 campaigns in AM ($\delta^{18}O = -5.35\%_0$, $\delta D = -35.58\%_0$ and $\delta^{18}O = -3.82\%_0$, $\delta D = -24.07\%_0$, respectively; see Table 2) were consistent with those from local rain samples $(\delta^{18}O = -6.4\%, \delta D = -29.2\%$ and $\delta^{18}O = -4.64\%$, $\delta D = -22.7\%$; see Table 1) and also with the isotopic fingerprint of groundwater, represented by the weighted average for annual rainfall ($\delta^{18}O = -5.15\%$, $\delta D = -27.68\%$, AM_{GNIP}; see Table 1). Contrasting with these data, plant-water for October-November, 2012 campaign was more enriched (δ^{18} O and δ D mean values: -2.47% and -16.10%, respectively) with isotopic ratios between those for rainwater collected in the same period (δ^{18} O and δ D mean values: -0.85% and 2.45%, respectively, n = 3; see Table 1) and groundwater (δ^{18} O and δ D mean values: -4.71% and -27.01%, respectively, n = 3; see Table 1). These data indicate that sugarcane crop absorbed both rain and groundwater prior to this sampling campaign, resulting in isotopic fingerprint from mixed sources. For soil-water (2012), there are two subsets of isotopic patterns in the

Table 2. Results for $\delta^{\scriptscriptstyle 18}O$ and δD in plant-water and soil-water

Sampling information		δ18Ο / %ο			δD / ‰					
		Soil-water Plant-water		Soil-water Plant-water			-water			
Location	Sugarcane variety	Campaign	0-5 cm	20-25 cm	OA-ICOS	WS-CRDS	0-5 cm	20-25 cm	OA-ICOS	WS-CRDS
AM	Unknown	Aug/2011	_	_	-5.35	_	-	-	-35.58	_
	Unknown	Nov/2011	-	-	-3.82	-	-	-	-24.07	-
	SP 813250	Oct/2012	0.54	-1.88	-3.33	-3.04	-6.76	-13.97	-22.03	-18.69
	SP 791011	Oct/2012	-1.99	-4.33	-2.64	-2.36	-9.46	-26.67	-17.53	-14.45
	RB 835486	Oct/2012	-2.39	-4.10	-1.90	-1.98	-22.45	-32.26	-14.21	-12.79
	SP 813250	Nov/2012	-0.14	-1.61	-2.32	_	-1.38	-10.33	-12.38	_
	RB 835486	Nov/2012	0.15	-1.05	-2.16	-1.96	1.48	-6.74	-14.34	-12.78
		Mean Nov/12	-0.77	-2.60	-2.47	-2.33	-7.71	-17.99	-16.10	-14.68
	Unknown	Aug/2011	_	_	-9.20	_	_	_	-68.94	_
	Unknown	Aug/2011	-	-	-9.53	-	-	-	-82.15	-
	Unknown	Aug/2011	_	-	-7.66	_	_	_	-58.93	_
	RB 867515	Aug/2011	-	-	-9.98	-	-	-	-77.04	_
		Mean Aug/11	-	_	-9.09	_	-	_	-71.76	_
	RB 867515	Jul/2012	_	_	-8.47	-7.70	_	_	-55.70	-51.62
	RB 867515	Jul/2012	2.19	-3.92	-5.62	-5.48	-5.32	-22.58	-48.78	-45.85
	RB 928064	Jul/2012	0.73	-4.59	-8.66	-8.35	-6.65	-28.15	-66.63	-65.13
) (T	RB 928064	Jul/2012	-0.98	-6.80	-6.81	-6.95	-11.95	-48.20	-55.20	-52.97
MT	RB 835486	Jul/2012	2.72	-8.64	-8.62	-7.83	-28.14	-67.99	-67.42	-64.57
	RB 92579	Jul/2012	_	_	-8.05	-7.71	_	_	-61.60	-59.50
		Mean Jul/12	1.17	-5.99	-7.71	-7.34	-13.01	-41.73	-59.22	-56.60
	RB 928064	Oct/2012	-2.12	-6.18	-1.54	-1.52	-25.49	-51.53	-20.32	-19.19
	SP 791011	Oct/2012	-3.07	-6.81	-7.39	-7.54	-33.05	-54.91	-61.56	-59.11
	RB 867515	Oct/2012	1.72	-0.20	-1.65	-1.45	-0.50	-2.91	-12.27	-9.14
	RB 867515	Oct/2012	3.89	0.94	-7.14	-4.89	1.78	-13.57	-52.67	-45.42
	RB 835486	Oct/2012	-2.87	-5.36	-3.03	_	-23.62	-33.93	-25.59	_
		Mean Oct/12	-0.49	-3.52	-4.15	-3.85	-16.18	-31.37	-34.48	-33.21
SPA	Unknown	Aug/2011	-	_	-6.23	_	-	-	-33.37	_
	Unknown	Nov/2011	_	_	-6.73	_	_	-	-30.93	-
	SP 813250	Aug/2012	-2.10	-6.69	-2.45	-3.03	-26.12	-44.00	-19.95	-19.70
	RB 867515	Aug/2012	0.02	-3.89	-3.97	-4.11	-16.46	-31.65	-27.53	-24.99
	RB 855156	Aug/2012	-1.63	-6.37	-2.48	-1.06	-18.40	-43.96	-22.98	-16.90
		Mean Aug/12	-1.24	-5.65	-2.96	-2.73	-20.33	-39.87	-23.49	-20.53
RS	RB 855156	Aug/2011	-	-	-4.87	-	-	-	-31.11	-
	RB 855156	Nov/2011	-	-	-4.67	-	-	-	-32.51	-
	RB 855156	Jul/2012	-2.01	-5.75	-4.11	-2.84	-13.85	-47.90	-29.52	-22.97
	RB 867515	Jul/2012	-1.56	-3.98	-3.19	-3.54	-6.41	-27.97	-21.66	-18.39
	SP 813250	Jul/2012	-1.09	-4.74	-3.43	-3.75	-1.53	-37.18	-22.08	-19.48
		Mean Jul/12	-1.55	-4.83	-3.58	-3.38	-7.26	-37.68	-24.42	-20.28
	SP 813250	Oct/2012	-9.54	-9.48	-2.17	-2.26	-81.91	-72.98	-8.81	-7.94
	RB 855156	Oct/2012	-12.09	-10.17	-4.73	-4.82	-83.71	-76.35	-31.89	-30.44
	RB 867515	Oct/2012	-9.58	-9.10	-6.65	-6.33	-78.47	-65.14	-47.87	-45.84
		Mean Oct/12	-10.40	-9.58	-4.52	-4.47	-81.36	-71.49	-29.52	-28.07

OA-ICOS: water analyzer from Los Gatos Research; WS-CRDS: water analyzer from Picarro Inc.

samples collected at 20-25 cm depth, suggesting major contribution of groundwater (δ^{18} O and δ D mean values -4.21% and -29.46%, respectively, n = 2; see Table 2) and influence of rainwater (δ^{18} O and δ D mean values -1.51% and -10.34%, n = 3; see Table 2). The plot of δ^{18} O *versus* δ D for plant-water and soil-water (Figure 4) shows

profiles consistent with those observed for meteorical water (Figure 2) in the location, although the obtained slope is somewhat smaller than that of $LMWL_{GNIP}$ Such profile supports the observations described above and indicates robust connections with the local water cycle possibly due to the high annual rainfall volume.



Figure 3. Graphical representation of Mato Grosso (MT) results for rainwater, and waters from river, sugarcane xylem and soil. In the detail: description of the observed effects of drought in this sampling location, adapted from (A) references 9, 10; and (B) reference 44.

Mato Grosso

Plant- and soil-water from MT were the most depleted as shown in Table 2. This site is located at 528 m high and 275 km northwest from the reference station MT_{GNIP} in Cuiabá (165 m height), therefore isotopically depleted rainwater should be expected in relation to Cuiabá due to altitude and continental effects.^{10,39,45,46} However, as the crop is not irrigated and the sampling location undergoes severe drought during all the harvest period, the only important sources of water in August, 2011 and July, 2012 were the vadose zone and the phreatic. $\delta^{18}O$ and δD mean values found for plant-water (-9.09% and -71.76%, respectively) in August, 2011 and in July, 2012 (-7.71%) and -59.22%, respectively; see Table 2 and Figure 3) differ from the predicted groundwater isotopic fingerprint based on the weighted average for annual rainfall (MT_{GNIP}^a; see Table 1). ¹⁸O and D were actually more depleted than the estimated groundwater based on historical MT_{GNIP} data for precipitation in the rain period extending from November to April (MT_{GNIP}^{b}) ; see Table 1 and Figure 1).

Although the sampling location in Campo Novo do Parecis and the GNIP station in Cuiabá pertain to different hydrographic regions,²⁷ water isotope ratios in the mill's catchment point (MT water 1; see Table 1) were not significantly different from those of the estimated November-April MT_{GNIP} groundwater. Nevertheless, MT water 1 values may be biased due to evaporative enrichment along the course of the river⁴⁷ or during the time it resided in the large open mill's reservoir. For further investigation on the location water and groundwater profile a complementary sampling was conducted in September, 2014. Samples were collected in four points: the mill's reservoir (same of MT water 1), Do Sangue River, Verde River, and in two artesian wells inside the mill's facilities. The new data were in good agreement with those from MT water 1 and confirmed the local groundwater isotopic fingerprint, the early recharge period, and the mean composition of precipitation in the catchment basin. The differences observed between these results and those from plant-water (Table 2) indicate that vadose zone may be the source of water for the sugarcane crop during 2011 and 2012 drought periods, rather than groundwater. The plot of δ^{18} O versus δ D for plant-water (Figure 4) shows similar slope to MT_{GNIP}, demonstrating that evaporative enrichment does not occur; the only possible conclusion is the occurrence of strongly depleted rainwater in Campo Novo do Parecis during the rainy season.

October marks the beginning of the rainy season and the end of the harvest in the mill. The integrated δ^{18} O and δ D for precipitation throughout this month were -1.60%and -1.80%, respectively, confirming the expected high enrichment caused by the amount effect, also registered in historical data for the reference station MT_{GNIP} (δ^{18} O and δ D mean values: -2.20% and -4.06%, respectively; see Figure 1). Plant-water isotopic ratios (October, 2012, Table 2) were quite variable indicating a progressive mixing of the new soil moisture, incoming from rainwater, with the residual old vadose water. It is also evident an isotopic enrichment of the plant-water reaching δ^{18} O and δ D mean values of -4.15% and -34.48%, respectively. Large isotopic enrichment was observed in soil-waters collected at 0-5 cm and at 20-25 cm depth (Table 2). The enrichment in the upper layer indicates incidence of evaporative process. The plot of δ^{18} O *versus* δ D for plant-water and for soil-water (Figure 4), shows profiles consistent with those observed for local meteorical water (Figure 2), except for 0-5 cm soil-water. In this case, regression slope was significantly smaller than that for LMWL_{GNIP}, confirming a remarkable evaporative enrichment in the upper soil layer.

São Paulo

Results in SPA were atypical since only one sugarcane sample, collected on August, 2011 campaign (RB 867515, Table 2), presented results for plant-water consistent with the other matrices, like the respective soil-water (Table 2), lagoon and process waters (SPA water 1 and SPA water 2, Table 1). Additionally, the two remaining soil samples were highly depleted, with δ^{18} O and δ D mean values of -6.53% and -43.98%, respectively. The considerably reduced slopes of the linear regression for both plant and soil-water versus LMWL_{GNIP} indicate strong isotopic enrichment in the SPA area (Figure 4). Groundwater isotopic trends in that area⁴⁸ (isoscape with $\delta^{18}O = -8\%$) and $\delta D = -50\%$ and results of highly depleted mineral water samples (δ^{18} O and δ D mean values of -8.00% and -54.54‰, respectively) collected in sites up to 150 km away from the mill³² (Águas de Santa Bárbara and Bauru cities) fitted well when interpolated in the regression line of LMWL_{GNIP} (plot not shown). These literature results support the predicted isotopic fingerprint for that area but, also highlight the extraordinary isotopic shifts obtained for SPA samples. Further investigation must be carried out to better understand the processes leading to such shifts.

Rio Grande do Sul

In RS, plant-water presented the most constant results, with δ^{18} O and δ D mean values of -4.22% and -28.18%, respectively. These values are consistent with those for the Uruguay River water obtained in July and November, 2012 (RS water 1 and RS water 2; see Table 1), with the weighted average for the annual rainfall (RS_{GNP}^a, Table 1), and also with groundwater isotopic trends in the area⁴⁹ (isoscape with δ^{18} O -4.7% and δ D -29%). These plant-water results do not concur with those for sampled precipitation, as the four rainwater samples collected presented isotopic shifts

in a broad range of $\delta^{18}O$ (-1.13 to -9.6%) and δD (-0.3 to -65%) (Table 1). Soil-water was significantly more depleted than plant-water (Table 2). In October, 2012, soil-water ¹⁸O and D were strongly depleted in respect to July, suggesting that an important recent rain event led to shifting δ^{18} O and δ D mean values to -9.99% (± 1.08%) and -76.43% (± 6.73%), respectively, in both sampled depths. The incidence of recent rain event and its effects on the isotopic results for October, 2012 soil-water is similar to the observed in November, 2011 for sample RS Rain 2 (Table 1), and also seem to have impact on the plant-water isotopic ratios. The spread of plant-water $\delta^{18}O$ and δD values suggest water uptake from mixed sources such as the highly isotopicaly depleted rain event, the sampled rainwater and groundwater. The $\delta^{18}O$ versus δD linear regressions for both plant-water and soil-water (Figure 4) were compared to the LMWL_{GNIP} showing similar slopes, reinforcing these conclusions.

Ethanol and sugarcane

IRMS and nuclear magnetic resonance (NMR) are the two main analytical techniques used for determination of stable isotopes in ethanol⁵⁰ to assess adulteration in beverages. In this work, the 21 samples of hydrated, anhydrous and neutral ethanol collected in the four studied mills in 2011 and 2012 harvests were analysed for δD and $\delta^{13}C$ using CF-IRMS. Results were in the range –216.1 to –186.9‰ for δD and –13.24 to –12.04‰ $\delta^{13}C$ (Table 3).

Comparison of δD for ethanol samples with those for water extracted from the sugarcane crops used in the respective mill (Tables 2, 3 and Figure 5) revealed similar general trends, showing that independently of the ethanol specification (hydrated, anhydrous or neutral ethanol), δD is strongly influenced by the isotopic ratio from the plant-water, and consequently by the isotopic fluctuations inherent to the water cycle. In fact, it should be expected since hydrogen atoms in the ethanol methylene group derives from water present in the fermentative medium, composed basically of plant-water.^{4,5}

ANOVA applied to δD from 2011 harvest showed that ethanol samples produced in the four studied mills are statistically different, however, for the 2012 harvest only the biofuel produced in MT is statistically different ($p \le 0.05$). For $\delta^{13}C$ occurred the opposite as there was for 2011 harvest no significant difference in isotopic fingerprint while MT sample in 2012 was significantly different (Figure S1).

MT ethanol samples were strongly enriched in D from July to October, 2012 (Table 4) as also observed for water extracted from sugarcane (Table 3). The rainy season begins in October, and the rainwater collected during this



Figure 4. Local soil- and plant-water lines and comparison with GNIP rainwater data.

entire month also showed large enrichment in deuterium (Tables 1 and 2).

Sugar and fibers are present in equivalent proportion in sugarcane, with respective content of 10 to 17% and 8 to 14%.² Comparison of carbon isotope ratios for sugarcane stalks with those for ethanol (Table 4), reveals a minor tendency to ¹³C depletion in sugarcane, confirming that the fermentative process does not cause important carbon isotope fractionation during ethanol production.⁷

A recent study⁵¹ on the impact of grape varieties in the ethanol isotopic ratios of C, O and H in wines produced in different parts of Italy, highlighted that proper evaluation needed deep knowledge of environmental factors, like rainfall and water sources. The connection between sugarcane variety and isotope ratios in plant-water and ethanol was evaluated, but no clear relationship was detected. Here, in addition to the non-linear impact of environmental factors, the use in Brazil of several sugarcane varieties, continuously exchanged intra- and inter-mills during the harvest, hinders associations.

Conclusions

In general, surface water samples collected in sugarcane mills and nearby not irrigated crops in the four studied locations reflected the weighted average isotopic ratios of annual rainwater, and consequently the isotopic fingerprint of groundwater. For plant-water, the largest ¹⁸O and D depletion were found in MT samples during the drought period that occurs in the middle of the year. It was evidenced that vadose zone, rather than groundwater, is the source of water for the sugarcane crop in this location since these are the only available sources and results for groundwater do not match those found in plant-water. Isotopic shifts in

Mill location	Harvest	$\delta D_{ethanol}$ / ‰	SD	$\delta^{_{13}}C_{_{etanol}}$ / %0	SD	$\delta^{13}C_{sugarcane}$ / %0	SD
AM	Aug/2011	-200.3	3.9	-12.61	0.10	-13.04	0.05
	Nov/2011	-190.7	4.8	-12.31	0.01	-13.03	0.16
	Oct/2012	-191.7	2.2	-12.42	0.00	-12.45	0.03
	Oct/2012 (N)	-199.3	0.9	-13.24	0.12	_	_
	Nov/2012	-199.8	1.7	-12.54	0.07	-12.74	0.06
	Nov/2012 (N)	-195.8	2.5	-12.63	0.16	-	
MT	Aug/2011	-216.1	3.4	-12.55	0.06	-12.78	0.06
	Jul/2012	-203.2	1.8	-12.25	0.08	-12.06	0.09
	Oct/2012	-188.8	1.5	-12.04	0.09	-12.69	0.07
	Oct/2012 (A)	-190.0	0.3	-12.53	0.08	-	-
	Nov/2012	-186.9	1.6	-12.23	0.11	-	-
SPA	Aug/2011	-203.3	1.0	-12.32	0.10	-12.22	0.11
	Nov/2011	-199.7	3.0	-12.63	0.05	-13.24	0.09
	Aug/2012	-196.7	2.6	-12.40	0.03	-12.25	0.09
RS	Aug/2011	-207.7	0.9	-12.31	0.15	-13.13	0.00
	Nov/2011	-208.0	1.6	-12.34	0.15	-13.25	0.08
	Jul/2012	-197.1	3.5	-13.00	0.04	-13.05	0.08
	Oct/2012	-195.9	1.0	-12.62	0.08	-12.96	0.08

Table 3. Average isotopic ratios for ethanol and sugarcane

AM: Amazonas; MT: Mato Grosso; RS: Rio Grande do Sul; SPA: São Paulo; A: anhydrous ethanol; N: neutral ethanol.



Figure 5. Comparison of δD for ethanol and for plant-water.

RS were practically constant over the observation period in the present study. Isotopic ratios for AM samples, showed a robust connection with local water cycle, indicating that rainwater in this site is the main water source for the soil and crop. SPA results showed poor correlation between the different evaluated matrices, revealing also an unexpected enrichment for both plant- and soil-water when compared to the local meteoric water line. Soil-water results were considered to be in good agreement with those of the respective plant-water for the depth of 20-25 cm.

Similar seasonal variation of δD in ethanol and plantwater should be expected due to hydrogen exchange during production^{4,5} and to plant-water results reflecting the weighted average for annual rainfall isotopic ratios. This fact highlights the influence of hydrological cycle on the isotopic fingerprint of the alcohol. A regional pattern was found only for MT samples, since the statistical evaluation (ANOVA) of δD indicated no distinction among the three remaining sampling sites in both harvest years. As for $\delta^{13}C$ results, ANOVA groupped all four locations as an unique homogeneous group in 2011. The results of the present study pointed out that the use of hydrogen and carbon isotopic ratios for determination of ethanol geographical origin in Brazil is not trivial due to large variations in δD imposed by the hydrologic cycle, which directly affects ethanol production, and to the homogeneity of δ^{13} C in the assessed locations.

Supplementary Information

Supplementary information about El Niño/La Niña, statistical comparison of plant water results and ANOVA of ethanol results is available free of charge at http://jbcs.sbq.org.br as PDF file.

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