High fat diet has a prominent effect upon the course of chronic schistosomiasis mansoni in mice

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This study investigated whether a long-term high-fat diet has an effect on the outcome of chronic murine schistosomiasis mansoni compared to a standard diet. Swiss Webster female mice (3 weeks old) were fed each diet for up to six months and were then infected with 50 Schistosoma mansoni cercariae. Their nutritional status was assessed by monitoring total serum cholesterol and body mass. Infected mice were examined 6-17 weeks post infection to estimate the number of eggs in faeces. Mice were euthanised the next day. Total serum cholesterol was lower in infected mice in comparison to uninfected controls (p = 0.0035). In contrast, body mass (p = 0.003), liver volume (p = 0.0405), spleen volume (p = 0.0124), lung volume (p = 0.0033) and faecal (p = 0.0064) and tissue egg density (p = 0.0002) were significantly higher for infected mice fed a high-fat diet. From these findings, it is suggested that a high-fat diet has a prominent effect on the course of chronic schistosomiasis mansoni in mice.

Key words: Schistosoma mansoni - cholesterol - chronic phase

The complexity of Schistosoma mansoni growth, migration and development implies that numerous levels of regulation occur as host glucose metabolism and its immune response (Saule et al. 2005, Loverde et al. 2007).

Schistosomes are blood-dwelling flukes that are highly dependent on the host metabolism (Saule et al. 2005). As a result, physiological changes associated with schistosomiasis infection have been considered important factors influencing this host-parasite relationship (Hulstijn et al. 2003). Early studies showed dramatic alterations in adult worm development (Neves et al. 2001, 2002, Oliveira et al. 2003), as well as diminished schistosomal periocular granuloma and milder chronic lesions in undernourished mice compared to mice fed a normal diet (Coutinho et al. 2003).

Animal models have shown reductions in the concentration of lipoproteins in plasma during experimental schistosomiasis mansoni infection (El-Marzouki & Amin 1997, Muller et al. 2001, Ramos et al. 2004). Furthermore, schistosome infection counteracts the effects of an atherogenic diet by modulating host lipid metabolism and inducing a reduction in total blood cholesterol (Doenhoff et al. 2002). Chronic exposure to schistosome eggs promoted the development of a Th2 response and reduced the levels of total cholesterol and LDL in serum, but it failed to reduce aortic lesion development (La Flamme et al. 2007).

As part of a series of studies being carried out in our laboratories on the role of dyslipidaemia during acute schistosomiasis infection, it was observed that a high-fat diet favours the development of adult worms (Neves et al. 2007a). Moreover, egg maturity, faecal output, viability and egg hatching were higher compared to mice fed a standard diet (Neves et al. 2007b). The purpose of this study was to determine whether a high-fat diet could alter the outcome of chronic schistosomiasis.

MATERIALS AND METHODS

The study design reported herein complies with the current laws regarding ethical procedures involving laboratory animals and was approved by Comissão de Ética de Uso de Animais-Fiocruz (L-0036/07), Rio de Janeiro, Brazil.

Animals and housing - Weanling (21 day) Swiss Webster female mice (Centro de Criação de Animais de Laboratório-Fiocruz) were housed conventionally in polypropylene cages (40 x 33 cm) with stainless steel-screened covers. Mice were kept in a temperature (21 ± 1°C) and humidity-controlled (60 ± 10%) room and exposed to a 12 h light/dark cycle with artificial lights and to an air exhaustion cycle.

Diet - Experimental animals were fed their specific diets from weaning at the age of 21 days. Hypercholesterolaemia was induced by feeding high-fat chow (HFC) composed of pork lard, egg yolks, wheat flour, corn starch, casein and balanced with vitamins and minerals (47% carbohydrates, 24% proteins, 29% lipids) (5.7 kcal/g body wt/day) (Neves et al. 2006) for more than a period of six months after weaning. Control groups were fed standard lab chow (SC) for mice (Nuvilab CR-1-NUVITAL Nutrients Ltda., Colombo, Paraná, Brazil)

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containing 12% fat, 28% protein and 60% carbohydrate (4.6 kcal/g body wt/day). Animals were weighed twice a week throughout the experiment. All animals were given free access to water and food during the study.

**Experimental infection protocol** - After six months of feeding on their respective diets, 20 mice were each subcutaneously infected with ~50 *S. mansoni* (BH strain) cercariae recovered by shedding from laboratory-reared *Biomphalaria glabrata* snails (Martinez et al. 2003). The *S. mansoni* life cycle has been maintained at the Laboratório de Malacologia (Instituto Oswaldo Cruz-Fiocruz), using *B. glabrata* snails and mice for the past 40 years.

A total of 30 mice were separated into four groups: (SC) (n = 5), uninfected mice fed HFC (n = 5), infected mice fed standard chow (ISC) (n = 10) and infected mice fed (HFC) (n = 10).

**Biochemical assay** - After the 6-month feeding period, total serum cholesterol was assayed using the colorimetric enzymatic method as previously described (Neves et al. 2006). Briefly, plasma was obtained from blood samples collected from food-deprived mice (overnight) by puncture of the retro orbital sinus. The biochemical assay was conducted both the day before experimental infection and at necropsy 17 weeks post infection, when mice were euthanised by jugular section (hypovolaemic shock). The blood was centrifuged at 1200 g for 15 min at rt to separate the serum, which was stored at -20°C until use.

**Biometry** - Body mass was evaluated at two points throughout the experiment: weeks zero and 36.

**Parasitologic examination** - Susceptibility to infection was evaluated by quantifying faecal egg excretion 6-17 weeks post-infection. The faeces were analysed in duplicate by the Kato-Katz technique processed in order to determine the number of eggs per gram of stool. The percentage of viable eggs in stool samples was determined by a quantitative sedimentation. The viability of these eggs was determined by flame cell activity and/or movement in miracidium. The prepatent period was determined by when the first faecal eggs were found (Martinez et al. 2003).

**Euthanasia** - Mice were euthanised the day after the last faecal examination was performed. Skin was opened by a midline incision in the thorax and abdomen to remove liver, intestine and spleen. Liver volume, intestine volume (IV) and spleen volume (SV) were determined by the Scherle liquid displacement method (Scherle 1970).

Adult worms were recovered from the portal system and mesenteric veins. The rate of infection (infectivity) was calculated as the percentage of maturation of cercariae into adult worms (Freire et al. 2003). For the enumeration of eggs, the liver and small intestine were removed from freshly killed mice. The organs were digested in 4% potassium hydroxide at 56°C and centrifuged (900 g) for 5 min. For oogram performance, duplicate 100 µL aliquots of the digest were placed on a glass slide and eggs were counted by light microscopy (100X) (Martinez et al. 2003).

**Statistical analysis** - Data were statically analyzed using the SPSS software. The Mann-Whitney and Student’s t-test were used to assess the total cholesterol, body mass index at week zero of experiment, infectivity and faeces eggs total were compared between IHFC and ISC groups. The analysis of variance (One Way Anova) was used to estimate the body mass and organs volume in different weeks of experiment. The Person’s correlation test was used to assess the correlation between the number of eggs passed in faeces and worm burden between IHFC and ISC mice. A p-value ≤ 0.05 was considered significant (Vieira 1991).

**RESULTS**

Three IHFC mice and one ISC mouse died during the experiment.

**Biochemical analysis** - As shown in Table I, mice fed the high-fat diet showed significantly increased total cholesterol levels both before (p = 0.0003) and after (p = 0.001) infection.

**Biometry analysis** - There was a significant difference (p = 0.0003) in the gain of body mass between mice fed each diet for 36 weeks. Mice fed the high-fat diet gained more body mass [HFC (58 ± 9) and IHFC (59 ± 12)] than standard controls [SC (43 ± 2) and ISC (39 ± 4)] (Table II).

Significantly greater liver (3.1 ± 0.6, p = 0.0031) and SVs (0.9 ± 0.7, p = 0.0039) were observed for IHFC the lipid-rich diet (Table II). IV decreased in the HFC group (4.0 ± 1.2, p = 0.0034) when compared to the SC group.

**Parasitologic examination** - Table II shows that the high-fat diet resulted in significantly (p = 0.0001) higher infectivity (22%) than the standard diet (14%). The male/female sex-ratio was 1.1:1 in the IHFC group and 1.2:1 in the ISC group. The mean prepatent period was 49 ± 1 days, irrespective of diet. High-fat feeding resulted in a higher total egg output in the IHFC group (61.056) in comparison to the ISC group (18.144) (p = 0.0064). The densities of the eggs in the intestine (25.236 ± 7.516; 4621 ± 2.942, p = 0.0002) and liver (10.804 ± 2828; 2606 ± 1.330, p = 0.0002) were statistically greater in the IHFC group that in the ISC group. Table II also shows that egg viability was higher in the IHFC group (4 ± 0.9) compared to the ISC group (2 ± 0.8) (p = 0.0276).

The pattern of egg production in the faeces is illustrated in Fig. 1. There was an initial rise in the egg output of the mice fed the high-fat diet by weeks 8-10. During the following counts, an irregular excretion was observed with marked increases (week 13) and decreases (week 14). In the later stages of the infection, there was a trend for high egg excretion. Despite the fact that mice fed a standard diet produced lower egg counts, weekly variations were less pronounced. In contrast to the other group, at the end of the observation period the egg count dropped rapidly (p < 0.05).

The relationship between the number of eggs passed in the faeces and worm burden in both groups was adequately described by a linear regression model. There was no significant correlation (Fig. 2A) in the IHFC group (r = 0.5264, p = 0.2248); however, there was a significant correlation (Fig. 2B) in the ISC group between egg count and worm pairs (r = 0.9943, p = 0.0001).
DISCUSSION

Over the last several years, the effects of metabolic disorders on the pathophysiological features of schistosomiasis have indicated that nutrient deprivation leads to a hostile microenvironment for adult worms (Ferreira & Coutinho 1999, Neves et al. 2002, Simões et al. 2002, Oliveira et al. 2003), as undernourished mice develop milder chronic lesions than normally fed mice (Coutinho et al. 2003). In contrast, a cholesterol-rich environment provides favourable conditions that ensure the reproduc-

### TABLE I

Biochemical and biometrical findings in schistosomiasis-infected SW mice fed high-fat chow or standard chow

<table>
<thead>
<tr>
<th>Parameters</th>
<th>HFC</th>
<th>IHFC</th>
<th>SC</th>
<th>ISC</th>
<th>P values&lt;sup&gt;c&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>TC&lt;sup&gt;a&lt;/sup&gt; (mmol/L)</td>
<td>1.6 ± 0.3</td>
<td>-</td>
<td>1.0 ± 0.2</td>
<td>-</td>
<td>0.0080</td>
</tr>
<tr>
<td>TC&lt;sup&gt;b&lt;/sup&gt; (mmol/L)</td>
<td>1.2 ± 0.1</td>
<td>1.2 ± 0.3</td>
<td>0.7 ± 0.2</td>
<td>0.9 ± 0.2</td>
<td>0.0055</td>
</tr>
<tr>
<td>BM&lt;sup&gt;c&lt;/sup&gt; (g/cm)</td>
<td>14 ± 2.5</td>
<td>-</td>
<td>15 ± 2</td>
<td>-</td>
<td>0.2034</td>
</tr>
<tr>
<td>BM&lt;sup&gt;d&lt;/sup&gt; (g/cm)</td>
<td>58 ± 9</td>
<td>59 ± 12</td>
<td>43 ± 2</td>
<td>39 ± 4</td>
<td>0.0003</td>
</tr>
<tr>
<td>LV&lt;sup&gt;e&lt;/sup&gt; (10&lt;sup&gt;3&lt;/sup&gt; mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>2.3 ± 0.9</td>
<td>3.0 ± 0.5</td>
<td>2.2 ± 0.2</td>
<td>2.4 ± 0.5</td>
<td>0.0405</td>
</tr>
<tr>
<td>IV&lt;sup&gt;f&lt;/sup&gt; (10&lt;sup&gt;3&lt;/sup&gt; mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>4.0 ± 1.1</td>
<td>6.0 ± 1.5</td>
<td>6.7 ± 1.1</td>
<td>5.9 ± 1.0</td>
<td>0.0034</td>
</tr>
<tr>
<td>SV&lt;sup&gt;f&lt;/sup&gt; (10&lt;sup&gt;3&lt;/sup&gt; mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>0.3 ± 0.2</td>
<td>0.8 ± 0.6</td>
<td>0.2 ± 0.1</td>
<td>0.3 ± 0.1</td>
<td>0.0124</td>
</tr>
<tr>
<td>HV&lt;sup&gt;f&lt;/sup&gt; (10&lt;sup&gt;3&lt;/sup&gt; mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>0.2 ± 0.0</td>
<td>0.1 ± 0.04</td>
<td>0.1 ± 0.02</td>
<td>0.1 ± 0.05</td>
<td>0.0039</td>
</tr>
<tr>
<td>LuV&lt;sup&gt;f&lt;/sup&gt; (10&lt;sup&gt;3&lt;/sup&gt; mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>0.2 ± 0.1</td>
<td>0.5 ± 0.2</td>
<td>0.2 ± 0.07</td>
<td>0.3 ± 0.1</td>
<td>0.0033</td>
</tr>
<tr>
<td>KV&lt;sup&gt;f&lt;/sup&gt; (10&lt;sup&gt;3&lt;/sup&gt; mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>0.7 ± 0.2</td>
<td>0.6 ± 0.08</td>
<td>0.4 ± 0.1</td>
<td>0.5 ± 0.1</td>
<td>0.0212</td>
</tr>
</tbody>
</table>

Values are the mean ± standard deviation, used statistics test: Test T (TC<sup>a</sup> and BM<sup>c</sup>) and Anova (other parameters). Significant differences (p < 0.05).<sup>a</sup>: after 41 weeks of experiment; <sup>b</sup>: after 50 weeks of experiment; <sup>c</sup>: when compared with the respective control group; HFC: uninfected mice fed high-fat diet; HV: heart volume; IHFC: infected mice fed high-fat chow diet; ISC: infected mice fed standard chow; IV: intestine volume; KV: kidney volume; LuV: lung volume; LV: liver volume; SC: uninfected mice fed standard chow; SV: spleen volume; TC: total cholesterol serum concentration; ◊: organs volume (Scherle’s method) after 49 weeks of experiment and 17 weeks post infection.

### TABLE II

Parasitologic findings in Swiss mice fed high-fat diet or standard chow, during the course of chronic phase of schistosomiasis

<table>
<thead>
<tr>
<th>Parameters</th>
<th>IHFC (n = 7)</th>
<th>ISC (n = 9)</th>
<th>p values</th>
</tr>
</thead>
<tbody>
<tr>
<td>Infectivity</td>
<td>22%</td>
<td>14%</td>
<td>0.0001</td>
</tr>
<tr>
<td>Faeces eggs (total)</td>
<td>61056</td>
<td>18144</td>
<td>0.0064</td>
</tr>
<tr>
<td>Intestine egg count</td>
<td>25236 ± 7516</td>
<td>4621 ± 2942</td>
<td>0.0002</td>
</tr>
<tr>
<td>Liver egg count</td>
<td>10804 ± 2828</td>
<td>2606 ± 1330</td>
<td>0.0002</td>
</tr>
<tr>
<td>Egg viability</td>
<td>4 ± 0.9</td>
<td>2 ± 0.8</td>
<td>0.0276</td>
</tr>
</tbody>
</table>

Values are the mean ± standard deviation. Mice were euthanised 17 weeks pi. IHFC: infected mice fed high-fat chow diet; ISC: infected mice fed standard chow.

### TABLE III

Effects of a high-fat diet on parasitologic aspects in Swiss mice chronically infected with *Schistosoma mansoni*

<table>
<thead>
<tr>
<th>Parameters</th>
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DISCUSSION

Over the last several years, the effects of metabolic disorders on the pathophysiological features of schistosomiasis have indicated that nutrient deprivation leads to a hostile microenvironment for adult worms (Ferreira & Coutinho 1999, Neves et al. 2002, Simões et al. 2002, Oliveira et al. 2003), as undernourished mice develop milder chronic lesions than normally fed mice (Coutinho et al. 2003). In contrast, a cholesterol-rich environment provides favourable conditions that ensure the reproduc-
The ability to adapt metabolic processes to environmental changes markedly influences the behaviour of schistosomes within the host due to a number of interesting nutritional adaptations (Halton 1997). In the present study, the IHFC group showed higher worm burden, tissue egg, egg viability and egg-laying. These data suggest that schistosomes are closely associated with the modulation of host lipid metabolism (El Ridi et al. 2004, Tallima & El Ridi 2005). The most plausible explanation is the presence of signalling pathways by which parasites detect host signals and respond to them in a way that presumably both increases their survival (Loverde et al. 2007) and reproduction of adult worms (Neves et al. 2007a). In this study, regression and linear correlation between faecal eggs and worm pairs found no correlation in the IHFC group. This suggests that although higher egg elimination does not correlate with worm pairs, another factor must account for the mechanism of egg production. Perhaps serum cholesterol induces higher egg production during egg embryogenesis among females from mice chronically infected and fed a high-fat diet, which is in line with our previous report (Neves et al. 2007a).

Paired schistosomes migrate toward mesenteric veins, where immature eggs are released. A proportion of these eggs is carried out by the portal flow back to the liver, while other reaches the lumen and are voided with faeces to the external environment (Neves et al. 2007a). Two mechanisms seem to favour their excretion to the outside environment. First, eggs incorporate external nutrients and growth factors as development takes place. Second, molecules secreted from eggs (Ashton et al. 2001) induce cytokine production (Brindley 2005) and local granulomatous inflammation, where inflammatory cells sustain egg passage (Lenzi et al. 1987). Moreover, both structural and functional changes are present during chronic murine schistosomiasis (Moreels et al. 2001). In a previous study, it was hypothesised that lipid-rich diets may be environmental modifiers which lead to exchanges between miracidium inside eggs and nutrients, primarily in the small intestine (Neves et al. 2007b).

As in other studies, mice fed a standard diet excreted few eggs by weeks 6-8 and this count then rose to a peak at week 12 and achieved stability between weeks 13-16, declining thereafter (Rocha et al. 1995, El Ridi et al. 2004).
In mice fed a high-fat diet, an initial low elimination was observed between 6–8 weeks, followed by significant increases at weeks 9 and 10, with week-to-week fluctuations from week 11-16, when excretion was increased. The main differences were the absence of stability and a progressive decrease, as previously reported during the chronic phase of schistosomiasis (Cheever et al. 1994, Rocha et al. 1995, Barth et al. 1996). This study complements the existing literature (Neves et al. 2007a, b) indicating that certain dietary patterns may be important for the outcome of chronic schistosomiasis. Altogether, these observations suggest that schistosome infection in mice fed high-fat diets may be a useful tool for understanding the consequences for human populations living in areas where both morbidities exist.

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REFERENCES


