Expression of S-100, EMA, CD34 and presence of mast cells in eight oral neurofibromas, and a review of 127 cases of the literature.

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ABSTRACT

Introduction: The rarity of oral neurofibromas (ONs) generates problems regarding their epidemiological and immunohistochemical characterization. Objectives: The aim of this study was to evaluate the expression of different markers in ONs and review epidemiologic data reported in the literature. Material and methods: Clinicopathologic, immunohistochemical (markers S-100, epithelial membrane antigen [EMA], CD34) and histochemical (modified-Ziehl-Neelsen-method) studies were performed in eight cases of ON diagnosed in the Department of Pathology and Legal Medicine (DPML), Universidade Federal do Ceará (UFC), Ceará, Brazil, between 1994 and 2010. Results: Oral neurofibromas represented 0.2% of the oral lesions diagnosed by our service in 16 years, and the buccal mucosa was the most frequent oral site (71.4%). Seven (87.5%) and 8 (100.0%) cases were positive for S-100 and CD34, respectively, and none for EMA. Mast cells were identified in seven cases (87.5%). The literature search indicated that solitary ONs are more common and occur preferentially in females, affecting patients between 30 and 40 years old. The alveolar ridge is the most commonly involved site. Conclusion: S-100- and CD34 markers proved to be of great value as a diagnostic tool, unlike EMA staining. Identification of mast cells in most cases suggests their involvement in this tumor pathogenesis. The clinicopathologic data retrieved from the literature enabled the establishment of a more consistent epidemiological profile.

Key words: neurofibroma; immunohistochemistry; oral cavity; mast cells.

INTRODUCTION

Neurofibroma is a slow-growing, relatively circumscribed, benign tumor. Along with schwannoma, it is one of the most common benign neoplasms of the neural sheath. Neurofibromas can occur as solitary lesions or in association with multiple other neurofibromas of the skin, café-au-lait macules, freckling in the axillary and inguinal regions, Lisch nodules, and abnormalities of bone development as part of a syndrome known as neurofibromatosis type 1 (NF1) or von Recklinghausen disease.

NF1 is a disorder of unknown etiology that is triggered by a mutation in the NF1 tumor suppressor gene, located on the long arm of chromosome 17. This gene encodes the protein neurofibromin, which inactivates the RAS intracellular signaling cascade, a key component of many growth factor signaling pathways.
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pathways. In the absence of neurofibromin, the RAS cascade is constitutively activated, resulting in increased cellular proliferation and survival(14, 33, 35).

Neurofibromas are rare tumors that typically affect the skin and subcutaneous tissue. In cases of isolated neurofibromas, patients are usually diagnosed between the ages of 20 and 40 years, although patients with tumors associated with the NF1 syndrome are usually diagnosed at younger ages(4, 18, 23). The prevalence of isolated oral neurofibromas (ONs) is uncertain; however, in patients with the syndrome, oral manifestations of the condition occur in up to 17% of patients(35). Some studies indicate a slight female predominance for both the syndrome and isolated cases(3, 7, 25, 28), whereas others observe no distinction between the sexes(50). Approximately 25% of neurofibromas are found in the head and neck region, and rare involvement of the oral cavity has been reported in sites such as the lips, tongue, buccal mucosa and alveolar ridge. Neurofibromas usually present as a superficial, painless nodule with nonspecific clinical symptoms and resemble fibrous hyperplasia, fibromas, or lipomas(27, 23, 24, 28).

Ultrastructurally, neurofibromas exhibit significant heterogeneity, largely due to the involvement of a variety of cellular constituents: Schwann cells, perineural-like cells, endoneurial fibroblasts, intermediate cells, and mast cells(35, 33). Immunohistochemistry (IHC) has been instrumental in confirming this heterogeneity and represents an auxiliary method in the diagnosis and identification of this neural tumor(6, 15, 29). The S-100 protein marker has been widely used in the identification of Schwann cells and is important for the diagnosis of neural sheath tumors(30). However, the differential diagnosis of neurofibromas from other lesions (schwannomas, leiomyomas, and myofibromas) requires the use of additional immunohistochemical stains and histochemical reagents to identify the several components of the tumor(4, 11, 24, 28).

The rarity of oral cavity involvement in neurofibroma is reflected by the paucity of studies that exclusively consider this tumor and its intra-oral location, what generates problems regarding the epidemiological and immunohistochemical characterization of ONs(7, 11, 28, 29).

The aim of this paper was to immunohistochemically characterize eight cases of neurofibromas located in the oral cavity using S-100, epithelial membrane antigen (EMA) and CD34 markers in addition to histochemical analyses using the modified Ziehl-Neelsen method. A literature review highlighting the epidemiological characteristics of ONs was performed to contribute to the identification and/or understanding of the behavior of neurofibromas.

MATERIAL AND METHODS

This study was approved by the Research Ethics Committee of Universidade Federal do Ceará (Protocol 280/05). We reviewed all cases of oral cavity neurofibromas that had been recorded between 1994 and 2010 in the files of the Department of Pathology and Legal Medicine (DPMIL, UFC, Ceará, Brazil). Qualitative analyses of the microscopic characteristics of the tumors were performed by an oral pathologist (A.P.N.A) after the tissue was sectioned, stained with hematoxylin and eosin (HE), and prepared for histochemical (modified Ziehl-Neelsen stain) and immunohistochemical study. Clinical data related to the patients, such as sex, age, and the tumor characteristics and anatomical site, were retrieved from the biopsy records (Table 1).

<table>
<thead>
<tr>
<th>Case nº</th>
<th>Age (years)</th>
<th>Sex</th>
<th>Location</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>49</td>
<td>F</td>
<td>Buccal mucosa</td>
</tr>
<tr>
<td>2</td>
<td>69</td>
<td>M</td>
<td>Buccal mucosa</td>
</tr>
<tr>
<td>3</td>
<td>76</td>
<td>M</td>
<td>Buccal mucosa</td>
</tr>
<tr>
<td>4</td>
<td>69</td>
<td>M</td>
<td>Buccal mucosa</td>
</tr>
<tr>
<td>5</td>
<td>39</td>
<td>F</td>
<td>Buccal mucosa</td>
</tr>
<tr>
<td>6</td>
<td>NI</td>
<td>F</td>
<td>Hard palate</td>
</tr>
<tr>
<td>7</td>
<td>41</td>
<td>M</td>
<td>NI</td>
</tr>
<tr>
<td>8</td>
<td>NI</td>
<td>M</td>
<td>Inferior lip</td>
</tr>
</tbody>
</table>

DPMIL: Department of Pathology and Legal Medicine; NI: non-informed.

Immunohistochemical analyses were performed using the standard streptavidin-biotin-peroxidase method in 5-μm-thick tissue sections that had been obtained from paraffin-embedded blocks and mounted on silanized microscopic slides. The steps from deparaffinization to heat-induced epitope retrieval were performed with an EZ Prep Solution (Ventana Tucson, AZ, USA) for 30 minutes. The primary antibodies used in our study included S-100 protein (1:2000, DakoCytomation® Carpinteria, CA, USA), EMA (1:200, DakoCytomation® Carpinteria, CA, USA) and CD34 (1:100, DakoCytomation® Carpinteria, CA, USA). The BenchMark™ XT IHC/in situ hybridization (ISH) (Ventana Tucson, AZ, USA) automated slide stainer was used, and the indirect immunoperoxidase was detected by the XT Ultraview diaminobenzidine (DAB) v3 system (Ventana Tucson, AZ, USA) followed by revelation of the reaction end product by exposure to DAB. The sections were subsequently counterstained with Mayer hematoxylin. Positive extrinsic and intrinsic control samples were used in each assay.
The positive results were expressed as the intensity (+ mild, ++ moderate or +++ intense) and extent (focal/diffuse) of staining. In addition, the term rare cells was applied to the samples in which the number of immunoreactive cells was considered very low.

To identify mast cells, a modified Ziehl-Neelsen stain was performed. Histological slides were prepared from the 5-μm-thick sections of the paraffin-embedded blocks. To deparaffinize the sections, a 60ºC solution of liquid paraffin and turpentine was dripped onto the histological sections and left for 20 minutes. Subsequently, the sections were hydrated with absolute ethanol and washed in running water. The sections were then stained with Ziehl Fuchsin for 50 minutes. After a second wash, the material was decolorized with 5% sulfuric acid until the sections were pale pink. Finally, the slides were counterstained, washed rapidly in running water, dried and mounted.

The histochemical reaction of the modified Ziehl-Neelsen stain was evaluated with an Olympus CX 41 optic microscope. The letter P (positive reaction) was used to identify positive cases, and the letter A (absence of expression), to identify negative cases (Table 2).

<table>
<thead>
<tr>
<th>Case nº</th>
<th>S-100</th>
<th>EMA</th>
<th>CD34</th>
<th>Ziehl-Neelsen</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>++++</td>
<td>D</td>
<td>+++</td>
<td>P</td>
</tr>
<tr>
<td>2</td>
<td>++</td>
<td>F</td>
<td>+++</td>
<td>P</td>
</tr>
<tr>
<td>3</td>
<td>++++</td>
<td>D</td>
<td>++</td>
<td>RC, A</td>
</tr>
<tr>
<td>4</td>
<td>A</td>
<td>+</td>
<td>D, RC</td>
<td>P</td>
</tr>
<tr>
<td>5</td>
<td>++++</td>
<td>D</td>
<td>+++</td>
<td>RC, A</td>
</tr>
<tr>
<td>6</td>
<td>++</td>
<td>RC</td>
<td>+</td>
<td>F, P</td>
</tr>
<tr>
<td>7</td>
<td>++</td>
<td>D</td>
<td>++</td>
<td>F, RC, A</td>
</tr>
<tr>
<td>8</td>
<td>++++</td>
<td>D</td>
<td>+++</td>
<td>F, P</td>
</tr>
</tbody>
</table>

EMA: epithelial membrane antigen; +++: intense immunoreactivity; ++: moderate immunoreactivity; +: mild immunoreactivity; D: diffuse extent of staining; F: focal extent of staining; A: absence of expression; RC: rare cells; P: positive reaction.

In addition, we searched the literature (PubMed Database) for case series of ONs that included epidemiological data. A manual search and the key words "neurofibroma", "oral cavity", and "immunohistochemistry" were used. Only the articles published in indexed journals were considered. On the basis of this search, seven articles within the period 1971 to 2011 were selected without date restrictions and after critical analyses (Table 3).

TABLE 2 – Immunohistochemical and histochemical profile of oral neurofibromas diagnosed between 1994 and 2010 in the DPML, Universidade Federal do Ceará, Brazil

RESULTS

ONs represented eight (0.2%) of the 3,730 oral lesions collected between 1994 and 2010 from the DPML of UFC.

Analyses of the eight cases of ONs revealed a male predominance (5:3), with patient ages varying from 39 to 76 years (mean = 57.1 ± 5.65 years). The most frequently involved intraoral site was the buccal mucosa (71.4%), and none of the lesions was associated with NF1 (Table 1).

Positive immunohistochemical staining for S-100 protein (Figure 1) was observed in seven specimens, four of which exhibited moderate staining, and three of which stained intensely. Of the seven S-100 positive specimens, five were diffusely stained, and two were focally stained (with one case exhibiting rare cells). EMA immunoexpression was not observed in any of the evaluated cases. All of the samples were immunopositive for CD34 (Figure 2), and the degree of staining intensity varied from mild (two cases) to moderate (three cases) to intense (three cases). The extent of staining was diffuse in five cases (with three cases exhibiting rare cells), and focal in three cases (with one case exhibiting rare cells). In all cases, vessels were positive for CD34 (Table 2). Histochemical analysis using a modified Ziehl-Neelsen stain revealed a small number of mast cells that were largely localized to the perivascular region and the periphery of the lesion (Table 2).

DISCUSSION

A literature search for case studies of ONs identified a total of 127 cases(3, 9-13, 22). ONs were observed in female patients in 64.3% of cases, with a male to female ratio of 1.00:1.80. The mean patient

FIGURE 1 – Positive immunohistochemical expression of S-100 protein in neurofibromas (LSAB 400×)

LSAB: labeled streptavidin-biotin method.
TABLE 3 – Clinicopathological data of 127 intraoral neurofibromas reported in the literature within the period 1971-2011 (PubMed database). Case studies and case reports

<table>
<thead>
<tr>
<th>Authors</th>
<th>Number of cases</th>
<th>Cases associated with NF1 N (%)</th>
<th>Mean age (variation) years</th>
<th>Gender (M/F)</th>
<th>Location (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cherrik and Eversole, 1971</td>
<td>19</td>
<td>4 (21)</td>
<td>NI</td>
<td>7/12</td>
<td>Buccal mucosa (7) Tongue (6) Palate (3) Lip (1) Alveolar ridge (1) Others * (1)</td>
</tr>
<tr>
<td>Chen and Miller, 1979</td>
<td>49</td>
<td>NI</td>
<td>39.5 (9 to 72)</td>
<td>21/28</td>
<td>Alveolar ridge* (24) Palate (9) Tongue (6) Buccal mucosa (5) Lip (5) Mandible (2) NI (2)</td>
</tr>
<tr>
<td>Crysomali et al., 1997</td>
<td>16</td>
<td>NI</td>
<td>45.1 (-)</td>
<td>5/10</td>
<td>Alveolar ridge (7) Palate (5) Lip (2) Buccal mucosa (1) NI (1)</td>
</tr>
<tr>
<td>Marocchio et al., 2007</td>
<td>11</td>
<td>2 (18.1)</td>
<td>NI</td>
<td></td>
<td>Alveolar ridge (5) Tongue (2) Buccal mucosa (1) Lip (1) Mandible (1) Others * (1)</td>
</tr>
<tr>
<td>Salla et al., 2009</td>
<td>12</td>
<td>2 (16.6)</td>
<td>31.2 (9 to 50)</td>
<td>2/10</td>
<td>Palate (4) Alveolar ridge (4) Tongue (2) Lip (1) Others * (1)</td>
</tr>
<tr>
<td>El-Gehani et al., 2009</td>
<td>8</td>
<td>NI</td>
<td>NI</td>
<td>4/4</td>
<td>Mandible (3) Tongue (5) Maxilla (1) Others * (1)</td>
</tr>
<tr>
<td>do Nacimiento et al., 2009</td>
<td>12</td>
<td>3 (25)</td>
<td>35.7 (6 to 78)</td>
<td>2/10</td>
<td>Alveolar ridge (5) Lip (2) Tongue (1) Maxilla (1) Buccal mucosa (1) Others * (4)</td>
</tr>
<tr>
<td>Total</td>
<td>127</td>
<td>11 (20.1)</td>
<td>37.8 (6 to 78)</td>
<td>41/74</td>
<td>Alveolar ridge (44) Palate (21) Tongue (20) Lip (10) Mandible (6) Buccal mucosa (13) Maxilla (2) Others * (8) NI (5)</td>
</tr>
</tbody>
</table>

NF1: neurofibromatosis type 1; M: male; F: female; NI: non-informed; *: gingival cases were included in this group; others *: retromolar area, mental region, floor of the mouth, maxillary vestibulum.
S-100 protein in 85% to 100% of the occasions\(^\text{19, 32}\), although rare findings of negative S-100 immunostaining can occur\(^6\), as seen in one case in the present study. Immunohistochemical markers have commonly been used in the identification of this tumor, especially when a differential diagnosis is necessary. Lesions such as schwannomas, leiomyomas, leiomyosarcomas, myofibromas, fibrosarcomas, fibrohistiosarcomas and malignant peripheral nerve sheath tumors also exhibit spindle-shaped cells\(^5, 7, 20, 26\).

S-100 protein is found predominantly, but not exclusively, in the nervous system. This protein is expressed by Schwann cells\(^5\) and is very useful for the identification of tumors composed of these cells\(^5\). Neurofibromas present a variable number of S-100 positive cells that may localize to different areas of the same tumor\(^5\), corroborating our findings of the variable extent of staining (diffuse or focal) for these cells. In addition, the cell nuclei and cytoplasm in the present study exhibited frequent variations in the staining intensity, which corroborates the findings of Nascimento et al.\(^7\). Neurofibromas are immunopositive for the S-100 protein in 85% to 100% of the occasions\(^9, 32\), although rare findings of negative S-100 immunostaining can occur\(^6\), as seen in one case in the present study.

FIGURE 2 – Positive immunohistochemical expression of CD34 in neurofibromas (LSAB 200×).

LSAB: labeled streptavidin–biotin method.
In our series, none of the cases exhibited spindle cells whose nuclei or cytoplasm stained positive for EMA. This marker is expressed in perineural cells, and normal nerves are characterized by EMA-positive cells and S-100 protein-negative cells(15). Despite the existence, as demonstrated by ultrastructural studies, of perineural-like cells in neurofibromas, immunohistochemical studies using EMA as a marker for these cells have produced variable expression(15, 25, 38).

The CD34 antigen, expressed by hematopoietic progenitor cells of myeloid and lymphoid lineages, has also been recently identified in peripheral nerve sheath tumors(21). CD34-positive cells in neurofibromas are distinct from Schwann cells, perineural cells and “conventional” fibroblasts, suggesting that they may correspond to the endoneurial fibroblasts that play a supportive role for Schwann cells(15, 21). The variable expression of CD34 immunostaining in the present study corroborates other reports(15, 21, 37), and those differences in expression levels have been attributed to the type of stroma, fibrous or myxoid(37), and the age of the lesion(21). The CD34 staining pattern in neurofibromas is different from the pattern seen in schwannomas. In the latter, only Antoni B areas, located near the capsule of the neoplasm, stain positive for CD34(37). Malignant peripheral nerve sheath tumors, neurothekeomas and perineuriomas express very low levels of the CD34 marker or none at all(15, 17, 34, 37).

The antibodies used in the present study were selected from a literature review. In addition to S-100 protein, other markers, such as EMA and CD34, have been used in ultrastructural studies to highlight the heterogeneity of neurofibromas(15). However, the shortage of immunohistochemical studies using these markers to identify intraoral tumors is reflected in the relative dearth of knowledge of these immunostains and their use in the daily practice of neurofibroma diagnosis. This lack of knowledge is a complicating factor when the identification of other components of the tumor is necessary for the differential diagnosis from other lesions.

With the rare incidence of solitary ONs and their unique histological and clinical characteristics, the diagnosis of this neoplasm is a challenge for many clinicians and oral pathologists. Therefore, we hope that the epidemiological and immunohistochemical data provided by this study will contribute to the understanding of the pathogenesis of these lesions and guide their correct diagnosis and treatment.

CONCLUSION

In the present study, the prevalence of ONs was rare, and no cases were associated with NF1 lesions. The analyzed lesions exhibited positive S-100 and CD34 expression as well as positive mast cells, which were identified through a modified histochemical method, and negative EMA expression. The association of these data with the clinicopathologic data of 127 cases retrieved from the literature enabled the establishment of a more consistent epidemiological profile. In addition, the immunohistochemical and histochemical aspects addressed here proved to be important to the determination of markers useful for the identification of tumorous cells, thereby aiding in correct diagnoses.

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RESUMO

Introdução: A rara ocorrência de neurofibromas orais (NO) gera problemas com relação à caracterização epidemiológica e imuno-histológica dessas lesões. Objetivos: Avaliar a expressão celular de diferentes marcadores em NO e revisar dados epidemiológicos reportados na literatura. Material e métodos: Estudos clinicopatológico, imuno-histológico (marcadores S-100, epithelial membrane antigen [EMA] e CD-34) e histoquímico (método Ziehl-Neelsen modificado) foram realizados em oito casos de NO diagnosticados no Departamento de Patologia e Medicina Legal (DPML) da Universidade Federal do Ceará (UFC), Ceará, Brasil, entre 1994 e 2010. Resultados: Os NO representaram 0,2% das lesões bucais diagnosticadas pelo serviço em 16 anos; mucosa jugal foi a localização mais frequente (71,4%); sete (87,5%) e oito (100%) casos apresentaram positividade para S-100 e CD-34, respectivamente, e nenhum para EMA. Adicionalmente, mastócitos foram identificados em sete casos (87,5%). A revisão
de literatura indicou que NO solitários são mais comuns e ocorrem preferencialmente em mulheres, afetando pacientes entre 30 e 40 anos. O rebordo alveolar é a localização intraoral mais comum. Conclusão: Os marcadores S-100 e CD34 provaram ser de grande valor como ferramentas diagnósticas, diferente da coloração EMA. A identificação de mastócitos na maioria dos casos sugere seu envolvimento na patogênese desse tumor. Os dados clinicopatológicos da revisão de literatura ajudaram no estabelecimento de um perfil epidemiológico mais consistente.

Unitermos: neurofibroma; imuno-histoquímica; boca; mastócitos.

REFERENCES


