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Understanding the complexity of *Tityus* serrulatus venom: A focus on high molecular weight components

Isadora Sousa de Oliveira^{1,2} ^(D), Nicoly Malachize Alano-da-Silva¹, Isabela Gobbo Ferreira¹, Felipe Augusto Cerni³, Jacqueline de Almeida Gonçalves Sachett^{4,5} ^(D), Wuelton Marcelo Monteiro^{4,5} ^(D), Manuela Berto Pucca⁶ ^(D), Eliane Candiani Arantes^{1*} ^(D)

¹Department of BioMolecular Sciences, School of Pharmaceutical Sciences of Ribeirão Preto, University of São Paulo, Ribeirão Preto, SP, Brazil. ²Department of Biotechnology and Biomedicine, Technical University of Denmark, Kongens Lyngby, Denmark.

³Health and Sciences Postgraduate Program, Federal University of Roraima, Boa Vista, RR, Brazil.

⁴School of Health Sciences, Amazonas State University, Manaus, AM, Brazil.

⁵Department of Teaching and Research, Dr. Heitor Vieira Dourado Tropical Medicine Foundation, Manaus, AM, Brazil.

⁶Department of Clinical Analysis, School of Pharmaceutical Sciences, São Paulo State University, Araraquara, SP, Brazil.

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Abstract

Tityus serrulatus scorpion is responsible for a significant number of envenomings in Brazil, ranging from mild to severe, and in some cases, leading to fatalities. While supportive care is the primary treatment modality, moderate and severe cases require antivenom administration despite potential limitations and adverse effects. The remarkable proliferation of T. serrulatus scorpions, attributed to their biology and asexual reproduction, contributes to a high incidence of envenomation. T. serrulatus scorpion venom predominantly consists of short proteins acting as neurotoxins (a and β), that primarily target ion channels. Nevertheless, high molecular weight compounds, including metalloproteases, serine proteases, phospholipases, and hyaluronidases, are also present in the venom. These compounds play a crucial role in envenomation, influencing the severity of symptoms and the spread of venom. This review endeavors to comprehensively understand the *T. serrulatus* scorpion venom by elucidating the primary high molecular weight compounds and exploring their potential contributions to envenomation. Understanding these compounds' mechanisms of action can aid in developing more effective treatments and prevention strategies, ultimately mitigating the impact of scorpion envenomation on public health in Brazil.

* **Correspondence:** ecabraga@fcfrp.usp.br

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Background

Scorpion stings represent a major public health challenge across the globe, with Brazil being one of the most severely impacted countries [1]. Despite the relatively low lethality rate of scorpionism in Brazil, the number of incidents in the country has risen dramatically over the past decade. Indeed, the number of scorpion sting incidents has increased by over 200% in the last ten years (Figure 1), from approximately 80,000 incidents in 2013 to more than 180,000 in 2022 [2]. The prevalence of scorpions in urban areas, coupled with factors such as deforestation and urbanization, has led to a surge in human-scorpion interactions. These interactions, often resulting in stings, have raised significant public health concerns across the country. The rise in scorpionism has prompted local authorities and healthcare providers to bolster their efforts in terms of prevention, treatment, and education to better address this growing issue and ensure the safety and well-being of the Brazilian population [3,4]. This trend is a cause for concern and underscores the need for effective prevention and treatment strategies to address this growing public health issue. Among the various scorpion species found in Brazil, those belonging to the Tityus genus are of medical importance. The Tityus serrulatus scorpion is responsible for the most severe cases of envenomation and fatalities [5,6], especially in areas of human population densities [7]. This scorpion's venom is a complex mixture of various molecules, including low molecular weight peptides such as neurotoxins and high molecular weight proteins such as enzymes [6,8]. While several studies have explored the toxic and mechanistic effects of neurotoxins, there is a lack of understanding regarding the role of high molecular weight proteins in the pathogenesis of *T. serrulatus* envenoming.

This review is dedicated to offering a comprehensive insight into the *T. serrulatus* scorpion, shedding light on the intricate array of molecules present in its venom. Our primary focus lies on the high molecular weight proteins, recognized for their significant



Figure 1. Trends in scorpionism in Brazil over the past decade. The left y-axis represents the number of scorpion sting incidents, while the right y-axis represents the lethality rate (%), calculated by the equation *Lethality rate* (%) = $\frac{number of deaths}{number of cases} \times 100$. The years 2020, 2021, and 2022 are still subject to review [2].

involvement in the pathogenesis of envenomation. It is crucial to clarify that, in this context, we define high molecular weight proteins as those exceeding 14 kDa, constituting approximately 20-25% of the venom composition [9]. Furthermore, this review will delve into the prominent high molecular weight proteins within *T. serrulatus* venom, underscoring the imperative need for further research to fully harness their potential applications.

Tityus serrulatus envenomation and treatment

T. serrulatus sting can lead to a wide range of clinical manifestations, varying from mild to severe, and can even result in death in some cases. Local symptoms such as pain, edema, erythema, sudoresis, and paresthesia are among the most commonly reported. These symptoms usually appear within hours of the sting and can last for several days. In addition to local symptoms, systemic manifestations can also occur. Tachycardia, diaphoresis, profuse sweating, psychomotor agitation, tremors, nausea, vomiting, sialorrhea, arterial hypertension, or hypotension are some systemic symptoms observed after *T. serrulatus* envenoming [3,5,6]. In severe systemic manifestations, other clinical manifestations may occur, including acute pulmonary edema, cardiovascular collapse, cardiac arrhythmia, congestive heart failure, and shock. In addition to clinical evaluation, complementary imaging, and biochemical tests are important for monitoring cases through an electrocardiogram, chest X-ray, echocardiogram, and biochemical tests to assess elevated creatine phosphokinase (CPK), and its MB fraction, hyperglycemia, hyperamylasemia, hypokalemia, and hyponatremia [10,11]. These symptoms have the potential to be life-threatening and necessitate prompt medical attention. Generally, the severity of T. serrulatus envenoming depends on the amount of venom injected, the time between the sting and medical intervention, and the individual's age and health status. Indeed, children under six and, less frequently, the elderly with comorbidities are more seriously affected and are related to most deaths [2,12].

A study was conducted on children under the age of 15 who experienced severe symptoms after being stung by *T. serrulatus* and were subsequently admitted to the intensive care unit. The study found that the most common symptoms reported by these children were tachycardia, sweating, and agitation. Furthermore, abnormal liver function tests were observed, with significant increases in aspartate aminotransferase (AST) and alanine aminotransferase (ALT) levels. There was also a high incidence of pulmonary edema, which in rare cases progressed to respiratory failure and even death. These findings underscore the importance of promptly recognizing and aggressively managing severe *T. serrulatus* envenomation in children, especially those with abnormal liver function tests and pulmonary edema [13].

The *T. serrulatus* envenoming treatment is primarily supportive, and early administration of analgesics, antihistamines, and benzodiazepines can help alleviate symptoms and prevent complications [14]. In mild cases, characterized only by local signs and symptoms, antivenom usage is not recommended, only in symptomatic treatment, and observation of the clinical condition for at least 6 hours after the incident is advised [15]. In moderate and severe cases, Brazil has two different antivenoms available: the arachnid antivenom (each vial with 5 mL contains a fraction of heterologous F(ab'), immunoglobulins that neutralize a minimum of 15.0 minimum lethal dose (MLD) of Loxosceles gaucho venom, 1.5 MLD of Phoneutria nigriventer venom, and 1.5 MLD of T. serrulatus venom per mL [16]) and the scorpion antivenom (each vial with 5 mL contains a fraction of heterologous F(ab'), immunoglobulins that neutralize a minimum of 5.0 mg of T. serrulatus reference venom [17]). For moderate cases, patients with signs of intense local pain associated with some manifestations are considered, thus, two to three vials of antivenom are administered. In severe cases, with the presence of more intense and severe local and systemic signs related to the respiratory and cardiovascular systems, four to six vials of antivenom are recommended [15].

Additional treatments may also be necessary, such as vasodilators, anti-arrhythmic agents, and inotropes. Therefore, healthcare professionals must thoroughly understand the clinical presentation and management of *T. serrulatus* envenoming to ensure optimal patient outcomes [6]. It is crucial to note that the use of antivenom should be based on clinical criteria, such as the severity of envenomation, rather than solely on the confirmation of a scorpion sting. Although antivenom is considered the mainstay of treatment for moderate and severe *T. serrulatus* envenoming, it is essential to understand that the use of heterologous antivenom has some limitations and can lead

to adverse effects [18]. As such, a careful risk-benefit assessment should be made before administering antivenom.

Tityus serrulatus biology

Popularly known as the yellow scorpion, *T. serrulatus* epitomizes a highly specialized species adapted to tropical and subtropical Brazilian ecosystems. Belonging to the arachnid class within the subphylum Chelicerata, scorpions possess four pairs of appendages distributed along their segmented body, comprising the prosoma (cephalothorax) and the opisthosoma (abdomen and tail). *T. serrulatus* exhibits well-developed chelicerae and pedipalps in the anterior cephalothorax, pivotal in facilitating the feeding process. In the terminal section of the opisthosoma, referred to as the telson, the venom-secreting glands are housing the stinger, a specialized apparatus responsible for venom delivery. Additionally, *T. serrulatus* showcases a distinctive anatomical feature in the tail (Figure 2), characterized by diminutive tooth-like structures or serrations, which have warranted the species' designation of "serrulatus" [6,19,20].

Parthenogenesis, a form of asexual reproduction, emerges as a pivotal factor propelling the proliferation of *T. serrulatus*. Within this process, eggs undergo development without the need for fertilization, a relatively uncommon phenomenon in nature, albeit observed in select scorpion species. Despite reports of male *T. serrulatus* individuals, the extent of sexual reproduction in this species remains incompletely elucidated, as the preponderance of females strongly suggests a propensity towards parthenogenetic reproduction as the primary reproductive mode [20,22].



Figure 2. *Tityus serrulatus* scorpion. *Tityus serrulatus*, commonly measuring between 7-9 centimeters (approximately 2.75-3.5 inches) in length, is characterized by its brown to dark brown color. The species name "serrulatus" is derived from the Portuguese term "serrilha", which refers to the serrated feature in its tail, indicated by a red circle in the image, setting it apart as a distinctive anatomical hallmark [21].

Similar to other scorpion species, *T. serrulatus* showcases remarkable resilience during prolonged periods of food deprivation, with reports documenting individuals surviving up to 400 days without sustenance. Nevertheless, this endurance does not extend to the absence of water access, which emerges as a critical determinant for the species' sustenance and survival [23].

Consequently, the synergistic combination of asexual reproduction and resistance to starvation contributes to the rapid expansion of *T. serrulatus* populations, thereby extending their habitat range and heightening the potential for human encounters and associated incidents [20].

Neurotoxicity triggered by low molecular weight compounds

T. serrulatus venom is a highly intricate combination of various compounds. It serves as a valuable repository of small neurotoxic proteins (refer to Table 1), playing crucial roles in prey capture, defense against predators [24,25], and interacting with diverse ionic channels in excitable membranes, contributing to their biological effects [26].

Voltage-gated Na⁺ channel toxins are the primary and highly reactive components responsible for the toxic effects of scorpion envenoming. These toxins are long-chain peptides and can be categorized into two classes: α - and β -scorpion neurotoxins [41,42]. The α -toxins specifically bind to site three, located on extracellular loops S3-S4 of domain IV of the ion channel. This binding hinders or even blocks the inactivation mechanism of these channels, resulting in their prolonged activation [43]. On the other hand, β -toxins bind to site four of the channel, immobilizing it and keeping it in the activated position [44,45]. The α and β -toxins, such as Ts1-5, Ts17, Ts18, Ts26-28, and Ts30, have a specific affinity for Na⁺ channels, thereby modulating the activated channels. Some of these toxins, like Ts5, may also interfere with the permeability of K⁺ channels [46].

K⁺ channel neurotoxins, including Ts6-9, Ts11, Ts12, Ts15, Ts16, and Ts19-25, exhibit inhibitory or blocking effects on K⁺ channels [26,47]. Specifically, Ts11-13, which were described by Pimenta et al. [48], are 29 amino-acid peptide sequences that contain four disulfide bridges. Another noteworthy toxin is Ts32, as reported by De Oliveira et al. [49]. Ts32 is a cell-penetrating peptide and represents the only Ca²⁺-specific toxin identified in *T. serrulatus* venom thus far. This particular toxin is capable of increasing intracellular Ca²⁺ release and holds promising biotechnological potential for the treatment of cancer cells [40,49].

Verano-Braga et al. [50] employed a proteomic approach to identify a novel group of peptides within *T. serrulatus* venom, referred to as hypotensins. These peptides are characterized by their random-coiled linear structure and possess a similar amino acid signature to bradykinin-potentiating peptides. The study revealed that hypotensins exhibit hypotensive effects and induce endothelium-dependent vasorelaxation, which is mediated by the release of nitric oxide (NO) [50].

Short-chain toxins found in *T. serrulatus* venom consist of 30-32 amino acid residues, primarily held together by three disulfide bridges, and this family of peptides constitutes a significant group that primarily targets K^+ channels [26]. These toxins exhibit diverse biological activities, including but not limited

Table 1. Small neurotoxins found on *Tityus serrulatus* scorpion venom that target ion channels.

Toxin	Target	Mechanism of action	Ref.
Ts1	Nav	Shifts the voltage of activation toward more negative potentials	[27]
Ts2	Nav	Inhibit the inactivation of the activated channels, blocking neuronal transmission; induces macrophage activation and production of immune mediators	[27,28]
Ts3	Nav	Inhibit inactivation of the activated channels, blocking neuronal transmission	[29]
Ts4	Nav	Induces release of neurotransmitters glutamic acid and gamma-aminobutyric acid; induces allergic reaction	[30]
Ts5	Nav and Kv	Inhibiting inactivation and blocking neuronal transmission; increases potassium permeability	[31]
Ts6	Kv	Block channels; induces macrophage activation and production of immune mediators	[28,32]
Ts7	Kv	Blocks multiple voltage-gated potassium channel subtypes; blocks Kv1.3 channel by occluding the pore	[32]
Ts8	Kv	Blocks potassium channels; inhibits Kv4.2 channel and produces nociception in vivo	[33]
Ts9	Kv	Blocks small-conductance calcium-activated potassium channels	[34]
Ts11	Kv	Blocks potassium channels at different proportions	[35]
Ts12	Kv	Blocks potassium channels	[35]
Ts15	Kv	Preferentially blocks Kv1.2, Kv1.3 and Kv2.1	[36,37]
Ts17	Nav	Change the kinetics of Nav1.2 and Nav1.5	[38]
Ts19 frag-II	Kv	Block Kv1.2	[39]
Ts32	Cav	Increases intracellular Ca ²⁺ release	[40]

to bradykinin-potentiating effects, antimicrobial properties, hemolytic activity, hypotensive effects (hypotensins), immunemodulating capabilities, and hormone-like activities [51]. Their wide range of biological activities highlights their versatility and potential for various applications.

Regarding the omic analysis of *T. serrulatus* venom, there have been two notable reports involving transcriptomic and proteomic analyses. The more recent study, conducted by De Oliveira et al. [49], identified new peptides capable of modulating ion channels. This analysis shed light on previously unknown components of the venom.

Additionally, *T. serrulatus* venom has been found to contain various low molecular weight components, which include antimicrobial peptides, hypotensins (previously mentioned), C-type natriuretic peptides, and non-disulfide peptides with angiotensin-converting enzyme inhibitor activity [52]. These findings demonstrate the diverse range of bioactive molecules present in *T. serrulatus* venom and their potential for various therapeutic applications.

According to the transcriptomic analysis conducted by Kalapothakis et al. [52], the most abundant toxin types in *T. serrulatus* venom are Na⁺ and K⁺ channel toxins, accounting for 45.24% and 38.10% of the total toxins, respectively. In addition, nine novel putative toxin sequences (Ts33-Ts41) were identified, being that, Ts33-35, Ts37, and Ts38 possess the conserved Toxin_3 domain, suggesting their potential action on Na⁺ channels. Ts36, Ts39, and Ts40 do not exhibit this domain, but show similarities, respectively, to toxins such as JAW07013.1 from *T. serrulatus*, AGT39262.1 from *Mesobuthus eupeus*, and ADY39581.1 from *Hottentotta judaicus*, respectively [52].

High molecular weight compounds and how they could interfere in the envenoming

Several studies have been conducted to investigate the venom of *T. serrulatus*, using transcriptomes and proteomics techniques. These studies have significantly contributed to our understanding of the venom's composition. Most proteins present in the *T*.



Relative front

Figure 3. Tricine-SDS-PAGE and densitometry of *Tityus serulatus* scorpion venom (TsV). (A) Tricine-SDS-PAGE profiles. Lanes 1 and 2: non-reduced TsV, 30 and 100 µg, respectively; Lanes 3 and 4: molecular weight markers (MW); Lanes 5 and 6: reduced TsV, 30 and 100 µg, respectively. (B-E) Densitometry of the bands obtained from Tricine-SDS-PAGE. (B-C) Densitometry showing the molecular weight of the standards. (D-E) Densitometry showing the percentage of the non-reduced TsV bands, respectively. Tricine-SDS-PAGE was performed as described by Schägger and von Jagow [53]. The densitometry analysis was performed using a densitometer Image Lab[™] Software.

serrulatus venom are neurotoxins with action on ion channels and molecular weights lower than 14 kDa. However, the venom also has many enzymes and other components with molecular weights higher than 14 kDa (~20-25%, Figure 3 A-E), still little characterized. Therefore, this work highlights the main venom compounds with molecular weight higher than 14 kDa.

Notably, Alvarenga et al. [54] identified various high molecular weight components using transcriptomic analysis, including antarease, zinc metalloproteases, proteins rich in cysteine, hyaluronidase, and phospholipase A_2 (PLA₂). Similarly, De Oliveira et al. [49] identified metalloproteinase, hyaluronidase, cysteine-rich secretory protein (CRISP), PLA₂, phospholipase C (PLC), and phospholipase D (PLD) through their research. Additionally, Amorim et al. [8] studies revealed the presence of metalloproteinases, CRISPs, phospholipases, and phospholiesterases (PDE).

Recently, Kalapothakis et al. [52] reported a novel transcriptomic approach that unveiled the presence of new components in the venom of *T. serrulatus*, including chitinase, peptidyl- α hydroxyglycine α -amidating lyase (PAL), peptidyl-glycine α -amidating monooxygenase A (PAM), and peptidylglycine α -hydroxylating monooxygenase (PHM). These findings shed light on the diverse array of bioactive molecules present in *T. serrulatus* venom.

Metalloproteases

Regarding proteases, metalloproteases are the most commonly present in animal venoms [55] and need a cofactor to perform the proteolytic activity, such as bivalent ions [56,57]. Some studies have identified the presence of metalloproteases in the venom of *T. serrulatus*. The study by Fletcher et al. [58] characterized a new metalloproteinase called antarease, which cleaves vesicle-associated membrane protein 2 (VAMP2) close to the transmembrane domain. VAMP2 is a protein that, along with the synaptosome-associated protein of 25 kDa (SNAP25) and Syntaxin, is essential for the release of a variety of biologically active molecules via exocytosis [59]. Zornetta et al. [60] produced a recombinant antarease from T. serrulatus venom and observed that it caused paralysis of the neuromuscular junction of insects and mammals, and they also indicated that this enzyme could act in voltage-gated calcium channel, inactivating it. Venancio et al. [61] identified dynorphin-cleaving metalloproteinases that may be antarease-like molecules.

The action of metalloproteases may be related to the acute pancreatitis that occurs in scorpion stings [6], which has already been reported. Machado and Silveira-Filho [62] showed hemorrhagic pancreatitis caused by the *T. serrulatus* toxin in dogs, while Novaes et al. [63] observed acute pancreatitis in rats after the injection of *T. serrulatus* toxin. Gallagher, Sankaran, and Williams [64] demonstrated that the scorpion venom indirectly prompted the release of amylase by acting on nerve endings to release neurotransmitters.

Carmo et al. [55] identified ten proteases named metalloserrulases (TsMs), which showed similarities (from 46

to 95%) with the antarease sequence. These TsMs have a zincbinding site and a conserved methionine, a common structure in the metzincin family, except for TsMs 10, which presents a great similarity with gluzincins and M13 metalloprotease families [55]. Metzincins are related to proteases A Disintegrin and Metalloprotease (ADAM) family, which in snakes are directly involved with the envenoming and blood clotting process [65]. Gluzincins are angiotensin-converting enzyme-like, and are involved in biological processes related to the conversion of angiotensin I into II [49,66]. Additionally, Carmo et al. [55] also reported that these proteases are involved in the maturation process of other toxins present in the venom, cleaving near arginine and lysine residues, which is also demonstrated by Martin-Eauclaire et al. [67] concerning the median lethal dose (LD₅₀) of different venom toxins.

The number of putative components in the transcriptome of *T. serrulatus* representing metalloproteases is considerable (~30%) [49,54] and in the proteome as well (~20%) [8]. However, a significantly larger amount of venom is required to detect any proteolytic activity, compared to snake venoms, for example [55].

Although metalloproteases share the same phylogenetic origin [49,55], Figure 4 illustrates a comparison among various metalloproteases, including antarease, antareaselike, metalloprotease, and metalloserrulases. While there are similarities between some of them, only one amino acid residue is common to all of these proteases, with few displaying any significant similarity.

Serine proteases

Although gangrene, hemolysis, and necrosis are infrequently documented in human envenomation cases caused by *T. serrulatus*, these manifestations can occur in animals, indicating the presence of proteolytic enzymes within the venom [69]. Almeida et al. [70] identified enzymes that provided gelatinolytic activities *in vitro*, which are potentially serine proteases, because they were inhibited by phenylmethylsulphonyl fluoride (PMSF), a serine protease inhibitor, and their optimal pH was eight, the same for serine proteases [71]. Amorim et al. [8] also detected serine protease activity using the Fraction I from *T. serrulatus*. It is important to emphasize that this component was only identified in venom transcriptomics and proteomics [49].

Hyaluronidases

Hyaluronidases are enzymes able to degrade hyaluronic acid, the major component of the extracellular [72], and are involved in several physiological and pathological activities such as fertilization, wound healing, embryogenesis, angiogenesis, diffusion of toxins and drugs, metastasis, pneumonia, sepsis, bacteremia, meningitis, inflammation, allergy, and others [73]. Being present in many animal venoms [73] and widely identified in scorpions [61,74,75], their major role is the facilitation of venom spread in the victim's tissues [76].

Hyaluronidase was isolated from *T. serrulatus* venom by Pessini et al. [77], which can confirm the spreading effect

AUAISSQNS6	MFLSLSSLLFVTIVSAVPTGREDIVIPTIETSRSGERRVRFRALGQDVDLRLESA	SNIFSEDFG	SLYEGEEAR-RLPSSVI	TILKNKLEKDNEKKAALTIDENDDGTSIEGVIGAKMKILPEQTVKNGKRAHKVFEVEEKEDNHNIKLNDEIIPPGVKGENGSTIKKTREIVPPGIKGRLNVAESSTERQGQQ I	190
AUAISSQNTS	MIVILAIALFIDAVSAVFTGREDVATENVETERSGVRRIKFRALDQNIDLRLESA	ANLISDDFE	SLIEGDGNRIRKPVDI	REKKRMTRNRETSARFIIDEDG-PLTINGIINSKERIEPIESEAMIRUGVRAHRIIEIR-RDENTIRENQIIPPGIRKVE	170
A0A076L332	MFVCVISLLFCASVSAIPNGRREVTYPSLETLRSGIKIVKFRAFGEDIELKLEPAG	SDVISDNFT	IMKDNL/G-RI-QTTDE	(SLKSRLPKAREKGAALYINEDG-FLKIKGVINSKLSIEPYNSDKMVKHGISAHVITESFAERKH-FNDKVMSMNLKKTFAKDNARIFNEDQ 1	101
A0A1S5QN52	MFVCVISLLFCASVSAIPNGRREVTYPSLETLRSGIKVVKFRAFGEDIELKLEPAG	GDVISDDFA	ALLDNYG-NS-QKTDVI	<pre>KSLKNKLFKDSEKGAVLYINDDG-ILEINGIINSKLRIEPNKSEETVNEGIKSHLIIELSAEKKR-FTDTVFRMNINENEKFYE-SVRDFDESE</pre>	166
A0A1S5QN54	MFTFMISLLFCASVLAISIGQTEVIYPSLETLASGMKRMKFRAFDQDMELNLEPAG	SDVISDNFT	FIIYSEG-KT-QATDV	<pre>{SLKKRLFRDPEKGSALYISEKG-LTEIEGVINSKLRIEPYETREVTKNEFKAHQIITTFPEDKH-FSDAMINFNMEKEKSYSERNDTNVNGDE</pre>	167
A0A1S5QN57	MLACLTILLFCTSVLAVSNGRIEVIYPSLETSRFGIKIIKFRAFDQDIILKLEPAG	DVISDDFT	TVMDEEG-KI-EPIDV	SFKGKLFRDQEKGAALYIDENE-FIEIEGIINFKLRIEPSESQKVGKYGMKEHVITETFDEKKR-FKDTVNNLNSQRSFLEDMGREFDENQ	167
A0A1S5QN59				ALYIDENG-FIKIKGIINSELRIEPSESEEVGKYGMKEHVIFETFDEKKH-FRDIVYNLNLQRSFSKNKAMNLSKDQ	75
A0A1S5QN60				ALYIDENG-FIKIKGIINSELRIEPSESEEVGKYGMKEHVIFETFDEKKH-FRDAVNNLTFQAPFSKDKSV-LNEDQ	- 74
A0A1S50N58	MICLVNILLFASVSTISNGRVDVVFPSVETSKSGMKTVKFRAFNODVELKMKPAG	GEILAKNFA	AFFDENG-OVPHPIDV	2NLNRKLFKNSEORAALIIDEDG-PLTIEGILNRKVRITPFESRKIIKNGIIAHRVLEEISEEKSYLHNNVTFTNIEREAENMKRMARVDO	167
A0A076L316	MELOEKTTMYFASEELFATVSA I PNGRVHVVEPTVETSRSGEKTLKLRAFDRDTELNLOPA	ETLAKNEG	SIMDDDH-BLEHPVDV	KNLORKLYKNSANGAALLIDEDG-PUTIEGNINAKUBIAPYESGGMDEAGBIAHWIEEEERDEKAYLRDDSTYFFNLWOEL-PPDIDTENTKRMDREGK	1.81
\$0\$076T \$¥7	WYEA CEFT FTTUCA TONODY/HUT/FDAVETCDC/2PKTI KTDA FODDTETNI ODA	PTI APARY	TMNENG_DI PUT VDVI		165
AGAD / CLAV /	WETKER DITTORTERDITION	PTIAKDES	PENDENI-REFILLVOV	NEIGORDERNOMMENDEDDEDG TVITEGEMEMENDERNE DEGEMENDEGENENGTE DEGEMENDE – KOTENDERNE – VDE – TOPENDERNE – VDE – TOPENDERNE – VDE – TOPENDERNE – VDE	122
AURISSUN //	VKIIKERALDRUVELINVKEA	SET DAKDEG	SPRIDERIN-QUIRFIDVI	WINSKEIRNORGAV DE DEDG-EVITEGTEDKEDTAPEDERNVDGGTARQIVELIGEE-KSILMUVERENG-UNK-	100
AUAU / 6L339	MILMUITPEATVSVIPSERIDVVPPEVETSRSGVKTIKERALDUDVELNLEPAG	BILKKSPG	32 VGENG-KVIHPIDV	INLKSKEPRNSSEGAALLIDEDG-QUTIEGRINSNERIAPHESGRMDEGGRIAHQIVEEIHEETSPEHIUVEPNNAEREVESKRMSKNOE I	100
A0A076L725	MYIVYIFLFAAVSAIPNSRNDIVFPTVETSRSGVKTIKFRALGQDVELNLEPAG	SQILGEREV	/FVGENG-QLYHPVDVI	(NLRSKLYRNSAKGAALLIDEEE-PLTIEGVVNEKLRIAPLESRRMDEDGRIAHQIVEEINEEKLPLHYDMIQMNNER-ELEREVESIKTLATDDQ 1	170
A0A076L882	MYIVYIFLFAAVSAIPNSRNDVVFPKVETSRSGVKTIKFRALGQDVELNLEPAG	SQILGEKFG	SFVDENR-QIYHPVNV	<pre>(NLRSKLYRNSAKGAALLIDEDE-PLTIEGVVNEKLRIAPHESGRMDEEGRIAHQIVEEINEEKLPLHYDMLQMDTGR-ELEREVESIKSMARDDQ]</pre>	164
P86392	MISYLASIFLLATVSAVPSGRVEVVFPSVETSRSGVKTVKFTALDQDVELKLRSAG	GEILGKRFA	AIQDVDVI	SLRRKIYRDSVNGAALLIDEDG-PLTIEGIVNSKLRIQPFESGRITKDGIIAHQIVEVIDDKKSYDRVAVIPENVKRNAENVSRMARDDD	160
V9Z9A3				DD	2
A0A076LAV6	MQTMIFSLAYIILLATVSAIPSGRVDIVFPSLETSRSGVKIIKFEALGQDIELNLEPAG	GEILAKDFA	AIVDLNN-QREHLTNVI	2DLKRKIYRDSVKGAALLIDENG-PLTMQGIINSKLRIVPYESGRVIKDGRIAHQIVELINDEKSYIN-DVMPLDVNVMGVMENVVKISKKSP 3	170
P85842					2
A0A076L3T0	DLFASVSAIPNGREDVVFPWVETSRSGAKTVKFRALGEDTELKLEPA	DILAFGEA	LEDSKN-OTKSSVDV	NURKETYRDSVNGAALITDDDE-PLSTOGTVNSKLETAPHESBELNOVRBRAHETVELTNDKNSSLEDDVISBN	1.60
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AUA1S5QN56	CIIFVVKVLIVTEYYFSNSFSSNA-EYEKYLAITIVYTQTLIDTMNFKIEIKLNAFLKYTK	SOPSFIER	CSTLPNYPQYFDCEAL.	.DNMCKFYSTG-SVALANNAHVIKMVSLRPMGDLVY-GKLDLRTLGIAFVGGICEIKYKCGVSIDDPNDFHEFINTFAHELAHLLGCPHDEDP-PVTYIKNSPGSLDCKWSYGHI 3	389
A0A1S50NT5	CILVELVFLIESNLTYRYKNNDETILKECYIIVTVMNSLAQSLKLNLTIQLQNVIKFTE	NEPSFIAN	SVIPEYPHILDSELL	(ANMGTHYRYK-RDVSNGDIVLLLMDRKMGVKSL-TSY-SLQLGLGYVGAACESRYKFGIAVYDS-KLDVFYDTCVHECAHVMGSPHDGDP-PVSHISNSPGSIDCLWKYGHI	361
A0A076L332	CVSIKYLFLTDSNFRSGFPNPKDMETYFATMFILVQEDMDTLKLNIKVSLIGIEPVK	NETNFVKE	ESLIPGE-EVFDFGHV	JGNLNVLNCKYKDNELYKKADSIMFITKRLLGNREPDGSVSTNTLGVANLGGACNFCLKTGVIKDYG-DMTLLANTVAHETAHQIGSPHDGED-APYSLPGSFTGEKCPGSQGYL	359
A0A1S5QN52	CVVIEYFCVADSTFSKLFNNDEKEIQTFVAGMFVKAQEMINTLKLNIKLVLIGLTVLK	DDLPFIRE	ESLIPGIDTIFNVGHI	4QNISKNYCHEKANNFVVQADLIFLITRRAIGEPRPDGTVFDGIMGVANLGGTCNPCTKYGVIHAEKENMLSAANSLAHESAHLLGSPHDGEG-ADLSLDGSPTGIKCPGKEGYI 3	361
A0A1S5QN54	CVEIEYLFLAENNFAAKFLDSEKLQVYLATVFVQVQTLMDTLNLNIKLRLIGTVIYK	DNPPFFQG	GSLIPGQ-DVLNAGNI	/KNMSNFLCKNEENELMKQADLVMLITINQIGYSSPSGMLFPSVIGVSYLGGACDPCKKCALIKDDG-RAFSTAYLIAHESAHLIGSPHDGAG-PIFSLPGSPTAVECPPFSGFI 7	359
A0A1S5QN57	CVEIEYYFLGEKNFTEGFPDLNKFKIYIGTVFVQTQTMMDTLKLKIKIRALGVLMLK	SDPPFITK	KSLFVGE-DVFNMELI	AKNMETYLCENKGIEKFQRADIIMLLTGRQIGIFDSKGTILVGVLGVANLGRACAQCTKLAVVKCDD-DKRETAVTLAHESAHLLGSPHDGEG-DYFSLPGSPGSTKCVASDGFI	359
A0A1S50N59	CVETEVVFLADKKFRDEVRNLDVFKKNIGSTVTOVONLLDTLNLNIKIKTLATLIFV	VDPPFLKD	OSLIPCE-EMENVAVI	PNPGKYTCEKKETGVIARADIILELSBREIGYMDRDGRIYKDILGASHLGGACOECKKVGFILWRD-DMHYTAVTVAHESAHLIGSPHDSEGGVAOSYPGSEGSEDCPLEDGFT	268
A0A1950N60	CVETDELEMMERKVINGEDGI EDVVGNLGI LETEVONTI DELNI KMKTVVI GEVNLV-	COUPETOR	CLIDCK-DIENVDAV		267
a0a1950N58	SUUTEVECUAESNETEHEKKD===EHITEVLTDMVTCUODILETIDICIKURIICUOAEKK	NDDEVIER	SATECVENTIONERT.	TRANSVERVEL A TOTA KENDET MET TOTA TO A SUBJECT TO A SUBJECT TO A SUBJECT TO A SUBJECT AND A SUBJECT AND A SUBJECT AND A SUBJECT AS A SUBJECT AND A SUBJECT AS A	361
RORISSYNDS	STATELLCARESNELERERKDERITELLIKATIGAODEDELDEGIKAKHEGAOARKK	MOPPILE	SOMIFOIENIBORRE.		201
AUAU/6L316	CIVIEILSVTDSNFTKRFSNIEELSEYITLTYAGVQTIMDTLEDGIKLKLIGIHAFTN	TEPPFIES	SNAIPGHERIVEIKRV.	INSMERTICEH-DIGLARDADIIIIITNDELLAGWDGGPNINTNVAGVAIHSGACDQCSEVAVSEHSS-SIFIEVIIIDHETAHSIGIDHDGGGRSENCSNGRGGF	300
AUAU/6LAV/	CIVIEILSVTDRDFTKRFSTYDDLTKYVSRTYLGVENIIERLELGIKVRLLGIQAFTN	TEPPFIED	DSAIDNHÖLAPNHÖLAPUNALKT	SSMEDYICEH-AYGLAKDADIIMLTTVRLLAEWDGS-KINTNIAGAARYSSVCNQCYKVGVSMHFS-YFTDRIEVIAHETAHLIGVPHDGSGPVTVAPNENPGAHNCSSYEGYF 3	359
A0A1S50N77	CIVIEFLSVIDSNVTKLFKTDEDLAKLMAETSIGVQNIIDTLDMGIKVRLLGIQTFTN	TDPSYIED	OSAIPKYERYLDHRRL	MMMGTYYCEH-ATGLAKSADIIMLITGRPLAAKLK-SGINANVGGVASPSDVCKQCHKVGAARFYQ-EFDARVNVIAHETAHLIGVPHDGEGAKSIYLFESPGALNCPYKSGYF	327
A0A076L339	CIVLEILSVIERTLTERFVPEEALTKHMTHTYMGVQNKIDTLELGIKVRLVGIEAFTK	TEPSFFEE	ETVIPGHDNYFSYLDI	/SKSKDYYCKH-DEGLAKDADIIMLSTERSLGTLGSDGEIETSVGGVASDSAVCDQCYKIGVAEHYN-DYSYRTNIITHEAAHLIGVPHDGGRGNSDSPGALDCPSEDSYI 3	357
A0A076L7Z5	CIVIEILSVTDKLVTKRFATDEALTQHMTLTYVKVQNIFDTLELGIKVRLIGIEAYTN	TEPSFIED	OSAIPGHEKYLHFVKL	.RNLGNYYCKQ-NEGLAKDADIIMLTTDRPLADISSEGKLNTNIGGVANYASVCHPCYKVGVGVYYS-YSYARVEVLAHEAAHLIGIPHDGEGEYYGMLGAKNCSVKYGYF	361
A0A076L882	CIVIEFLSVTDOLVTKRFETVEALTEHMTLTYLGVONILDMLELGIKVRLIGIEAYTN	TEPSFIED	OSAIPGHEKYLHYVKL	IRNLENYYCKR-NEGLAKDADIIMLTTDRPLASTPSEGKLNTNIGGAANYAAVCSOCSKVGVGVYYS-YSYARVETIAHEAAHLIGIPHDGEGGVYGIPGAKNCSSKYGYF	355
P86392	CIVVEYYIVTDSAFTKRFKSNSALTNYVTVMFTGVONLMDTLELGIGVRLLGVTTFTE	TEPSFIKD	ONLIPGPPAAFDPDVL	ISAMSKYYCNH-OTGLAKDTDLIFLITARGMGDPREDGTVDINTAGIANSAGVCRPCFKSGIATDDS-DYNERVDTLAHESVHLLGSPHDGEGPNLVSLEGSPGAANCPAKAGYI	355
107033	CTURPYY TUPDON PTYPER ONON TRAVENER OTONI MOST FLOTOURI I CUTTERED	TPDOPTVD	NIT TROPPA APOPDUT		107
1000767 506	CITATELLA COMPAREMENTED DE MAN ANTI AL DEL MARTINE CONTRACTOR DE LA CONTRACTÓR DE LA CONTRACT	NEDCEIRD	AND FOUL ON THE OF DESIGN OF THE OF T		265
ACAC / BLAVE	CITIDIDE VIETE IERCENINRELEBITIVE IGV VIETE IERCENING	WPL SE TPP	SAIFGRQQVLDFVDL	INNAARIICNA-ARGBARDADIINDISWARDGBLQDDGIVAINIAGISLGSGVCAQCSAVGVDDS-DINEAVDIVARBIABIIGAFIDEGFEQIGGSGFGARDCFESDGII	305
P85842	CITIDYLCVTETCFCERFRTNRELLEYITVMFTGVQNLLDTLNLGI			VGVAQDYS-DYNERVDTVAHETAHLIGAPHDEEGPCQDT	8.9
A0A076L310	CIVVESLCVTESRFTRRFKTNQALTEYVTLMFTGAQNMYDTMNLGIKLRLIGVQAFAI	TEPSFIKT	PNEVKNGK-YLK-FSI	.DDMNSYYCKN-ATGLAKDADIIILIITRTMVLMKGS-QIENEAVGLALGGGACLTCEKSLVMADET-DYNERTISLAHEAAHLLAVPHDGDDFKVSDIPNSPGAKSCRYGEGYI 3	352
A0A1S5QN67	CIVIEFLCATESKFTERFKSNQALTEYVILMYAGAEILIRGLDLGIKFRLTGIQAYTK	NEPAFIKE	ENELANGS-MINGNNV	(RSFGNYVCKN-LTGLSKKADIAMLIVTRTMTQKKPS-GVA-NAAGLAYYGKVCDECHKVGATVDKS-RGLNTITTLAHEIAHLLGVPHDGESIPAVPGSPGAESCSPKEGYI 3	335
				.::.:*	
			Identity (%)		
A0A1S5QN56	MSYESNAVNGSKFSPCSAACAKHLLDLTKCVKVPCPV	423	100.0		
A0A1S5QNT5	MSYKYNIHNSTLFSSCSRNNIAHLCLTIKCLQCK	395	42.05		
A0A076L332	MGDSKGENKGKFSPCTRENVKFFLNKDEASCVVSACKSTV	399	32.47		
A0A1S50N52	MGNDYGENERKESPCSKANIMYFLGKPRASCIVGAC	397	31.70		
3031550N54	MGNTFGVTACMECKCCBENIKVFI.CKNFASCIT_SCNCTTI_	399	32 30		
NON1 GEONET		400	24 54		
AGALGOUND /	I CONEC ENERGY DESCRIPTION OF THE PROPERTY INTERPORT	200	27.05		
NONTRONNOA	DODNEGENEDRESTUTIKNVKIFLSQNBASULIINCKNRVL	309	21.95		
AUA1S5QN60	LGDNSGANKGKFSVCSQENVKYFLGQPQASCLLQPCENLVL	308	28.72		
A0A1S5QN58	MGDLSDRVHKYRFSECSKSCLRHLLSLPRANCLFEVCKNCLQ	403	31.38		
A0A076L316	MGRSDSSLHPTFQNFRDYRDEAFSECSKENAKYFLSTSKADCLFEECPND-NITSLFET	424	28.72		
A0A076LAV7	MGLRWGENANSFSECSKKCAAYLLSLPKADCLYEECONDDNITSLFEI	407	30.93		
A0A1S5QN77	MGNKNLYKYKFSECSKACAKYLLSLSRADCLFEECNKYP	366	32.58		
A0A076L339	MGDWGKNHDKFSECSKICAKYLLSLPKANCVYEOCKTDY	396	33.51		
3030761775	MCNACKNUTVECCCSKANAEVIISIERARCVIEQCRIDI	402	34.70		
3030761992	MOVE	105	32.16		
AUAU / 01002	HORDGRINNI RESECONNCHEILLSEPANDULIEDUEVEWIEIFK-	105	33.10		
P86392	MUNENDKNKYKFSPCTKKCVEYLLSKPTASCIFQQCTDF	394	34.81		
V929A3	MGNRNDKNKYKFSPCTKKCVEYLLSKPTASCIFQQCRD	235	31.74		
A0A076LAV6	MGSGNNKVNKFKFSKCTKKCVEHLLSLPRASCVLADC	402	38.21		

Figure 4. Multiple align among metalloproteases from *Tityus serulatus* scorpion venom. Align among antarease (P86392), antarease-like (V9Z9A3), metalloprotease (P85842) metalloserrulase 2 (A0A076LAV6), 3 (A0A076L310), 4 (A0A076L332), 5 (A0A076L7Z5), 6 (A0A076L882), 7 (A0A076LAV7), 8 (A0A076L316), 9 (A0A076L339), 11 (A0A1S5QN60), 12 (A0A1S5QN59), 13 (A0A1S5QN77), 14 (A0A1S5QN54), 15 (A0A1S5QN58), 16 (A0A1S5QN57), 17 (A0A1S5QN55), 18 (A0A1S5QN52), 19 (A0A1S5QN56) and 20 (A0A1S5QN67). Red ID represents data from UniProtKB/TrEMBL. The purple box represents the signal peptide. Amino acid residues are indicated in black. *: fully conserved residues; :: residues with very similar properties; .: residues with dissimilar properties [68].

34.52

393

performed by this enzyme. Furthermore, the presence of this component may be related to the lethality of the venom. Horta et al. [78] produced an anti-hyaluronidase antibody from *T. serrulatus*, which inhibited the enzyme's action both *in vitro* and *in vivo*, effectively reducing the venom's toxicity. The same antibody was used by Oliveira-Mendes et al. [79], who demonstrated that hyaluronidase not only played a crucial role in venom spreading but also inhibiting it, resulting in a delay in venom biodistribution from the bloodstream to target organs (*e.g.*, lungs and liver), being this inhibitor a potential and a valuable first-aid agent for this type of envenoming.

-PNMLKFSKCSIACAKYFLSLPQASCLYEDCPSSAY

The structures of hyaluronidases are already deposited in the UniProtKB database [68], demonstrating that the six deposited sequences show high identity among them (> 79%) (Figure 5A). Additionally, the molecular model of hyaluronidases exhibits secondary structures, such as α -helix and β -sheets (Figure 5B-C), indicating the presence of different epitopes distributed throughout the molecule's structure, as demonstrated by Horta et al. [78]. In this study, the authors performed a systematic mapping of continuous epitopes which were recognized by anti-hyaluronidase serum with three antigenic regions common to

both hyaluronidases TsHyal-1 and TsHyal-2 and could identify among these regions, the active site D¹⁰¹ and E¹⁰³. Also, the three antigenic regions were mapped onto the 3D models of both hyaluronidases and were found to surround the active sites, which could indicate that the neutralization of Ts venom by anti-hyaluronidase serum was a result of the binding of serum antibodies to specific residues in the Ts hyaluronidase active site [78].

Phospholipases

Phospholipases are enzymes that hydrolyze steric bonds of phospholipids, that could infer in the membrane function and structure [82]. They can be involved in phospholipid metabolism, signal transduction, or other cellular functions, or extracellular when they are present in mammalian pancreatic juice and animal venom and act as platelet aggregators in the blood or as catalysts in the release of arachidonic acid, triggering inflammatory reactions [83].

Although PLA_2 activity was not detected in fraction I of *T. serrulatus* venom, Amorim et al. [8] detected phospholipases in the venom proteome. In addition, De Oliveira et al. [49]

1

4	P85841 A0A218QWX6 A0A218QX64 A0A7S8RGE3 A0A218QX67 W0HFN9 A0A7S8MU79	DFKVYWEVPSPLCSKRFKINVTEVLTSHEILVN PISIFSIVISVICAVQADFKVYWEVPSPLCSKRFKINVTEVLTSHEILVN MNPISIFSIVISVICAVQADFKVYWEVPSPLCSKRFKINVTEVLTSHEILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSIVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYWEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVISVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVVISVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVISVISVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVISVISVISVICAVQADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVISVISVISVICAVADFKVYMEVPSPLCSKRFNINVTQVLTSHKILVN MNPISIFSVISVISVISVISVISVISVISVISVISVISVISVISVIS	QGESFNG QGESFNG QGESFNG QGESFNG QGESFNG QGESFNG QGESFNG	DKIVIFYENQLGKYPHIDSNNVEINGGILQVADLAKHLKVAKDNITKFVPNPNFNGVGV DKIVIFYENQLGKYPHIDSNNVEINGGILQVADLAKHLKVAKDNITKFVPNPNFNGVGV DKIVIFYENQLGKYPHIDSNNVEINGGILQVADLAKHLKVAKDNITKFVPNPNFNGVGV DKIVMFYENQLGKYPYINSNNVEINGGILQVADLLKHLKVAKDNITKLVPNPNFNGVGV DKIVMFYENQLGKYPYIDSNNVEINGGILQVADLLKHLKVAKDNITKLVPNPNFNGVGV DKIVMFYENQLGKYPYIDSNKVEINGGILQVADLLKHLKVAKDNITKLVPNPNFNGVGV DKIVMFYENQLGKYPYIDSNKVEINGGILQVADLLKHLKVAKDNITKLVPNPNFNGVGV	100 116 118 118 118 118 118					
	P85841 A0A218QWX6 A0A218QX64 A0A7S8RGE3 A0A218QX67 W0HFN9 A0A7S8MU79	IDWEAWRPSWEPNWGKLKVYKEKSIDLVKSKHPEWPSDRVEKVAKEEWEESA IDWEAWRPSWEPNWGKLKVYKEKSIDLVKSKHPEWPSDRVEKVAKEEWEESA IDWEAWRPSWEPNWGKLKVYKEKSIDLVKSKHPEWPSDRVEKVAKEEWEESA IDWESWLPSWEPNWGKWYYRKSIDLVKSKHPEWPSBRVEKVAKEEWEKSA IDWESWLPTWDPNNDKMKVYRKKSIDLVKSKHPEWPSBRVENVAKEEWEKSA IDWESWLPTWDPNNDKMKYYRKSIDLVKSKHPEWPSBRVENVAKEEWEKSA IDWESWLPTWDPNNCKKYYRKSIDLVKSKHPEWPSBRVENVAKEEWEKSA	KEWMVKI KEWMVKI KEWMVKI KEWMVKI KEWMVKI KEWMVKI	LKLAQEMRPNAVWCYYLFPDCYNYFGKDQPSQFSCSSRIQKENSRLSWLWNQSTAICLS LKLAQEMRPNAVWCYYLFPDCYNYFGKDQPSQFSCSSRIQKENSRLSWLWNQSTAICLS LKLAQEMRPNAVWCYYLFPDCYNYFGKDQPSQFSCSSRIQKENSRLSWLWNQSTAICLS LKLAQEMRPNAVWCYYLFPDCINYGGRAQPSRKTCNKLUQLENDRIFNUKQSTAICPS LKLAQELRPNAVWCYYSFPDCYNYSRKDEPSPLACIRKVLVENDRISWLWKQSTAICPS LKLAQELRPNAVWCYYSFPDCYNYSRKDEPSPLACIRKVLVENDRISWLWKQSTAICPS LKLAQELRPNAVWCYYSFPDCYNYSRKDEPSPLACIRKVLVENDRISWLWKQSTAICPS	218 234 236 236 236 236 236 236					
	P85841 A0A218QWX6 A0A218QX64 A0A7S8RGE3 A0A218QX67 W0HFN9 A0A7S8MU79	IYIQESHVTKYNMSQRTWWIDARLREAIRVSEHRPNIPIYPYINYILPGTNQTVPAMDFKRTLGQIASLGLDGALLWGSSYHVLTESQCKITSDYVKSVIAPTVATVVLNTNRCSQII IYIQESHITKYNMSQRTWWIDARLREAIRVSEHRPNIPIYPYINYILPGTNQTVPAMDFKRTLGQIASLGLDGALLWGSSYHVLTESQCKITSDYVKSVIAPTVATVVLNTNRCSQII IYIQESHITKYNMSQRTWWIDARLREAIRVSEHRPNIPIYPYINYILPGTNQTVPAMDFKRTLGQIASLGLDGALLWGSSYHVLTESQCKITSDYVKSVIAPTVATVVLNTNRCSQII IYIQESHITKYSMSQRVWWIDARLREAVRLSMTRRLIPIYPYINYILPGTNQTVPAMDFKRTLGQIASLGLDGALLWGSSYHLLSESQCKITSDYVKSVIAPTVATVVLNTNRCSQII IHIQESHITKYSMSQRVWWIDARLREAVRLSMYHRNIPIYPYINYILPGTNQIVPVMDFKRTLGQIASLGLEGVILWGSSYHLFESQCKITFDYVKNVIAPTVATVVLNTNRCSQLI IHIQESHITKYSMSQRVWWIDARLREAVRLSMYHRNIPIYPYINYILPGTNQIVPVMDFKRTLGQIASLGLEGVILWGSSYHLFESSQCKITFDYVKNVIAPTVATVVLNTNRCSQLI IHIQESHITKYSMSQRVWWIDARLREAVRLSMYHRNIPIYPYINYILPGTNQIVPVMDFKRTLGQIASLGLEGVILWGSSYHLFESSQCKITFDYVKNVIAPTVATVVLNTNRCSQLI IHIQESHITKYSMSQRVWWIDARLREAVRLSMYHRNIPIYPYINYILPGTNQIVPVMDFKRTLGQIASLGLEGAILWGSSYHLFSESQCKITFDYVKNVIAPTVATVVLNTNRCSQLI								
	P85841 A0A218QWX6 A0A218QX64 A0A7S8RGE3 A0A218QX67 W0HFN9 A0A7S8MU79	CKGRGNCVWPEEPFSSWKYLVDPKMPVFKPTNIHCKCKGYLGRYCEIPK- CKGRGNCVWPEEPFSSWKYLVDPKMPVFKPTNIHCKCKGYLGRYCEIPK- CKGRGNCVWPEPFSSWKYLVDPKMPVFKPTNIHCKCKGYLGRYCEIPK- CKGRGNCFWYEPSLSWKYLVDPKMPVFNPTNIYCRCNGYMGRYCGFNYY CKGRGNCIWPAEPFSSWKYLLDPKMPVFKPMKIICKCKHYLGRYCQIPK-	385 401 403 404 336 403 403	<pre>dentity (%) 100.0 99.74 99.74 79.74 82.39 82.86 82.86</pre>						
3		\square		C						



A0A218QX64, A0A7S8RGE3, A0A218QX67 and A0A7S8MU79. Red ID represents data from UniProtKB/Swiss-Prot, while black ID represents data from similar properties; .: residues with dissimilar properties [68]. (B) Front and (C) back view of hyaluronidase-1 (P85841) structure, based on the amino acid sequence using Alphafold [80,81]. a-helix and β -sheet are represented in pink and yellow, respectively.

observed the presence of transcripts of PLA₂, PLC, and PLD, without proteomic evidence.

some articles [61,74,75], with a need for further study on these molecules and their presence in the venom.

Cysteine-rich secretory protein

CRISP was also identified in T. serrulatus venom, and its role is still unclear [8]. However, CRISPs are widely distributed in animal venoms, such as snake venoms, being their role on ion channels also demonstrated [84], and in humans, they are involved with the immune system [85]. The molecular model of CRISP is demonstrated in Figure 6A-B. Despite the presence of these compounds in proteomic and transcriptomic approaches of T. serrulatus venom [8,49], their activity was not detected in

Others

Phosphodiesterases are enzymes that hydrolyze cyclic nucleotides and play a role in regulating intracellular levels of cyclic adenosine monophosphate (cAMP), cyclic guanosine monophosphate (cGMP) and, therefore, cell function [86,87]. Its presence in the venom was detected by proteome [8].

PAL, PAM, and PHM were found in the transcriptome analysis [52], and these enzymes are responsible for post-translational modifications of venom toxins, such as C-terminal amidation,



Figure 6. Structure prediction of CRISP from *Tityus serrulatus* scorpion venom. The structure was predicted through the amino acid sequence of CRISP (A0A218QX58) using Alphafold [80,81]. (A) Front and (B) back view. α -helix and β -sheet are represented in pink and yellow, respectively.

which plays a fundamental role in enhancing their lethal effect [48,88,89].

Chitinases play a crucial role in the digestive process of *T. serrulatus* by being present in its intestinal system [90]. Consequently, identifying these enzymes in the transcriptome could be directly linked to the effective digestion of prey organisms [52].

Identifying antarease, metalloproteases, peptides rich in cysteine, and phospholipases highlights the venom's potential enzymatic activity and its role in disrupting various physiological processes. Moreover, the presence of hyaluronidase suggests a possible involvement in tissue degradation and facilitating venom spread, while CRISP proteins may contribute to modulating the victim's immune response. PDE identification is noteworthy as this enzyme can impact intracellular signaling pathways. Some described components are also involved with the enhancement of the lethality of toxins, as well as the prey's digestion. These studies have significantly improved our understanding of the T. serrulatus venom composition overall by identifying and characterizing several important venom components. Further research building upon these findings can contribute to developing novel therapeutic interventions and enhancing our knowledge of the molecular mechanisms underlying envenomation.

Conclusion

T. serrulatus envenoming is of great medical importance in Brazil, and it can be more severe and frequent in children and patients with comorbidities. Antivenom treatment

is available in healthcare services and recommended for moderate and severe cases. Notably, T. serrulatus scorpion venom is an extraordinary source of proteins with different molecular weights and performing different roles. The high molecular weight components can play crucial roles during the *T. serrulatus* envenomation strategy and have evolved to subdue prey or defend against predators effectively; thus, some considerations can be inferred. Many of these components in T. serrulatus venom are enzymatic proteins and play various functions, such as facilitating the breakdown of tissues, interfering with physiological processes, disrupting the prey's defense mechanisms, increasing the lethality of toxins, or helping the digestion of prey. Enzymes identified in T. serrulatus venom often have larger molecular weights due to their complex structures and functional domains, exhibiting complex tertiary structures, which could provide stability and protection against degradation.

Additionally, these components in *T. serrulatus* venom can participate in intricate interactions with target molecules in the prey or victim's body, may involve binding to specific receptors or interfering in signaling pathways, targeting different physiological systems or employing multiple mechanisms of action simultaneously, increasing their chances of subduing prey or defending themselves effectively, suggesting the enhancement of venom's potency. Although the high molecular weight components were identified in *T. serrulatus* venom, and some of them were isolated, further research into the venom's composition and function can provide deeper insights into the precise roles of these components and their impact on envenomation.

Abbreviations

ADAM: A Disintegrin and Metalloprotease; ALT: alanine aminotransferase; AST: aspartate aminotransferase; cAMP: cyclic adenosine monophosphate; cGMP: cyclic guanosine monophosphate; CPK: creatine phosphokinase; CRISP: cysteine-rich secretory protein; LD_{50} : median lethal dose; MLD: minimum lethal dose; NO: nitric oxide; PAL: peptidyla-hydroxyglycine a-amidating lyase; PAM: peptidyl-glycine a-amidating monooxygenase A; PDE: phosphodiesterase; PHM: peptidylglycine a-hydroxylating monooxygenase; PLA₂: phospholipase A₂; PLC: phospholipase C; PLD: phospholipase D; PMSF: phenylmethylsulphonyl fluoride; SNAP25: synaptosomeassociated protein of 25 kDa; TsMs: metalloserrulases; VAMP2: vesicle associated membrane protein 2.

Availability of data and materials

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Competing interests

The authors declare that they have no competing interests.

Authors' contributions

ISO and ECA conceived the main idea of this work and draft the manuscript. NMAS, IGF, FAC, JAGS, WMM and MBP wrote parts of the review. All authors read and approved the final manuscript.

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