Seroepidemiological Studies of Cutaneous Leishmaniasis in the Campeche State of Mexico

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Seroepidemiological studies of cutaneous leishmaniasis were carried out in 169 individuals in a rural area of the Campeche state of México. Fifty showed cutaneous lesions suggestive of leishmaniasis, 70% were parasite positive and 96% skin test positive. An overall 40% positivity to skin test with Montenegro’s antigen was found. Most of the affected individuals were males from 11 to 30 years-old. Antibodies were determined by immunofluorescent antibody test (IFA) and by Western blot. Two antigen preparations were used, one from a Leishmania mexicana strain which produced localized cutaneous leishmaniasis (LCL) and the other from a diffuse cutaneous leishmaniasis (DCL). In the general population from the area of study 19% gave positive IFA tests with DCL antigen and 20% with LCL antigen while for the patients 67% gave positive IFA tests with DCL and 71% with LCL. By Western blot analysis most of the patients recognized more antigens in the DCL than in the LCL strain. In the DCL strain 78% of patients recognized a 105 kDa, 34% a 139 kDa, 28% a 117 kDa and 26% a 205 kDa MW antigen. In the LCL strain 40% of patients recognized a 205 kDa and 22% a 175 kDa antigens.

Key words: seroepidemiology - cutaneous leishmaniasis - Campeche state - Mexico - Western blot

American cutaneous leishmaniasis is caused by species belonging to two complexes: Leishmania mexicana and L. braziliensis. Species of L. mexicana produce either self-healing ulcerative cutaneous lesions (localized cutaneous leishmaniasis: LCL) with early positive skin test and low titers of serum antibodies or nodular diffuse cutaneous lesions (diffuse cutaneous leishmaniasis: DCL) with very heavy loads of parasites in macrophages, with negative skin test, higher serum antibodies titers and which do not respond to treatment.

In Mexico, cutaneous leishmaniasis (CL) was recorded for the first time in Campeche state by Seidelin (1912) who called it “Chicleros Ulcer” because it was frequently found in chicle workers. Nowadays CL is becoming important because of the number of cases appearing per year, some of such cases being DCL. CL in Mexico has been found mainly in the forests of Campeche, Quintana Roo, Tabasco, Chiapas, Oaxaca and Veracruz states, it has also been found in semi-arid regions of the country (Velasco et al. 1989). Nevertheless, incidence and prevalence rates in those places are unknown because CL became a notifiable disease in Mexico only at the end of 1986 and proper epidemiological studies have not been carried out on CL in other states apart from Campeche. “Chicleros ulcer” was recorded in Campeche state by several authors at different times, Seidelin (1912), Beltrán and Bustamante (1942), Biagi (1953a) and Andrade-Narvaez et al. (1990).

The most reliable methods to confirm the diagnosis of leishmaniasis are the identification of the parasite in thin smears made from superficial scrapings of the ulcers and isolation of the parasite by inoculation of culture media or susceptible animals (Navin et al. 1990), but CL parasitological diagnosis is difficult in a patient with more than one year of evolution and furthermore in the field and small towns or rural areas there are no facilities to grow the parasite in culture or in animals. For these reasons it is necessary to find methods for an accurate diagnosis of CL but especially those which can be used in small laboratories in the endemic areas.

It has been shown by Andrade-Narvaez et al. (1990) and Souza et al. (1992), that the Montenegro skin test (MST) is very useful to identify or characterize areas in which there is a risk of CL infec-
tion. MST has a very good sensitivity, but patients who have been cured remain giving positive skin reactions. Serological techniques together with the skin test could be useful to identify accurately patients with CL (Garcia-Miss et al. 1990, Guimarães et al. 1990).

The aim of this work was to determine the reactivity to MST of individuals living in the endemic district of Holpechen, Campeche as well as to measure by indirect immunofluorescent antibody test (IFA) their specific antibodies to antigens of two strains of *L. mexicana* isolated in Mexico. We also searched by Western blot analysis which antigens of this parasite are recognized by patients with LCL of the same area to try to find a more dependable method of diagnosis of CL.

**MATERIALS AND METHODS**

**Study population** - The state of Campeche has an area of 56,850 km². Tropical humid, high forests predominate, covering about 60% of the state. Campeche state has a population of 533,703, around 45% of which work in the forest (Andrade-Narvaez et al. 1990). People from rural areas (169 persons) of the Holpechen district accepted to participate in the study (101 males and 68 females). The following data was obtained from all participants: age, sex, occupation and places where they had lived or traveled. MST was performed on the individuals and sera were obtained for serological analysis.

In the first part of the study individuals with ulcerative skin lesions were considered as suggestive cases of leishmaniasis. Parasitological diagnosis of cutaneous leishmaniasis was attempted in individuals with suggestive lesions by inoculation of aspirate into biphasic Senekji’s and RPMI media and microscopic search of amastigotes in Giemsa stained thin smears made from superficial scrapings of the ulcers. All patients were treated with intraleisonal administration of glucantime. A confirmed case of cutaneous leishmaniasis was considered if a positive *Leishmania* culture was observed or if the parasite was found in the thin smears. The other patients were considered leishmaniasic because, although the parasite could not be demonstrated, they were MST positive and responded to the treatment. Two patients in whom *Leishmania* could not be found, were MST negative and did not respond to the treatment were later diagnosed as skin cancer by histopathological studies. The lesion evolution time in the patients was from 4 months to 8 years.

**Strains of Leishmania** - Two *L. mexicana* strains, a *L. m. mexicana* MHOM/MX/88/HRC 1S isolated from a patient with LCL and one *L. m. mexicana* MHOM/MX/84/SET GS from a patient with DCL were used to prepare antigens. Both patients came from the Tabasco state of Mexico, very close to Campeche.

**Sera** - Two hundred and eighteen sera were analyzed for specific antibodies. 119 sera were obtained from individuals living in the endemic area of Holpechen district (healthy individuals from endemic area). 48 were from patients with leishmaniasis (described above); 31 were from patients from Mexican states other than Campeche and with diagnosis of Chagas’ disease and 20 came from healthy people that have lived in Mexico city (non-endemic area), who have never been in endemic areas, were MST negative and had no lesions suggestive of cutaneous leishmaniasis (healthy individuals from non-endemic area).

**Preparation of antigens** - *Leishmania* parasites were grown in RPMI 1640 medium (Sigma Chem, St Louis, Mo, USA) supplemented with 10% fetal calf serum. Leishmanin was prepared with promastigotes of 9-10 days of culture which were harvested by centrifugation (at 1500 g for 5 min at 4°C) and washed three times in phosphate buffered saline (PBS) pH 7.4. The pellet was resuspended in 0.05% phenol 0.85% NaCl at a final concentration of 5×10⁶ parasites/ml for skin test (Bray 1980), or in an antiproteolytic electrophoresis sample buffer containing 0.04M NaCl, 10mM PMSF, 10mM TLCK and Tris pH 8.0 for Western blot analysis.

**Skin test** - 0.1 ml of Montenegro’s antigen (leishmanin) were injected intradermally in the volar surface of the forearm. The reaction was observed at 24, 48 and 72 hr. A positive reaction was considered when an induration of 5 mm or more was observed.

**Antibody test** - The IFA test was used to measure anti *L. mexicana* antibodies (IgG). Promastigotes of 9-10 days of culture in RPMI medium were used as antigen. They were washed twice with PBS, 8×10⁴ promastigotes were spread as a thin “dot smear” on slides and allowed to dry thoroughly and fixed with absolute methanol. The assay was performed as described by Bray (1980), using as a starting point a 1:4 serum dilution. Fluorescent labeled goat anti human IgG (Cappel laboratories, USA), was used to develop the reaction.

**Antigen recognition by Western blot** - Antibodies against different antigens of *Leishmania* were determined by Western blot analysis as described by Towbin et al. (1979) with some modifications: promastigotes of both LCL and DCL strains of *Leishmania* were used for the analysis. Electrophoresis in 10% SDS-PAGE 10 cm gels was carried out with 3×10⁸ promastigotes from one or the other strain of *L. mexicana*. The gels were run for 40 min at 30 mA and electrophoretically transferred to nitrocellulose membranes (Amersham, 0.45 pm pore-size) overnight at 100 mA. After blocking overnight with bovine serum albumin (Sigma
Chem., USA) the membranes were incubated with patient’s sera, diluted 1:20, sera from healthy individuals living in endemic areas or healthy individuals from non-endemic areas. To develop the reaction peroxidase-labeled goat antihuman IgG (Cappel laboratories, USA) was used.

RESULTS

Parasitological studies - Lesions suggestive of CL were found in 50 of the 169 individuals examined from the endemic area. 86% of them were male and 14% female and the most affected age was 11 to 30 years of age (Table I). Lesions occurred mainly on the ear (39%) followed by the forearm (18%) and on the face (11%). In 70% of the patients the parasite was either isolated or observed by direct microscopy.

Skin test - Skin test was applied to a total of 191 individuals, 169 from endemic area and 22 from non-endemic area. The strongest reaction was observed at 48 hr with around 1 cm reactions being common. 16% of healthy people, and 96% of patients from the endemic area gave positive skin tests. This gives a 40% overall positivity in this endemic area. Healthy individuals with no lesions from non-endemic areas gave negative skin tests. Two patients from the endemic area showing lesions similar to cutaneous leishmaniasis and who were later diagnosed as skin cancer (see materials and methods), were also skin tested and gave a negative result (Table II).

Serum antibodies by IFA - When the LCL strain of *L. mexicana* was used as antigen, all the 20 sera from donors living in non-endemic areas (Mexico city) gave negative results while only 3 from patients with leishmaniasis gave negative results. The other 45 from patients with leishmaniasis gave positive tests between 1:16 and 1:1024 dilutions. Most of the individuals from the endemic area gave negative tests (112) and 7 were positive between 1:4 and 1:32 dilutions. Only two patients with Chagas’ disease diagnosis gave a positive test, both at 1:32 dilution while the rest (29) were negative (Fig. 1). One of the patients diagnosed as skin cancer gave a negative test and the other gave a 1:16 titer. Neither of them were included in Fig. 1. Table III shows the positivity results of the different groups using as cut-off point a 1:32 dilution which gives a 100% specificity for the assay. It is clear that the results using two different strains of *Leishmania* (DCL and LCL) are very similar, although LCL gave a positive result with two more patients. The patients with leishmaniasis gave around 70% positivity which gives an overall positivity of 20% for the population from the endemic area (Table III). When the antibody titers obtained with one or the other *Leishmania* strain are compared, there seems to be a tendency to obtain higher titers with LCL (Fig. 2).

Western blot analysis - All the 48 sera from patients with leishmaniasis plus 19 from healthy donors living in non-endemic areas were later diagnosed as skin cancer (see materials and methods), were also skin tested and gave a negative result (Table II).

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of subjects</th>
<th>No. positive (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Healthy individuals from EA</td>
<td>119</td>
<td>19 (16)</td>
</tr>
<tr>
<td>Patients with skin lesions</td>
<td>50</td>
<td>48 (96)</td>
</tr>
<tr>
<td>Total from EA</td>
<td>169</td>
<td>67 (40)</td>
</tr>
<tr>
<td>Healthy individuals from NEA</td>
<td>22</td>
<td>0 (0)</td>
</tr>
</tbody>
</table>

EA: endemic area
NEA: non endemic area

![Fig. 1: antibody titer distribution of the different groups of sera tested using localized cutaneous leishmaniasis antigen. The numbers at the bottom of the columns indicate the number of negative sera for each group at 1:4 dilution.](image)
TABLE III

Immunofluorescence test results of the different groups of sera tested

<table>
<thead>
<tr>
<th>Groups</th>
<th>No. of sera</th>
<th>DCL(%)</th>
<th>LCL(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Healthy individuals from EA</td>
<td>119</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Patients with leishmaniasis</td>
<td>48</td>
<td>32 (67)</td>
<td>34 (71)</td>
</tr>
<tr>
<td>Total from EA</td>
<td>169</td>
<td>32 (19)</td>
<td>34 (20)</td>
</tr>
<tr>
<td>Healthy individuals from NFA</td>
<td>20</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Patients with Chagas’ disease</td>
<td>31</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

EA: endemic area, NFA: non endemic area, DCL: diffuse cutaneous leishmaniasis; LCL: localized cutaneous leishmaniasis

![Antibody titre distribution](image)

Fig. 2: antibody titre distribution, expressed as percentage of the total of patients with leishmaniasis (48) when assayed with two different Leishmania strains. Open bars are results with diffuse cutaneous leishmaniasis (DCL) and hatched bars results with localized cutaneous leishmaniasis (LCL).

![Percentage of recognition](image)

Fig. 3: percentage of recognition of different Leishmania antigens by Western blot by sera of 48 patients with leishmaniasis. Open bars are results with diffuse cutaneous leishmaniasis (DCL) and hatched bars results with localized cutaneous leishmaniasis (LCL).

In the present study carried out in the Holpachen district of Campeche state a 40% overall positivity to MST was found (Table II). In patients with suggestive lesions MST became positive in early stages of the infection as patients with a month of evolution gave skin reactions greater than 5 mm; there was no relationship between the time of evolution and the size of the skin test (data not shown). These results confirm those of Biagi (1953b), who found 36 out of 84 (43%) field workers with positive MST reactions in Escárcega district of Campeche, Andrade-Narváez et al. (1985) who found 66 out of 330 (20%) members of the Mexican army with positive MST in the Peninsula of Yucatán (near Campeche) and Andrade-Narváez et al. (1990), who found in several rural areas of Campeche state a 43% MST positivity. It seems that the incidence of CL has been maintained at the same level at different times in different endemic areas of Campeche.

As Biagi (1953a) and Andrade-Narváez et al. (1990) reported, we found also a correlation among age, sex, occupation and positive skin lesions. Table I shows that the range of age more affected was from 11 to 30 years and that the most affected group was 21-30 year-olds. 86% of the 50 patients were male and 14% female (compared to 60% males and 40% females in the population studied from the endemic area), and agriculture was their
main occupation. It seems that the risk of infection is due to the agricultural occupation, because the women and the 3 children under 10 years (2 males and 1 female) who were infected (Table I), regularly went to work in the field. There appears to be no urban transmission of leishmaniasis in the area studied.

In serological studies Garcia-Miss et al. (1990) found 76% of patients with “Chicleros Ulcer” with positive antibodies by ELISA in some rural areas different from Holpecenh and report that their assay has a 76% sensitivity and 100% specificity. In the present study (Fig. 1, Table III), 71% of positivity by IFA test was found in our patients with leishmaniasis which results in 20% positivity for the general population of Holpecenh using a cut off point of 1:32. Using the healthy population from the endemic area to evaluate the performance of our assay, we have a 70.8% sensitivity and 100% specificity.

We were also interested in finding out if the stain of Leishmania used for antibody determinations influences the results. Fig. 2 shows the titers obtained in the same patients using both strains of Leishmania (LCL and DCL) and we can see that there is not much difference between the reactivity against both strains of Leishmania, although with the LCL strain the IFA titers were slightly higher, resulting in only 4% more positives among leishmaniasis cases (Table III).

Our results show that antibody determination alone is not of much help for the diagnosis of leishmaniasis as around 30% of the patients gave titers similar to those of healthy people in the endemic area and three patients were even negative (Fig. 1). A combination of antibody determination and skin test seems to be more valuable as an aid for the diagnosis of leishmaniasis in this endemic area. As mentioned above, two patients who had skin lesions were MST negative and were later diagnosed as having skin cancer. One of these patients was negative by IFA and the other had a titer of 1:16 (data not shown).

Excluding these two patients, who obviously are not leishmaniasic, the performance of the MST in this endemic area is a 100% sensitivity and 84% specificity with a 100% negative predictive value. The usefulness of such a high negative predictive value is illustrated by the two cases of skin cancer unexpectedly found in our studied population and who were MST negative.

On the other hand, if the cut-off point for the IFA assay is set to 1:16, the sensitivity is increased importantly to 85.4% while the specificity is lowered only slightly (to 97.4%).

We can conclude that for the endemic area studied, the presence of an ulcerative skin lesion, an antibody titer by IFA above 1:16 and a positive Montenegro skin test can be considered as a positive diagnosis of leishmaniasis even if the parasite cannot be isolated or demonstrated.

Our results and those of Garcia-Miss et al. (1990) clearly indicate that the serological assays for leishmaniasis must be improved, specially in their sensitivity. As a first step toward this goal, we analyzed the reactivity of our serum samples by Western blot. As Fig. 3 shows, there was an important difference in antigen recognition of the sera of the patients with CL between both strains of L. mexicana. In spite of the fact that the LCL strain was slightly better for detection of Leishmania antibodies than DCL by the IFA test, by Western blot the patients recognized more antigens in DCL and some of them with higher frequency than in LCL. Another important feature is that in both strains of Leishmania few patients recognized antigens in a range of 40-70 kDa where GP63, one of the important antigens of Leishmania, is found (Moody 1993). This antigen has been considered as a candidate for a Leishmania vaccine. On the other hand 78% of patients recognized a 105 kDa antigen in the DCL strain. In this range of Mw the secreted acid phosphatase (SAP) would be found; this enzyme is thought to contribute to parasite survival in the host, as it is involved in pathogenicity and parasite nutrition (Hassan & Coombs 1987, Ilg et al. 1991). In the LCL strain 40% of the patients recognized a 205 kDa antigen, but we do not know the significance of this, because there is no information on important antigens of this molecular weight in L. mexicana.

The Western blot analysis indicates that it might be possible to develop better serological assays for epidemiology and diagnosis. The 105 kDa antigen was recognized by almost 80% of patients and by none of the 19 sera from healthy persons from endemic area which included some sera with IFA titers of 1:32 and below. Other antigens were also recognized—by 20-30% of the patients (Fig. 3) so we think that it would be possible to find specific antigens of Leishmania which could be used to improve the serological assays for the diagnosis and seroepidemiology of leishmaniasis.

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REFERENCES


