Genotypes of pathogenic Leptospira spp isolated from rodents in Argentina

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Leptospirosis is the most widespread zoonosis in the world and significant efforts have been made to determine and classify pathogenic Leptospira strains. This zoonosis is maintained in nature through chronic renal infections of carrier animals, with rodents and other small mammals serving as the most important reservoirs. Additionally, domestic animals, such as livestock and dogs, are significant sources of human infection. In this study, a multiple-locus variable-number tandem repeat analysis (MLVA) was applied to genotype 22 pathogenic Leptospira strains isolated from urban and periurban rodent populations from different regions of Argentina. Three MLVA profiles were identified in strains belonging to the species Leptospira interrogans (serovars Icterohaemorrhagiae and Canicola); one profile was observed in serovar Icterohaemorrhagiae and two MLVA profiles were observed in isolates of serovars Canicola and Portlandve. All strains belonging to Leptospira borgpeterseni serovar Castellonis exhibited the same MLVA profile. Four different genotypes were isolated from urban populations of rodents, including both mice and rats and two different genotypes were isolated from periurban populations.

Key words: Leptospira spp - pathogenic - multiple-locus variable-number tandem repeat analysis - genotyping - rodent population - Argentina

Leptospirosis is the most widespread zoonotic disease in the world. It is caused by pathogenic Leptospira spp spirochetes, which find an ideal niche for transmission in subtropical and tropical regions. There are no available data on the incidence of leptospirosis in several countries and the disease is frequently not recognised and is therefore severely neglected (Hartskeerl et al. 2011). Pathogenic Leptospira strains mainly infect mammals, but they can also be found in reptiles and amphibians (Levett 2001, Adler & Moctezuma 2010). This zoonosis is maintained in nature through chronic renal infection of carrier animals, with rodents and other small mammals being the most important reservoirs. Additionally, livestock and domestic animals, such as dogs, are significant sources of human infection. In Argentina, this disease is endemic and no epidemiological control or prevention programs targeting leptospirosis have been implemented thus far. From 2012 until July of 2013, an estimated 1,056 human cases were reported to the National Health Ministry, 99 of which were confirmed as leptospirosis (MS 2013). Recently, Draghi et al. (2011) reported the presence of leptospirosis in cattle from Corrientes province, which is endemic for this disease. An outbreak in this province in 2009 resulted in the death of 100 calves in one week, demonstrating the significance of leptospirosis in this region.

The occurrence of leptospirosis has also been reported in the rodent populations of urban and periurban areas of Argentina (Cacchione et al. 1967, Cacchione 1973, Arango et al. 2001, Martin et al. 2002, Marder et al. 2008, Scialfa et al. 2010), but no information on the molecular characteristics of the Leptospira strains infecting these animals is available.

Several schemes for the genotyping of the pathogenic species of Leptospira spp have been developed around the world in recent years (Cerqueira & Picardeau 2009, Cerqueira et al. 2009, Levett & Haake 2010, Nalam et al. 2010). Multiple-locus variable-number tandem repeat analysis (MLVA) has been successfully employed to genotype pathogenic Leptospira spp strains (Majed et al. 2005, Slack et al. 2005, 2006, Salaün et al. 2006, Pavan et al. 2008, 2011a, Li et al. 2012). Using this methodology, Pavan et al. (2011b) identified eight distinct MLVA genotypes in the Leptospira spp isolated from domestic animals and humans from Argentina.

The present study applied MLVA to molecularly characterise a collection of strains isolated from urban and periurban rodents, including Rattus norvegicus, Rattus rattus, Rattus sp. and Mus musculus, from the Argentine provinces of Buenos Aires, Santa Fe and Entre Ríos.
**MATERIALS AND METHODS**

*Strains* - Twenty-two pathogenic *Leptospira* spp strains were genotyped. Six of these strains were isolated during the years 2009-2011 and 16 of the strains were part of the bacterial collection of the Leptospirosis Laboratory at the Pathobiology Institute of the Argentine National Institute of Agricultural Technology, which is a World Organization for Animal Health reference laboratory. The isolates used in this study were previously serotyped using the cross agglutinin absorption test (Pavan et al. 2008). The sources, dates and the locations of isolation for the strain are listed in Tables I, II. The reference *Leptospira interrogans* strains employed in this study were Pomona (serovar Pomona, serogroup Pomona), M20 (serovar Copenhageni, serogroup Icterohaemorrhagiae), RGA and Ictero No. 1 (serovar Icterohaemorrhagiae, serogroup Icterohaemorrhagiae), Hond Utrecht IV (serovar Canicola, serogroup Canicola) and MY 1039 (serovar Portlandvere, serogroup Canicola). The reference strain used for *Leptospira borgpetersenii* was Castellon3 (serovar Castellonis, serogroup Ballum).

*Genotyping* - The reference strains and isolates were grown in Fletcher medium (Difco Laboratories) at 28°C. The MLVA strain typing procedure was performed as described by Majed et al. (2005), Salaün et al. (2006) and Pavan et al. (2008). Briefly, 100 µL of a culture sample was incubated at 100°C for 10 min and this suspension was used as the DNA template. MLVA typing was performed using two sets of oligonucleotides specific for pathogenic *L. interrogans, Leptospira kirschneri* and *L. borgpetersenii*. Oligonucleotides that hybridised to the flanking regions of the VNTR4, VNTR7, VNTR9, VNTR10, VNTR19, VNTR23 and VNTR31 *loci* were used to discriminate strains of *L. interrogans* and oligonucleotides that hybridised to the flanking regions of the VNTR4, VNTR7, VNTR10, Lb4 and Lb5 *loci* were used for *L. kirschneri, L. borgpetersenii* and *L. interrogans* strains (Majed et al. 2005, Salaün et al. 2006). The final volume (50 µL) of each reaction mixture contained polymerase chain reaction (PCR) buffer (20 mM Tris-HCl, pH 8.4, 50 mM KCl), 200 µM deoxynucleoside triphosphates, 2 µM each of the corresponding forward and reverse primers, 2 mM MgCl₂, 1.25 U of Taq DNA polymerase (Invitrogen) and 5 µL of DNA template. PCR amplifications were carried out in a Thermo Scientific PxE 0.2 Thermal Cycler, using the following cycling parameters: 94°C for 5 min, followed by 35 cycles of denaturation at 94°C for 30 s, annealing at 55°C for 30 s and extension at 72°C for 90 s, with a final cycle at 72°C for 10 min. The amplified samples were examined following electrophoresis in ethidium bromide-containing 2% agarose gels in TAE buffer (40 mM Tris-acetate, 1 mM EDTA, pH 8.0) at 100 V for 50 min. Amplified DNA bands were visualised through ultraviolet light expo-

<table>
<thead>
<tr>
<th>MLVA genotype (copy number of VNTR4, VNTR7, VNTR9, VNTR10, VNTR19, VNTR23, VNTR31 <em>loci</em>)</th>
<th>Isolates (n)</th>
<th>Strain</th>
<th>Source</th>
<th>Province and date of isolation</th>
<th>Genotype reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>“Ictero” (2,1,13,7,2,0,3)</td>
<td>8</td>
<td>Entre Ríos I</td>
<td>Human</td>
<td>Entre Ríos, 1981</td>
<td>Pavan et al. (2011b)</td>
</tr>
<tr>
<td>Reconquista II</td>
<td>River</td>
<td>Buenos Aires, 1996</td>
<td>Pavan et al. (2011b)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cañuelas III</td>
<td>Pig</td>
<td>Buenos Aires, 1996</td>
<td>Pavan et al. (2011b)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>K4</td>
<td><em>Rattus sp.</em></td>
<td>Buenos Aires, 1961 (P)</td>
<td>This paper</td>
<td></td>
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</tr>
<tr>
<td>K5</td>
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<td>Buenos Aires, 1963 (P)</td>
<td>This paper</td>
<td></td>
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<tr>
<td>Rat2 to Rat4</td>
<td><em>Rattus sp.</em></td>
<td>Buenos Aires City, 2010 (U)</td>
<td>This paper</td>
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<td></td>
</tr>
<tr>
<td>Hond Utrecht IV (1,10,2,3,10,2,3)</td>
<td>5</td>
<td>M.4</td>
<td>Human</td>
<td>Unkown, 2004</td>
<td>Pavan et al. (2011b)</td>
</tr>
<tr>
<td>M.5</td>
<td>Cow</td>
<td>Buenos Aires, 2004</td>
<td>Pavan et al. (2011b)</td>
<td></td>
<td></td>
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<tr>
<td>Comadreja</td>
<td>Opossum</td>
<td>Buenos Aires, 2005</td>
<td>Pavan et al. (2011b)</td>
<td></td>
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<tr>
<td>E3</td>
<td>Human</td>
<td>Buenos Aires, 2005</td>
<td>Pavan et al. (2011b)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Roedor 104</td>
<td><em>Rattus norvegicus</em></td>
<td>Buenos Aires City, 2011 (U)</td>
<td>This paper</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MY 1039 (1,10,2,3,10,2,4)</td>
<td>3</td>
<td>Corrientes 266</td>
<td>Cow</td>
<td>Corrientes, 1985</td>
<td>Pavan et al. (2011b)</td>
</tr>
<tr>
<td>Corrientes 289</td>
<td>Cow</td>
<td>Corrientes, 1985</td>
<td>Pavan et al. (2011b)</td>
<td></td>
<td></td>
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<tr>
<td>Roedor 1507</td>
<td><em>R. norvegicus</em></td>
<td>Buenos Aires City, 2011 (U)</td>
<td>This paper</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>16</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

strains isolated from periurban and urban rodent populations are marked with (P) and (U), respectively. MLVA: multiple-*locus* variable-number tandem repeat analysis.
sure (Uvi Tec transliluminator BTS-20.M, Manufacturer UviTec, St. John’s Innovation Centre, Cowley Road, Cambridge, England). Amplicon sizes were estimated using CienMarker (Biodynamics) and the GelAnalyzer 2010a program. To calculate repeat copy numbers, the following formula was used: number of repeats (bp) = [fragment size (bp) - flanking regions (bp)]/ repeat size (bp). Repeat copy numbers were rounded down to the closest whole number. If the copy number was less than one, it was rounded to zero.

RESULTS

The MLVA profiles of 22 pathogenic strains isolated from periurban and urban rodent populations were analysed in this study based on seven loci described by Majed et al. (2005) and five loci described by Salaün et al. (2006) (Tables I, II). All of the strains analysed in this study were also characterised using agglutination absorption tests, which generated the same results as the MLVA typing. A total of seven strains belonged to *L. interrogans*. Five strains presented an MLVA profile (2,1,3,7,2,0,3) that was indistinguishable from that of strains M20 (serovar Copenhageni) and Ictero No. 1 (serovar Icterohaemorrhagiae) (Majed et al. 2005), both of which come from serogroup Icterohaemorrhagiae. Two of these strains were isolated from periurban rodent specimens from Buenos Aires province (K4 and K5) and three were isolated from urban rodents (*Rattus* sp. in the city of Buenos Aires (Rat2, Rat3 and Rat4)). Additionally, two strains were isolated from urban rodents (*R. norvegicus*). One of these strains was characterised as serovar Portlandvere strain (Roedor 1507) which showed the same MLVA profile (1,10,2,3,10,2,4) as MY 1039. The other was characterised as serovar Canicola (Roedor 104) and showed the same profile as Hond Utrecht IV (1,10,2,3,10,2,3). Additionally, we identified 15 strains belonging to *L. borgpetersenii*, all of which presented the MLVA pattern (1,1,4,6) of Castellon 3 (serovar Castellonis). Six of these strains were isolated from periurban rodents, including one strain (C16) from *M. musculus* in Buenos Aires province and five (ROE1-ROE5) from *R. rattus* in Entre Ríos province. The other nine isolates came from urban rodent populations, with a total of eight strains being isolated from *M. musculus* in the city of Santa Fe (Santa Fe I to Santa Fe VIII) and one strain (Rat 1) being isolated from *Rattus* sp. in city of Buenos Aires. The sources, locations and dates of collection for the isolated strains examined in this study are indicated in Tables I, II.

In this study, we used both sets of MLVA typing primers described by Majed et al. (2005) and Salaün et al. (2006) to genotype all three pathogenic *Leptospira* species found in Argentina. No *L. kirschneri* genotypes were found in the rodent isolates obtained in this work. The newly isolated strains were included in serogroup Icterohaemorrhagiae and serogroup Canicola. Five new strains isolated from *Rattus* sp. (K4, K5, Rat2, Rat3 Rat4) were added to serogroup Icterohaemorrhagiae, represented by the reference strains M20, RGA and Ictero I. This MLVA pattern had previously been found in strains isolated from pigs (Cañuelas III) and humans (Entre Ríos I) and in an isolate from a water sample from a river (Reconquista II); all of these samples were isolated in the provinces of Buenos Aires and Entre Ríos (Pavan et al. 2011b). In addition to the serogroup Canicola strains present in Argentina, Portlandvere (MY 1039) has been found in strains isolated from cows (Corrientes 289 and Corrientes 266, from Corrientes province) and rodents (Roedor 1507, isolated from *R. norvegicus* in the city of Buenos Aires). The *Leptospira* spp strains from Buenos Aires province that corresponded to the MLVA profile of Hond Utrecht IV were represented by strains isolated from a white-eared opossum (*Didelphis albiventris*, opossum), a rodent (*R. norvegicus*, Roedor 104), cows (M5) and humans (E3 and M4). Serovar Castellonis (Castellon 3) was more homogeneous and included only isolates from rodents found in the urban and periurban areas of the Santa Fe, Entre Ríos and Buenos Aires provinces, confirming that the strains harbouring this genotype were circulating pathogenic strains.

### TABLE II

Strains of *Leptospira borgpetersenii* typed in this study using the five loci described by Salaün et al. (2006)

<table>
<thead>
<tr>
<th>MLVA genotype (copy number of VNTR4, VNTR7, VNTR10, Lb4 and Lb5 loci)</th>
<th>Isolates (n)</th>
<th>Strain</th>
<th>Source</th>
<th>Province and date of isolation</th>
<th>Genotype reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Castellon3 (1,0,1,4,6)</td>
<td>1</td>
<td>C16</td>
<td><em>M. musculus</em></td>
<td>Buenos Aires, year unknown (P)</td>
<td>This paper</td>
</tr>
<tr>
<td>8</td>
<td>Santa Fe I to Santa Fe VIII</td>
<td>1039</td>
<td><em>M. musculus</em></td>
<td>Santa Fe City, 1998 (U)</td>
<td>This paper</td>
</tr>
<tr>
<td>5</td>
<td>ROE1 to ROE5</td>
<td>Rattus rattus</td>
<td>Paraná, 2006 (P)</td>
<td>This paper</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Rat1</td>
<td><em>Rattus</em> sp.</td>
<td>Buenos Aires City, 2010 (U)</td>
<td>This paper</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>15</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

strains isolated from periurban and urban rodent populations are marked with (P) and (U), respectively. MLVA: multiple-locus variable-number tandem repeat analysis.
**DISCUSSION**

Different rodent species have been described as reservoirs of distinct *Leptospira* spp serovars (Levett 2001, Vanasco et al. 2003, Levett & Haake 2010, Vedhagiri et al. 2010, Hartskerel et al. 2011). Rats have been found to carry the Icterohaemorrhagiae or Copenhageni serovars, while leptospires isolated from mice have been shown to belong to serogroup Ballum. Hartskerel et al. (2011) proposed that the main reservoirs of pathogenic and highly virulent serovars are rodents and this is the reason why leptospires (and leptospirosis) are unlikely to be eradicated.

In contrast to previous reports (Zuerner & Alt 2009), this study showed that tandem repeat analysis (using VNTRs and MLVA) was unable to achieve a higher resolution (stratification) than the serotyping of isolates from Argentina. We genotyped a strain isolated from *Rattus* sp. in the city of Buenos Aires that presented an MLVA profile identical to that of Canicola Portlandvare MY 1039. The MY 1039 MLVA profile had previously only been reported in cows from Corrientes province (Pavan et al. 2011b). New wildlife carriers can be determined through strain isolation and molecular typing methods. Using this approach, Brihguea et al. (2007) isolated and genotyped a *L. interrogans* Canicola Canicola Hond Utrecht IV strain from *D. albiventris* (white-eared opossum) in a rural area in Buenos Aires province. This genotype has also been isolated from strains from rats (this work), cows and humans (Pavan et al. 2011b). On the other hand, the white-eared opossum (*D. albiventris*) seems to be a candidate for the environmental dissemination of *L. borgpetersenii* (Jorge et al. 2012). In this study, 142 *R. norvegicus* and a number of opossums from 10 domiciles of patients with leptospirosis were captured and examined. The infecting serovar in the rat population was Copenhageni and rats could be identified as an important leptospirosis reservoir. *L. borgpetersenii* Castellonis was the pathogenic leptospire isolated from *D. albiventris*. In a recent study in New Caledonia (Pacific Ocean), Perez et al. (2011) obtained similar results for strains isolated from rats and mice. Using a multilocus sequence typing protocol, *L. borgpetersenii* Ballum was identified in isolates from and *L. interrogans* Icterohaemorrhagiae was identified in isolates from *R. rattus* and *R. norvegicus* (Perez et al. 2011). Moreover, an isolate recovered from *R. norvegicus* field rats in South India was identified as *L. borgpetersenii* serogroup Javanica (Vedhagiri et al. 2010). Previous studies performed in humans during a leptospirosis outbreak in 1998 in Santa Fe, Argentina, which occurred after excessive rain and floods, determined that human sera were reactive to the serotypes Ballum, Canicola, Icterohaemorrhagiae and Pyrogenes (Vanasco et al. 2000, 2003). Additionally, *L. borgpetersenii* strains isolated from rodents were serotyped as serovar Arborea serogroup Ballum (Vanasco et al. 2000).

In summary, 15 pathogenic *L. interrogans* and *L. borgpetersenii* strains were isolated from urban rodent populations (from Santa Fe and the city of Buenos Aires), which displayed a total of four different MLVA genotypes. Additionally, seven pathogenic strains were isolated from periurban rodent populations from Buenos Aires and Entre Ríos provinces and these strains contained both pathogenic species as well as two different genotypes. These results show that several MLVA genotypes have circulated among Argentine rodents in the last 50 years and the same genotype is sometimes shared by strains isolated from urban and periurban rodent populations.

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**REFERENCES**


