Cellular composition of induced sputum in healthy adults*

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Abstract

Objective: To establish reference values for cellularity in induced sputum samples collected from healthy adults.

Methods: Induced sputum samples were obtained from 88 healthy adult never-smokers (39 males). The mean age was 36 years (range, 18-68 years). The participants had been residing in the city of Florianópolis, Brazil (a medium-sized non-industrial city) for at least two years. After the samples had been processed, we obtained total and differential cell counts.

Results: The mean total cell count was $4.8 \pm 4.2 \times 10^6$ cells/g. There was a predominance of macrophages (mean, $77.5 \pm 14.7\%$) and neutrophils (mean, $23.4 \pm 14.3\%$). Eosinophils were virtually absent (mean, $0.1 \pm 0.3\%$). Lymphocytes and bronchial epithelial cells were scarce. Neither age nor atopy had any effect on the total or differential cell counts.

Conclusions: In the induced sputum of this healthy adult population, macrophages and neutrophils predominated. However, the proportion of neutrophils was lower than that reported in previous studies, which suggests that reference values might vary depending on geographic location.

Keywords: Sputum; Reference values; Brazil.

Resumo

Objetivo: Estabelecer valores de referência para a celularidade de amostras de escarro induzido coletadas de indivíduos adultos saudáveis. Métodos: O escarro induzido foi obtido de 88 adultos saudáveis que nunca fumaram (39 homens) com média de idade de 36 anos (variação: 18-68 anos) residentes há pelo menos dois anos em Florianópolis, uma cidade brasileira não industrial e de tamanho médio. As amostras foram processadas, e foi realizada a contagem total e diferencial das células. Resultados: A média da contagem celular total foi de $4,8 \pm 4,2 \times 10^6$ células/g. Houve predominio de macrófagos (média de $77,5 \pm 14,7\%$) e de neutrófilos (média de $23,4 \pm 14,3\%$). Os eosinófilos estiveram virtualmente ausentes (média de $0,1 \pm 0,3\%$). A proporção de linfócitos e de células broncoepiteliais foi pequena. Não houve efeito da idade ou de atopia sobre a contagem celular total ou diferencial. Conclusões: Nesta população de indivíduos saudáveis, macrófagos e neutrófilos foram as células predominantes no escarro induzido. Contudo, a proporcão de neutrófilos foi inferior à previamente relatada, sugerindo que os valores de normalidade podem variar de acordo com o local onde ele é amostrado.

Descritores: Escarro; Valores de referência; Brasil.
Introduction

Nearly two decades ago, the introduction of induced sputum to assess airway inflammation provided a breakthrough in the understanding of the pathophysiology of airway diseases. Currently, there is sufficient evidence that induced sputum is the most comprehensive method for noninvasive examination of airway inflammation because of its properties (reliability, reproducibility, and responsiveness). In addition, the results of recent studies have consistently shown that induced sputum is an important tool for the determination of the phenotype of asthma, COPD, and other airway diseases, as well as for the study of the effect of treatment of these diseases. The method has been important for the study of the pathogenesis of obstructive and infectious respiratory diseases. In contrast, few studies have examined the cellular composition of induced sputum in healthy subjects.

Knowledge of normal values for induced sputum measurements, especially as regards the proportion of eosinophils and neutrophils (the two cell types most commonly used for characterizing the inflammatory response of the airways), is essential to understand the result of this test. The use of induced sputum, together with pulmonary function tests, makes it possible to monitor the pattern of inflammation more appropriately in specific respiratory conditions. However, total and differential cell counts in induced sputum samples should be delimited by normal reference values.

To date, only three studies, all of which involved large samples of healthy subjects, have provided reference values for total and differential cell counts in induced sputum. The only difference among the results of those studies was in the proportion of neutrophils.

Reference values for induced sputum cellularity can be influenced by regional characteristics (such as ambient air pollution), atopy, and age, as well as by the techniques employed in the induction and processing of sputum. The objective of the present study was to establish reference values for total and differential cell counts in induced sputum samples collected from healthy adult never-smokers residing in the city of Florianópolis, Brazil.

Methods

A total of 99 healthy adult never-smokers who had been residing in Florianópolis, a medium-sized non-industrial city located in southern Brazil, for at least two years were recruited through advertisements in the local media and posters displayed at the local university. We applied the following exclusion criteria: presenting with respiratory symptoms (nasal or pulmonary); having a history of respiratory diseases, including asthma and allergic rhinitis; having a history of occupational exposure to dust, chemical materials, or smoke; and having experience symptoms of respiratory infection in the last four weeks. All subjects had normal spirometry results (FEV₁ > 80% of predicted, FEV₁/FVC ratio ≥ 80%, and normal methacholine responsiveness—the provocative concentration of methacholine causing a 20% fall in FEV₁ [PC₂₀] being > 8.0 mg/mL). Of the 99 subjects evaluated, 27 were atopic, as indicated by the presence of positive skin test reactivity to one or more common allergens. The study was approved by the Human Research Ethics Committee of the Universidade Federal de Santa Catarina (UFSC, Federal University of Santa Catarina; Protocol no. 084/84). All volunteers gave written informed consent.

This was a single-visit, cross-sectional study. All subjects were evaluated in the research laboratory of the UFSC Center for Airway Inflammation Research. After having been screened for the inclusion and exclusion criteria, the subjects with normal spirometry results and normal methacholine responsiveness underwent sputum induction and skin testing for the investigation of atopy.

The characteristics of the volunteers were documented with a structured questionnaire. Spirometry was performed in accordance with the American Thoracic Society guidelines. The predicted reference values were those established by Crapo et al. Methacholine challenge was performed in accordance with the method described by Juniper et al., and the results are expressed as PC₂₀ in cumulative units. Skin testing was performed by a modified puncture technique with the use of 14 extracts of common inhalant allergens, 1 negative control (glycerol), and 1 positive control (histamine). Test results were considered positive when the papule size was 3 mm greater than was that of
the negative control. Atopy was defined as the presence of one or more positive test results.

Sputum was induced and processed by the method described by Pizzichini et al.[3]. In brief, the procedure was initiated 15 min after the administration of 200 µg of inhaled albuterol, through the inhalation of increasing concentrations of saline (3%, 4%, and 5%), each inhaled for 7 min consecutively or until there was a decrease in FEV₁ ≥ 20% in relation to baseline values. Saline nebulization was performed with a Fison ultrasonic nebulizer (Fisons, Pickering, Ontario, Canada), with an output rate of 0.87 mL/min and particles presenting a median aerodynamic mass diameter of 5.58 µm. After each inhalation period, FEV₁ was measured to ensure the safety of the test. If there was a decrease in FEV₁ ≥ 10% in relation to baseline values, the saline concentration was not increased. Samples were considered appropriate if total and differential cell counts could be obtained from material with cell viability of at least 50% and contamination by oropharyngeal squamous cells lower than 20%.

The sample size was estimated on the basis of previous calculations.[14-16] Categorical variables (gender, race, atopy, and induction success) are expressed as absolute frequencies and percentages. Continuous variables were initially evaluated in terms of the criteria of normality. Variables with a continuous distribution (age, weight, height, baseline FEV₁, FEV₁/FVC ratio, and post-albuterol FEV₁) are expressed as means and standard deviations or ranges (minimum and maximum). Because of the non-Gaussian distribution of the total cell counts and of some cellular components (eosinophils and lymphocytes), sputum cytology results are expressed as means and standard deviations, as medians and interquartile ranges, and as the 10th and 90th percentiles. Differences among groups were analyzed by ANOVA or unpaired t-test. Values of p < 0.05 were considered statistically significant. All tests were two-tailed. Data were analyzed with the Statistical Package for the Social Sciences, version 18.0 for Windows (SPSS Inc., Chicago, IL, USA).

**Results**

Sputum induction produced an adequate sample in 88 participants (39 males). The mean age was 36 years (range, 18-68 years). The success rate of sputum induction was 88.9%. We excluded 11 sputum samples because of insufficient sputum volume for evaluation or lack of cell viability. In the remaining samples, the quality of sputum obtained was satisfactory, as indicated by a mean viability of 77.5 ± 11.1%, with a median (90th percentile) of 79.0% (94.0%).

### Table 1 – Total and differential cell counts (× 10⁶ cells/g) in induced sputum samples collected from healthy adult subjects.

<table>
<thead>
<tr>
<th>Cell counts</th>
<th>Mean ± SD</th>
<th>Median (II)</th>
<th>10th percentile</th>
<th>90th percentile</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>4.8 ± 4.2</td>
<td>3.5 (4.0)</td>
<td>1.2</td>
<td>10.8</td>
</tr>
<tr>
<td>Eosinophils</td>
<td>0.004 ± 0.010</td>
<td>0 (0)</td>
<td>0</td>
<td>0.020</td>
</tr>
<tr>
<td>Neutrophils</td>
<td>1.4 ± 2.1</td>
<td>0.9 (1.4)</td>
<td>0.1</td>
<td>3.0</td>
</tr>
<tr>
<td>Macrophages</td>
<td>3.2 ± 2.5</td>
<td>2.3 (3.0)</td>
<td>0.6</td>
<td>6.8</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>0.10 ± 0.10</td>
<td>0.09 (0.20)</td>
<td>0</td>
<td>0.30</td>
</tr>
<tr>
<td>Bronchial epithelia cells</td>
<td>0.10 ± 0.10</td>
<td>0.10 ± 0.10</td>
<td>0.01</td>
<td>0.30</td>
</tr>
</tbody>
</table>

II: interquartile range.

### Table 2 – Differential non-squamous cell counts in induced sputum samples from healthy subjects in the study.

<table>
<thead>
<tr>
<th>Differential cell counts</th>
<th>Mean ± SD</th>
<th>Median (II)</th>
<th>10th percentile</th>
<th>90th percentile</th>
</tr>
</thead>
<tbody>
<tr>
<td>Eosinophils</td>
<td>0.1 ± 0.3</td>
<td>0 (0)</td>
<td>0</td>
<td>0.5</td>
</tr>
<tr>
<td>Neutrophils</td>
<td>23.4 ± 14.3</td>
<td>21.7 (21.0)</td>
<td>6.9</td>
<td>41.3</td>
</tr>
<tr>
<td>Macrophages</td>
<td>68.4 ± 14.7</td>
<td>71.0 (17.9)</td>
<td>47.9</td>
<td>86.0</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>3.1 ± 2.4</td>
<td>3.0 (3.0)</td>
<td>0</td>
<td>6.5</td>
</tr>
<tr>
<td>Bronchial epithelial cells</td>
<td>5.2 ± 7.9</td>
<td>2.8 (4.9)</td>
<td>0.5</td>
<td>13.3</td>
</tr>
</tbody>
</table>

II: interquartile range.
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Eosinophils and lymphocytes, are similar to those in the literature.\(^{14,15}\) The percentage of healthy adults excluded from the sample evaluated is also similar to that reported in previous studies.\(^{14,15}\) However, the proportion and the absolute number of neutrophils found in our study were lower than those previously reported. Knowledge of reference values for sputum cellularity, especially as regards neutrophils and eosinophils, is important for understanding and interpreting the meaning of inflammatory subtypes—neutrophilic, eosinophilic, mixed granulocytic, and paucigranulocytic—which have often been used for determining the phenotype of asthma.\(^{21}\)

The lower proportion of neutrophils found in the present study cannot be attributed to technical aspects, because the sputum induction and processing methods employed are similar to those used in previous studies.\(^{14,15,22}\) Nor does it seem to be due to bias in the selection of participants, because we found that neither age nor atopy had any effect on the total or differential cell counts. We speculate that this lower proportion of neutrophils was caused by local variations, such as environmental pollution.

The present study was not designed to investigate the effect of environmental pollution on airway secretions. Instead, it was designed to establish normal values for induced sputum cellularity in healthy adults not exposed to environmental pollution, because all participants had been residing in a non-industrial city for at least two years. We cannot rule out previous household or environmental exposure with certainty. However, none of the study subjects

### Table 3 - Total and differential cell counts in induced sputum samples from healthy subjects in the study by age bracket.\(^{a}\)

<table>
<thead>
<tr>
<th>Cell counts</th>
<th>Age bracket, years</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>18-29</td>
</tr>
<tr>
<td></td>
<td>(n = 39)</td>
</tr>
<tr>
<td>Total</td>
<td>4.1 ± 3.2</td>
</tr>
<tr>
<td>Eosinophils</td>
<td>0.10 ± 0.30</td>
</tr>
<tr>
<td>Neutrophils</td>
<td>20.2 ± 14.5</td>
</tr>
<tr>
<td>Macrophages</td>
<td>73.0 ± 14.8</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>3.1 ± 2.5</td>
</tr>
<tr>
<td>Bronchial epithelial cells</td>
<td>4.3 ± 5.2</td>
</tr>
</tbody>
</table>

\(^{a}\)Values expressed as mean ± SD.

### Table 4 - Total and differential cell counts in induced sputum samples from healthy subjects in the study by presence or absence of atopy.\(^{a}\)

<table>
<thead>
<tr>
<th>Cell counts</th>
<th>Atopy</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Present</td>
</tr>
<tr>
<td></td>
<td>(n = 27)</td>
</tr>
<tr>
<td>Total</td>
<td>3.5 ± 3.0</td>
</tr>
<tr>
<td>Eosinophils</td>
<td>0.20 ± 0.30</td>
</tr>
<tr>
<td>Neutrophils</td>
<td>26.7 ± 16.4</td>
</tr>
<tr>
<td>Macrophages</td>
<td>64.1 ± 15.1</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>3.8 ± 2.9</td>
</tr>
<tr>
<td>Bronchial epithelial cells</td>
<td>4.9 ± 6.1</td>
</tr>
</tbody>
</table>

\(^{a}\)Values expressed as mean ± SD.
had been exposed to environmental pollution in the last two years.

Environmental exposure results from contact with a variety of gaseous and particulate pollutants, among which the most harmful to human health seems to be particulate matter. Repeated inhalation of particulate pollutants can result in a moderate level of oxidative stress and a moderate degree of inflammatory response in the airways and lungs. In recent years, various studies have investigated the effect of acute and chronic exposure of the lungs to environmental pollution. Studies using bronchoalveolar lavage or induced sputum have shown that healthy subjects exposed to ozone or diesel exhaust have an inflammatory response characterized by an increase in neutrophils and IL-6. Changes in induced sputum can be observed as early as six hours after exposure. In addition, it has been shown that healthy workers who are exposed to traffic pollution on a daily basis have sputum neutrophilia. Occupational exposure should also be considered a major modulator of the cellular composition of induced sputum. Although it is reasonable to assume that the low proportion of neutrophils in our sample is associated with lower exposure to environmental pollution, this hypothesis needs to be confirmed or refuted in studies specifically designed to investigate this aspect.

Finally, it is important to mention that, in 85% of the study subjects, there were no eosinophils in the induced sputum samples. This result confirms previous reports and supports the concept that the presence of eosinophils in sputum is a robust and reliable marker of eosinophilic inflammation. Our results also suggest that the current cut-off value of 3.0% for eosinophilic inflammation can be lowered.

In summary, the results of the present study show that, in this population of healthy subjects, there was a predominance of macrophages and neutrophils in induced sputum samples. However, the proportion of neutrophils was lower than that reported in previous studies, which suggests that reference values might vary by geographic location.

References


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